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Studies on Myxosporidia

Richard Roksabro Kudo



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Nos. 3 and 4



STUDIES ON MYXOSPORIDIA

A SYNOPSIS OF GENERA AND SPECIES
OF MYXOSPORIDIA

WITH 25 PLATES AND 2 TEXTFIGURES

ROKUSABURO KUDO

Price \$3.00

PUBLISHED BY THE UNIVERSITY OF ILLINOIS UNDER THE AUSPICES OF THE GRADUATE SCHOOL URBANA, ILLINOIS

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STUDIES ON MYXOSPORIDIA

A SYNOPSIS OF GENERA AND SPECIES OF MYXOSPORIDIA

WITH 25 PLATES AND 2 TEXTFIGURES

BY ROKUSABURO KUDO

Contributions from the Zoological Laboratory of the University of Illinois No. 158



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INTRODUCTION

Ten years have elapsed since Auerbach (1910) published *Die Cnidosporidien* in which he gave a synopsis of the genera and species of Myxosporidia known up to that time. During this period new genera and a number of new species have been added to the list of this particular group of parasitic protozoa from the various parts of the world. It is, therefore, desirable to have a complete monographic work including all the forms reported up to the present time.

The main objects of the present paper are: 1) to describe a new genus and a number of new species which have come under the observation of the writer; 2) to collect all the genera and species recorded by various authors; 3) to propose a new classification by which some of the confusion now existing may, probably, be avoided; 4) to show the geographical, zoological and organal distribution in the light of more recent observations; and 5) to present a complete list of the names of the hosts in which Myxosporidia occur.

The writer believes himself to be in possession of as complete references as possible under present conditions. However, he may be unaware of some works which have not reached him owing to the war.

The Myxosporidia recorded by Labbé (1899) are arranged in almost the same order as that author listed them, with some slight change such as placing the type species at the front of each genus or removing a few species to other genera, while those species which have been described since 1898 are arranged chronologically, no matter whether names are given the species or not.

Some of the references are omitted, especially when they can be found in Gurley (1894), Thélohan (1895), Labbé (1899), or Auerbach (1910). The description of each species is given according to the first observer. The observations of subsequent investigators are then mentioned in the second place.

Each species is described according to the following scheme:

- 1) Specific name
- 2) Synonyms and literature
- 3) Habitat, including the locality and the date of observation
- 4) Vegetative form
- 5) Spore
- 6) Remarks

I wish to express my appreciation to Professor Henry B. Ward whose kindness has made the completion of this paper possible.



GENERAL REMARKS ON RECENT OBSERVATIONS

The total number of species of Myxosporidia reported up to date and described in the following pages, excluding 12 ambiguous forms, reaches 237 of which 125 are species which have been observed since 1910.*

The distribution of these new forms is as follows:

Africa:	6	species
Asia		
Australia	1	species
Europe	31	species
North America		
South America	1	species

Thus, the majority of the species were observed in other lands than Europe, nearly half being recorded from North American waters. It is not hard to anticipate from the observations made by Awerinzew, Davis, Kudo, Mavor, Johnston and Bancroft, and others, that further investigations on the parasites in the localities where the study of the protozoa under consideration was neglected, will bring out not only new and interesting forms which will be quite different from the comparatively well studied European species, but also many important facts that will clear unknown or doubtful phases concerning the life history and structure of Myxosporidia.



^{*} Three species are included here which have been described (in Nipponese) by Miyairi in 1909.

MYXOSPORIDIA RECORDED IN THE PRESENT PAPER LIST I

Order MYXOSPORIDIA Bütschli I Suborder EURYSPOREA nom. nov. (see page 56) I Family CERATOMYXIDAE Doflein

Genus 1 LEPTOTHECA Thélohan [15 species]

- 1) L. agilis Thélohan (type species)
- 2) L. elongata Thélohan
- 3) L. polymorpha (Thél.) Labbé
- 4) L. parva Thélohan
- 5) L. renicola Thélohan
- 6) L. hepseti Thélohan
- 7) L. perlata (Gurley) Labbé
- 8) L. sp. Awerinzew
- 9) L. macrospora Auerbach
- 10) L. informis Auerbach
- 11) L. longipes Auerbach
- 12) L. fusiformis Davis
- 13) L. scissura Davis
- 14) L. lobosa Davis
- 15) L. glomerosa Davis

Genus 2 CERATOMYXA Thélohan [35 species]

- 1) C. arcuata Thélohan (type species)
- 2) C. sphaerulosa Thélohan
- 3) C. pallida Thélohan
- 4) C. globurifera Thélohan
- 5) C. appendiculata Thélohan
- 6) C. truncata Thélohan
- 7) C. reticularis Thélohan
- 8) C. inaequalis Doflein
- 9) C. linospora Doflein
- 10) C. ramosa Awerinzew
- 11) C. drepanopsettae Awerinzew
- 12) C. tylosuri Awerinzew
- 13) C. (?) spari Awerinzew
 14) C. sp. (?) Awerinzew
- 15) C. sp. (?) Awerinzew
- 16) C. acadiensis Mayor

- 17) C. sp. Georgévitch
- 18) C. coris Georgévitch
- 19) C. herouardi Georgévitch
- 20) C. mesospora Davis
- 21) C. sphairophora Davis
- 22) C. taenia Davis
- 23) C. attenuata Davis
- 24) C. recurvata Davis
- 25) C. lunata Davis
- 26) C. abbreviata Davis
- 27) C. flagellifera Davis
- 28) C. agglomerata Davis
- 29) C. amorpha Davis
- 30) C. monospora Davis
 31) C. streptospora Davis
- 32) C. aggregata Davis
- 33) C. undulata Davis
- 34) C. navicularia Davis
- 35) C. spinosa Davis

Genus 3 MYXOPROTEUS Doflein

[3 species]

- 1) M. ambiguus (Thélohan) Doflein (type species)
- 2) M. cordiformis Davis
- 3) M. cornulus Davis

Genus 4 WARDIA nov. gen.

[2 species]

- 1) W. ovinocua nov. spec. (type species)
- 2) W. ohlmacheri (Gurley) Kudo

Genus 5 MITRASPORA Fujita emend. Kudo [3 species]

- 1) M. cyprini Fujita (type species)
- 2) M. caudata (Parisi) Kudo
- 3) M. elongata nov. spec.



II Suborder SPHAEROSPOREA nom. nov. (see page 57) I Family CHLOROMYXIDAE Thélohan

Genus 1 CHLOROMYXUM Mingazzini

[22 species]

- 1) C. leydigi Mingazzini (type species)
- 2) C. caudatum Thélohan
- 3) C. quadratum Thélohan
- 4) C. fluviatile Thélohan
- 5) C. mucronatum Gurley
- 6) C. diploxys (Gurley) Thélohan
- 7) C. protei Joseph
- 8) C. truttae Léger
- 9) C. cristatum Léger
- 10) C. dubium Auerbach

- 11) C. sp. Awerinsew
- 12) C. thymalli Lebzelter
- 13) C. koi Fujita
- 14) C. magnum Awerinsew
- 15) C. funduli Hahn
- 16) C. misgurni Kudo
- 17) C. fujitai Kudo
- 18) C. clupeidae Hahn
- 19) C. granulosum Davis
- 20) C. trijugum nov. spec.
- 21) C. catostomi nov. spec.
- 22) C. wardi nov. spec.

II Family SPHAEROSPORIDAE Davis

Genus 1 SPHAEROSPORA Thélohan

[10 species]

- 1) S. divergens Thélohan (type species)
- 2) S. elegans Thélohan
- 3) S. rostrata Thélohan
- 4) S. masovica Cohn
- 5) S. platessae Woodcock
- 6) S. angulata Fujita
- 7) S. sp. Davis
- 8) S. polymorpha Davis

- 9) S. (?) sp. Southwell et Prashad
- 10) S. carassii nov. spec.

Genus 2 SINUOLINEA Davis

[5 species]

- 1) S. dimorpha Davis (type species)
- 2) S. capsularis Davis
- 3) S. arborescens Davis
- 4) S. opacita Davis
- 5) S. brachiophora Davis

III Suborder PLATYSPOREA nom. nov. (see page 57) I Family MYXIDIDAE Thélohan

Genus 1 MYXIDIUM Bütschli

[26 species]

- 1) M. lieberkühni Bütschli (type species)
- 2) M. incurvatum Thélohan
- 3) M. sphaericum Thélohan
- 4) M. kistophilum Thélohan
- 5) M. sp. Gurley
- 6) M. danilewskyi Laveran
- 7) M. giganteum Doflein
- 8) M. barbatulae Cépède
- 9) M. giardi Cépède
- 10) M. pfeifferi Auerbach
- 11) M. inflatum Auerbach
- 12) M. bergense Auerbach
- 13) M. procerum Auerbach
- 14) M. mackiei Bosanquet
- 15) M. macrocapsulare Auerbach16) M. sp. Awerinzew
- 17) M. depressum Parisi
- 18) M. oviforme Parisi 19) M. anguillae Ishii
- 20) M. sp. Mavor

- 21) M. gadi Georgévitch
- 22) M. glutinosum Davis
- 23) M. phyllium Davis
- 24) M. striatum Cunha et Fonseca
- 25) M. kagayamai nov. spec.
- 26) M. americanum nov. spec.

Genus 2 SPHAEROMYXA Thélohan

[7 species]

- 1) S. balbianii Thélohan (type species)
- 2) S. immersa (Lutz) Thélohan
- 3) S. incurvata Doflein
- 4) S. sabrasesi Laveran et Mesnil
- 5) S. hellandi Auerbach
- 6) S. exneri Awerinzew
- 7) S. gasterostei Georgévitch

Genus 3 ZSCHOKKELLA Auerbach [4 species]

- 1) Z. hildae Auerbach (type species)
- 2) Z. nova Klokacewa
- 3) Z. acheilognathi Kudo
- 4) Z. globulosa Davis

II Family MYXOSOMATIDAE Poche

Genus 1 MYXOSOMA Thélohan [3 species]

- 1) M. dujardini Thélohan (type species)
- 2) M. (?) lobatum Nemeczek
- 3) M. funduli Kudo

Genus 2 LENTOSPORA Plehn [6 species]

- 1) L. cerebralis (Hofer) Plehn (type species)
- 2) L. multiplicata Reuss
- 3) L. encephalina Mulsow
- 4) L. asymmetrica Parisi
- 5) L. acuta (Fujita) Kudo
- 6) L. dermatobia Ishii

III Family MYXOBOLIDAE Thélohan

Genus 1 MYXOBOLUS Bütschli

- [63 species] 1) M. mülleri Bütschli (type species)
- 2) M. piriformis Thélohan
- 3) M. unicapsulatus Gurley
- 4) M. fuhrmanni Auerbach
- 5) M. oculi-leucisci Trojan
- 6) M. toyamai Kudo
- 7) M. notatus Mavor
- 8) M. sp. Kudo
- 9) M. rokitae Southwell et Prashad
- 10) M. seni Southwell et Prashad
- 11) M. misgurni nov. spec.
- 12) M. pfeifferi Thélohan
- 13) M. inaequalis Gurley
- 14) M. dispar Thélohan
- 15) M. ellipsoides Thélohan
- 16) M. exiguus Thélohan
- 17) M. oviformis Thélohan
- 18) M. lintoni Gurley
- 19) M. globosus Gurley 20) M. oblongus Gurley
- 21) M. transovalis Gurley 22) M. obesus Gurley
- 23) M. cycloides Gurley
- 24) M. sphaeralis Gurley
- 25) M. anurus Cohn
- 26) M. sp. Gurley
- 27) M. sp. Gurley
- 28) M. sp. Gurley
- 29) M. cyprini Doflein
- 30) M. neurobius Schuberg et Schröder
- 31) M. aeglefini Auerbach
- 32) M. gigas Auerbach
- 33) M. volgensis Reuss
- 34) M. scardinii Reuss
- 35) M. physophilus Reuss
- 36) M. macrocapsularis Reuss
- 37) M. sandrae Reuss
- 38) M. bramae Reuss

- 39) M. cyprinicola Reuss
- 40) M. balleri Reuss
- 41) M. squamae Keysselitz
- 42) M. cordis Keysselitz
- 43) M. musculi Keysselitz
- 44) M. sp. Miyairi
- 45) M. sp. Wegener
- 46) M. permagnus Wegener
- 47) M. rotundus Nemeczek
- 48) M. minutus Nemeczek
- 49) M. sp. Lebzelter
- 50) M. magnus Awerinzew
- 51) M. carassii Klokacewa
- 52) M. sp. Southwell
- 53) M. funduli Kudo
- 54) M. pleuronectidae (Hahn)
- 55) M. capsulatus Davis
- 56) M. nodularis Southwell et Prashad
- 57) M. kylae Johnston et Bancroft
- 58) M. aureatus Ward
- 59) M. miyairii nov. spec.
- 60) M. koi nov. spec.
- 61) M. orbiculatus nov. spec.
- 62) M. discrepans nov. spec.
- 63) M. mesentericus nov. spec.

Genus 2 HENNEGUYA Thélohan [32 species]

- 1) H. psorospermica Thélohan (type species)
- 2) H. texta (Cohn) Labbé
- 3) H. minuta (Cohn) Labbé
- 4) H. oviperda (Cohn) Labbé
- 5) H. lobosa (Cohn) Labbé
- 6) H. peri-intestinalis Cépède
- 7) H. media Thélohan
- 8) H. brevis Thélohan
- 9) H. schizura (Gurley) Labbé
- 10) H. creplini (Gurley) Labbé

- 11) H. linearis (Gurley) Labbé
- 12) H. gurleyi Kudo
- 13) H. strongylura (Gurley) Labbé
- 14) H. monura (Gurley) Labbé
- 15) H. kolesnikovi (Gurley) Labbé
- 16) H. macrura (Gurley) Thélohan
- 17) H. sschokkei (Gurley) Doflein
- 18) H. sp. (Gurley) Labbé
- 19) H. sp. (Gurley) Labbé
- 20) H. tenuis Vaney et Conte
- 21) H. müsslini Schuberg et Schröder
- 22) H. legeri Cépède
- 23) H. acerinae Schröder
- 24) H. gigantea Nemeczek
- 25) H. (?) sp. Nemeczek
- 26) H. gasterostei Parisi
- 27) H. neapolitana Parisi
- 28) H. wisconsinensis Mavor
- 29) H. brachyura Ward
- 30) H. salminicola Ward

- 31) H. miyairii nov. spec.
- 32) H. mictospora nov. spec.

Genus 3 HOFERELLUS Berg [1 species]

1) H. cyprini Doslein

Appendix: Myxosporidia of unknown genera and species [11 forms]

- 1) Gen. et spec. incert. Leydig
- 2) Gen. et spec. incert. Leydig
- 3) Gen. et spec. incert. Leydig
- 4) Gen. et spec. incert. Heckel et Kner
- 5) Gen. et spec. incert. Borne
- 6) Gen. incert. merlucii Perugia
- 7) Gen. incert. congri Perugia
- 8) Gen. et spec. incert. Linton
- 9) Gen. et spec. incert. Mingassini
- 10) Gen. et spec. incert. Nufer
- 11) Gen. et spec. incert. Mavor
- 12) Gen. et spec. incert. Mavor

DISTRIBUTION OF MYXOSPORIDIA

A. GEOGRAPHICAL DISTRIBUTION

As will be seen from List III, Myxosporidia are common parasites of fish in various parts of the world.

It is interesting to notice that the same species are found among freshwater or marine fish from waters in widely separated countries. It is possible to think that Myxosporidia in marine fish may be carried into remote waters by the migration of their hosts, while those infecting freshwater fish may be brought from one place to another by the transportation of infected fish for breeding purpose, etc. It should be noted in this connection that no intermediate host has yet been found in relation to myxosporidiosis.

The followings are the common species found in different localities:

Leptotheca parva Thél.
Ceratomyza sphaerulosa Thél.
C. appendiculata Thél.
C. drepanopsettae Awerinzew
Chloromyzum leydigi Ming.
C. quadratum Thél.
Sphaerospora elegans Thél.
S. divergens Thél.
Myzidium lieberkühni Bütsch.

M. incurvatum Thél.
M. bergense Auerbach.
M. oviforme Parisi
S phaeromyxa balbianii Thél.
M yzosoma dujardini Thél.

Marseille, Bergen
Monaco, Roscoff, Bergen
Roscoff, Marseille, Rovigno
Murman coast, Bergen, Woods Hole
Roscoff, Monaco, Napoli, Rovigno, Beaufort
Roscoff, Marseille, Napoli, Beira
Bretagne, Karlsruhe, Lago di Garda
Napoli, Roscoff, Smalfjorden
Lago Maggiore, France, Germany, Lake
Mendota, Georgian Bay
Napoli, Monaco, Roscoff, Bergen, Beaufort
Bergen, St. Andrews
Napoli, Norwegian coast
Roscoff, Napoli, Beaufort
France, Germany, Tokio(?)

On the other hand, some species are limited to certain localities. Five species classified in the genus Sinuolinea by Davis are reported only from Beaufort, N. C., U. S. A. The two species of the genus Wardia have been found solely in the state of Illinois, U. S. A.

More detailed data are shown in the following list.

LIST II

ASIA

I NIPPON

Myxosporidia of fresh water fish

1) Northern Part (Hokkaido)

Sapporo: Mitraspora cyprini Fujita
Chloromyxum koi Fujita
Sphaerospora angulata Fujita
Lentospora acuta (Fujita) Kudo

2) Central part (Hondo)

Tokio: Mitraspora cyprini Fujita
Chloromyxum misgurni Kudo
Chloromyxum fujitai Kudo
Sphaerospora carassii nov. spec.
Myxidium hagayamai nov. spec.
Zschokkella acheilognathi Kudo
Myxosoma dujardini (?) Thélohan
Myxobolus toyamai Kudo
Myxobolus misgurni, nov. spec.

Myzobolus koi nov. spec.

Numazu: Myxidium anguillae Ishii Lentospora dermatobia Ishii

Southern part (Kiushiu)
 Fukuoka: Myzobolus sp. Miyairi
 Myzobolus miyairii nov. spec.
 Henneguya miyairii nov. spec.

II India

A. Myxosporidia of fresh-water fish

Katwan, Mirzapore (U.P.): Myzobolus sp. Southwell

Mirpur, Decca district: Myzobolus rokitae Southwell et Prashad

Myzobolus seni Southwell et Prashad Myzobolus nodularis Southwell et Prashad

B. Myxosporidian of reptiles Myxidium mackiei Bosanquet

Bombay:

III BURMA

In the vicinity of Ruby Mines: Sphaerospora sp. Southwell et Prashad

IV KAMTSCHATKA

?Henneguya salminicola Ward

AUSTRALIA

Myxosporidian of amphibia
In the vicinity of Sidney: Myxobolus kylae Johnston et Bancroft

AFRICA

Nile:

A. Myxosporidia of fresh-water fish Myxobolus unicapsulatus Gurley Henneguya strongylura Gurley
B. Myxosporidia of marine fish

1) Indian Ocean

Beira:

Algoa Bay:

Chloromyxum magnum Awerinzew Chloromyxum quadratum Thélohan

East London: Chloromyxum magnum Awerinzew Lorenço Marques: Ceratomyxa tylosuri Awerinzew

Ceratomyza spari Awerinzew
Ceratomyza sp(?). Awerinzew
Ceratomyza sp (?). Awerinzew
Sphaeromyza exneri Awerinzew

2) South Atlantic Ocean

Lüderitz Bay: Chloromyxum magnum Awerinzew

NORTH AMERICA

I United States

A. Myxosporidia of fresh-water fish

1) From Rivers emptying into Atlantic Ocean

Carlius, Va. (tribt. of Potomac River): Myxobolus transovalis Gurley Myxobolus globosus Gurley

Columbia, S. C. (Santee River):

Myxobolus globosus Gurley

Kinston, N. C. (Neuse River): West Falmouth, Mass.:

Myxobolus sp. Kudo

Woodbury, N. J. (Delaware River):

Henneguya monura Gurley

2) From Lakes and Rivers opening into the Gulf of Mexico Fox River, trib. Mississippi: Myzobolus globosus Gurley

Lake Mendota, Wis.:

Myxidium lieberkühni Bütschli

Henneguya wisconsinensis Mavor et Strasser

Neches River, Palestin, Tex.: Henneguya macrura (Gurley) Thélohan

Storm Lake, Ia .:

Henneguya gurleyi Kudo

Stony Creek, Ill.:

. Chloromyxum trijugum nov. spec.

Myzobolus orbiculatus nov. spec.

Henneguya mictos pora nov. spec.

Homer Park, Ill.:

Chloromyxum trijugum nov. spec.

Myxobolus orbiculatus nov. spec.

Salt Fork, Urbana, Ill.:

Wardia ovinocua nov. gen. nov. spec. Chloromyxum catostomi nov. spec.

Myxobolus discrepans, nov. spec.

Crystal Lake, Urbana, Ill.:

Mitraspora elongata nov. spec.

Myxidium americanum nov. spec.

Myxobolus mesentericus nov. spec.

3) From the rivers opening into the Great Lakes

Black River, Ohio: Gen. et spec. incert. Linton

Put-In-Bay, Ohio:

Myxobolus aureatus Ward

Henneguya brachyura Ward

B. Myxosporidia of marine fish (Atlantic Ocean)

Leptotheca fusiformis Davis Beaufort, N. C.:

Leptotheca scissura Davis

Leptotheca lobosa Davis

Leptotheca glomerosa Davis

Ceratomyxa mesospora Davis

Ceratomyxa sphairophora Davis

Ceratomyxa taenia Davis

Ceratomyxa attenuata Davis

Ceratomyxa recurvata Davis

Ceratomyxa lunata Davis

Ceratomyxa abbreviata Davis Ceratomyxa flagellifera Davis

Ceratomyxa agglomerata Davis

Ceratomyxa amorpha Davis

Ceratomyxa monospora Davis

Ceratomyxa streptospora Davis

Ceratomyxa aggregata Davis Ceratomyxa undulata Davis

Ceratomyza navicularia Davis Ceratomyxa spinosa Davis Myzoproteus cordiformis Davis Myxoproteus cornutus Davis Chloromyxum leydigi Mingazzini Chloromyxum granulosum Davis Sphaerospora polymorpha Davis Sinuolinea dimorpha Davis Sinuolinea capsularis Davis Sinuolinea arborescens Davis Sinuolinea opacita Davis Sinuolinea brachiophora Davis Myxidium incurvatum Thélohan Myxidium glutinosum Davis Myxidium phyllium Davis Sphaeromyza balbianii Thélohan Zschokkella globulosa Davis Myxobolus capsulatus Davis

Woods Hole, Mass: Ceratomyxa drepanopsettae Awerinzew

Chloromyxum funduli Hahn
Chloromyxum clupeidae Hahn
Myzosoma funduli Kudo
Myzobolus lintoni Gurley
Myzobolus funduli Kudo
Myzobolus pleuronectidae Hahn

Locality unrecorded: Henneguya schisura (Gurley) Labbé

C. Myxosporidian of Amphibia

Sycamore, Ill.: Wardia ohlmacheri (Gurley) Kudo

II CANADA

A. Myxosporidia of fresh-water fish Georgian Bay (south. part): Myxidium lieberkühni Bütschli Myxobolus notalus Mavor Gen. et spec. incert. Mavor

B. Myxosporidia of marine fish (Atlantic Ocean)

Passamaquoddy Bay (at or near the mouth of St. Croix

River), New Brunswick: Ceratomyza acadiensis Mavor

Myxidium bergense Auerbach

M. sp. Mavor

Gen. et spec. incert. Mavor

III Alaska

Klutina Lake: Chloromyzum wardi nov. spec. Stickeen River: Henneguya salminicola Ward

SOUTH AMERICA

A. Myxosporidia of fresh-water fish from the waters connected with Atlantic Ocean

Guiana: Myxobolus inaequalis Gurley Surinam: Myxobolus inaequalis Gurley Locality?: Henneguya linearis (Gurley) Labbé B. Myxosporidian of marine fish (Atlantic Ocean)

Rio de Janeiro: Myzidium striatum Cunha et Fonseca

C. Myxosporidian of Amphibia

Brazil: Sphaeromyza immersa (Lutz) Thélohan

EUROPE

I ITALY

A. Myxosporidia of fresh-water fish from lakes and rivers opening into Adriatic Sea

Lago di Como: Mitraspora caudata (Parisi) Kudo

Myxidium lieberkühni Bütschli

Sphaerospora elegans Thélohan Lago di Garda:

Henneguya gasterostei Parisi

Lago di Varamo: Henneguya minuta (Cohn) Lago Maggiore: Myxidium lieherkühni Bütschli Milano:

Myxidium lieberkükni Bütschli

Myxobolus pfeifferi Thélohan Myxobolus gigas Auerbach Pavia:

Myxobolus ellipsoides Thélohan Henneguya peri-intestinalis Cépède

Ticino River: Henneguya minuta (Cohn)

B. Myxosporidia of marine fish

1) Ligurian Sea

Genova: Chloromyzum leydigi Mingazzini

Gen. incert. merluccii Perugia

Gen. incert. congri Perugia

2) Tyrrhenian Sea

Napoli: Leptotheca agilis Thélohan

Leptotheca elongata Thélohan Ceratomyxa arcusta Thélohan

Ceratomyxa appendiculata Thélohan

Ceratomyxa truncata Thélohan

Ceratomyxa inaequalis Doflein

Ceratomyxa linospora Doflein

Myzoproteus ambiguus (Thél.) Doflein

Chloromyxum leydigi Mingazzini

Choromyxum quadratum Thélohan

Sphaerospora divergens Thélohan

Myxidium incurvatum Thélohan

Myxidium giganteum Doflein

Myxidium depressum Parisi

Myxidium oviforme Parisi

Sphaeromyxa balbianii Thélohan

Sphaeromyza incurvata Doflein

Sphaeromyza sabrasesi Laveran et Mesnil

Lentos pora asymmetrica Parisi

Myxobolus exiguus Thélohan

Myxobolus mülleri Bütschli

Henneguya neapolitana Parisi

II Monaco

Myxosporidia of fish from Ligurian Sea

Leptotheca elongata Thélohan

Ceratomyxa sphaerulosa Thélohan

Ceratomyza arcuata Thélohan Ceratomyza pallida Thélohan Ceratomyxa herouardi Georgévitch Ceratomyxa sp. Georgévitch Chloromyxum leydigi Mingazzini Myxidium incurvatum Thélohan Sphaeromyza sabrasesi Laveran et Mesnil

III FRANCE

A. Myxosporidia of fresh-water fish

1) From Rivers opening into Atlantic Ocean

Aigne:

Myxobolus pfeifferi Thélohan

Bretagne: Lorraine:

Sphaerospora elegans Thélohan Myxobolus oviformis Thélohan

Nancy:

Myxobolus pjeifferi Thélohan

Marne:

Myxobolus pfeifferi Thélohan Myxobolus pjeifferi Thélohan

Seine: Paris:

Chloromyxum fluviatile Thélohan

Wimereux:

Myxidium giardi Cépède

2) From Rivers opening into Mediterranean Sea

Dauphiné:

Myxobolus mülleri Bütschli

Drac River:

Myxobolus mülleri Bütschli

Myxobolus pjeifferi Thélohan

Grenoble:

Chloromyxum cristatum Léger Myxidium barbatulae Cépède

Isère River:

Myxobolus oviformis Thélohan Myxobolus mülleri Bütschli Myxobolus cycloides Gurley

Henneguya légeri Cépède

Lac d'Annecy:

Myxobolus mülleri Bütschli Lac de Paladru: Myxobolus cycloides Gurley

Lac du Bourget: Myxobolus obesus Gurley

Henneguya peri-intestinalis Cépède Henneguya tenuis Vaney et Conte

Lyon?: Rhone River:

Myxobolus pfeifferi Thélohan

Saone River:

Myxobolus pfeifferi Thélohan

B. Myxosporidia of marine fish

1) From Atlantic Ocean

Arcachon

Sphaeromyxa sabrasesi Laveran et Mesnil

Concarneau:

Ceratomyxa arcusta Thélohan Chloromyxum leydigi Mingazzini

Chloromyxum quadratum Thélohan Sphaerospora divergens Thélohan Myxidium incurvatum Thélohan Sphaeromyxa balbianii Thélohan

Le Croisic:

Leptotheca elongata Thélohan Leptotheca parva Thélohan

Leptotheca renicola Thélohan Ceratomyxa appendiculata Thélohan Myxoproteus ambiguus (Thél.) Doflein Sphaerospora rostrata Thélohan

Concarneau:

Ceratomyxa arcuata Thélohan Chloromyxum leydigi Mingazzini Chloromyxum quadratum Thélohan Sphaerospora divergens Thélohan Myxidium incurvatum Thélohan Sphaeromyza balbianii Thélohan

Roscoff:

Ceratomyxa sphaerulosa Thélohan Ceratomyza arcuata Thélohan Ceratomyxa appendiculata Thélohan Chloromyxum leydigi Mingazzini Chloromyxum quadratum Thélohan Sphaeros pora rostrata Thélohan Sphaerospora divergens Thélohan Myxidium incurvatum Thélohan Myxidium gadi Georgévitch Sphaeromyxa balbianii Thélohan

Sphaeromyxa sabrasesi Laveran et Mesnil Sphaeromyxa gasterostei Georgévitch

Myxobolus mülleri Bütschli

Le-Vivier-sur-mer: Leptotheca parva Thélohan

Myxidium sphaericum Thélohan

Myxobolus exiguus Thélohan

St.-Valery-en-caux: Ceratomyza sphaerulosa Thélohan

2) From Mediterranean coast

Marseille:

Leptotheca elongata Thélohan Leptotheca parva Thélohan Leptotheca renicola Thélohan Leptotheca hepseti Thélohan Ceratomyxa arcuata Thélohan Ceratomyxa pallida Thélohan Ceratomyxa globulifera Thélohan Ceratomyxa appendiculata Thélohan Ceratomyxa reticularis Thélohan Chloromyxum leydigi Mingazzini Sphaerospora rostrata Thélohan Myxidium incurvatum Thélohan Myxidium sphaericum Thélohan Sphaeromyxa balbianii Thélohan Myxobolus exiguus Thélohan

Banyuls:

Leptotheca elongata Thélohan

Leptotheca polymorpha (Thél.) Labbé Ceratomyxa arcuata Thélohan Ceratomyxa globulifera Thélohan Ceratomyxa appendiculata Thélohan Ceratomyxa reticularis Thélohan Chloromyxum leydigi Mingazzini Sphaerospora rostrata Thélohan Myxidium incurvatum Thélohan Myxidium sphaericum Thélohan Sphaeromyxa balbianii Thélohan Myxobolus exiguus Thélohan

Villefranche:

Ceratomyxa pallida Thélohan

Ceratomyxa truncata Thélohan Ceratomyxa coris Georgévitch Sphaeromyxa balbianii Thélohan

Locality unknown: Leptotheca agilis Thélohan

Leptotheca perlata (Gurley) Labbé Myxidium lieberkühni Bütschli Myxidium histophilum Thélohan Myxosoma dujardini Thélohan Myzobolus piriformis Thélohan Myxobolus dispar Thélohan Myxobolus obesus Thélohan Henneguya psoros permica Thélohan

Henneguya media Thélohan Henneguya brevis Thélohan Hoferellus cyprini Doflein

C. Myxosporidian in a reptile Myxidium danilewskyi Laveran

IV GERMANY

A. Myxosporidia of fresh-water fish

1) From Rivers opening into North Sea

Throughout country: Myzobolus cyprini Doflein Berlin: Henneguya oviperda (Cohn) Bodensee: Chloromyxum dubium Auerbach

Myxobolus mülleri Bütschli

Gutach: Myxobolus neurobius Schuberg et Schröder

Henneguya nüsslini Schub. et Schröder

Karlsruhe and its

Chloromyxum mucronatum Gurley vicinity: Sphaerospora elegans Thélohan

Myxidium lieberkühni Bütschli Myxidium pfeifferi Auerbach Myxidium macrocapsulare Auerbach

Henneguya ovi perda (Cohn) Henneguya lobosa (Cohn) Myxobolus gigas Auerbach

Leipzig: Myxobolus sp. Gurley Mosel:

Myxobolus pfeifferi Thélohan Myxobolus squamae Keysselitz Myxobolus cordis Keysselitz Myxobolus musculi Keysselitz

Neckar: Myxobolus exiguus Thélohan (Heidelberg)

> Myxobolus mülleri Bütschli Myxobolus pfeifferi Thélohan Myxobolus squamae Keysselitz Myxobolus cordis Keysselitz Myxobolus musculi Keysselitz Henneguya psorospermica Thélohan Henneguya acerinae Schröder (Heidelberg)

Rhine: Myxidium lieberkühni Bütschli

> Myxobolus mulleri Bütschli Henneguya psoros permica Thélohan Lentospora encephalina Mulsow

2) From Rivers opening into Baltic Sea

Alle: Myxobolus mülleri Bütschli

Frisches Haff: Myxidium lieberkühni Bütschli

Myxosoma dujardini Thélohan
Myxobolus piriformis Thélohan
Myxobolus dispar Thélohan
Myxobolus exiguus Thélohan
Myxobolus oviformis Thélohan
Myxobolus mülleri Bütschli
Myxobolus cycloides Gurley
Myxobolus anurus Cohn
Myxobolus sp. Wegener
Myxobolus permagnus Wegener
Henneguya psorospermica Thélohan

Henneguya texta (Cohn)
Henneguya minuta (Cohn)
Henneguya lobosa (Cohn)
Henneguya creplini (Gurley)

Kurisches Haff: Myxosoma dujardini Thélohan

Myxobolus exiguus Thélohan Myxobolus oviformis Thélohan Myxobolus cycloides Gurley Henneguya psorospermica Thélohan

Henneguya texta (Cohn)

Henneguya creplini (Gurley) Labbé

Masurische Seen: Sphaerospora masovica Cohn

Myxobolus dispar Thélohan Myxobolus ellipsoides Thélohan Myxobolus cycloides Gurley Myxobolus anurus Cohn

Henneguya psorospermica Thélohan

Henneguya texta (Cohn)

Pregel: Myzosoma dujardini Thélohan

Myxobolus piriformis Thélohan Myxobolus dispar Thélohan Myxobolus exiguus Thélohan Myxobolus oviformis Thélohan Myxobolus mülleri Bütschli Myxobolus cycloides Gurley Myxobolus anurus Cohn Myxobolus permagnus Wegener Henneguya psorospermica Thélohan

Henneguya texta (Cohn)
Henneguya minuta (Cohn)
Henneguya lobosa (Cohn)

Henneguya creplini (Gurley) Labbé

Myxobolus cyprini Doflein

3) Localities unknown: Chloromyxum leydigi Mingazzini

Weichsel:

Myxidium lieberkühni Bütschli

Myxidium sp. Gurley

Lentos pora cerebralis (Hofer) Plehn

Henneguya schisura (Gurley) Labbé

Hojerellus cyprini Doslein Gen. et spec. incert. Leydig Gen. et spec. incert. Leydig Gen. et spec. incert. Leydig Gen. et spec. incert. Borne

V NETHERLAND

Myxosporidian of marine fish Chloromyxum quadratum Thélohan

Helder:

VI ENGLAND

Myxosporidia of marine fish

Firth of Clyde, More-

camb, etc.: Liverpool (?): Myzobolus aeglefini Auerbach Sphaerospora platessae Woodcock

VII NORWAY

Myxosporidia of marine fish

Abelvaer:

Myxidium bergense Auerbach Myxidium ovijorme Parisi Zschokkella hildae Auerbach Myxobolus aeglefini Auerbach

Lepiotheca parva Thélohan

Bergen:

Leptotheca macrospora Auerbach
Leptotheca informis Auerbach
Leptotheca longipes Auerbach
Ceratomyxa sphaerulosa Thélohan
Myxidium incurvatum Thélohan
Myxidium inflatum Auerbach
Myxidium bergense Auerbach
Myxidium procerum Auerbach
Sphaeromyxa hellandi Auerbach
Zschokkella hildae Auerbach
Myxobolus aeglefini Auerbach

Bergsfjord:

Myxidium bergense Auerbach Zschokkella hildae Auerbach Zschokkella hildae Auerbach

Boadsfjord: Bodö:

Myxidium bergense Auerbach Zschokkella hildae Auerbach

Finkongkjeilen:

Ayridium bergense Auerbach

Zschokkella hildae Auerbach

Grönöy:

Myxidium bergense Auerbach Zschokkella hildae Auerbach

Hammerfest:

Myxidium bergense Auerbach Myxidium ovijorme Parisi Zschokkella hildae Auerbach

Harstad:

Myxidium bergense Auerbach Zschokkella hildae Auerbach

Honnigsvaag:

Myxidium bergense Auerbach Zschokkella hildae Auerbach

Kabelvaag:

Ceratomyxa drepanopsettae Awerinzew

Myzidium bergense Auerbach

Zschokkella kildae Auerbach

Kiberg: Zschokkella kildae Auerbach
Kirkenes: Myxidium bergense Auerbach
Kristiana: Myxidium bergense Auerbach
Kristiansand: Lieptoliene parea Thélohan

Leptotheca macrospora Auerbach Myzidium oviforme Parisi Zschokkella hildae Auerbach

Lödingen: Myxobolus aeglesini Auerbach
Makur: Myxidium bergense Auerbach

Zschokkella hildae Auerbach

Mosjöen: Zschokkella kildae Auerbach

Myxidium bergense Auerbach

Nusfjord: Myxidium bergense Auerbach

Zschokkella kildae Auerbach

Rörvik: Ceratomyza drepanopsettae Awerinzew

Myxidium bergense Auerbach

Rossfjord: Myxidium oviforme Parisi Skjervö: Zschokkella hildae Auerbach Skjötningsberg: Zschokkella hildae Auerbach

Myzidium bergense Auerbach

Smalfjorden: Sphaerospora divergens Thélohan

Zschokkella kildae Auerbach Leptotheca parva Thélohan

Stavanger: Leptotheca parva Thélohan

Myxidium bergense Auerbach
Myxidium bergense Auerbach

Svolvaer: Myxidium bergense Auerbach
Zschokkella hildae Auerbach

Leptotheca informis Thélohan

Ceratomyxa drepanopsettae Awerinzew

Myxidium bergense Auerbach

Torghatten: Sphaeromyza kellandi Auerbach
Trondhjem: Myzidium bergense Auerbach

Myxidium oviforme Parisi

Zschokkella hildae Auerbach

Vikholmen: Zschokkella hildae Auerbach
Vardö: Myzidium bergense Auerbach

Myxidium oviforme Parisi Zschokkella hildae Auerbach Myxobolus aeglefini Auerbach

VIII SWITZERLAND

Myxosporidia of fresh-water fish

1) From Lakes connected with North Sea

Tjömö:

Thun:

Neuchatel: Myxobolus fuhrmanni Auerbach

Myxobolus mülleri Bütschli Henneguya oviperda (Cohn)

Henneguya zschokkei (Gurley) Doslein Henneguya zschokkei (Gurley) Doslein

Zurich: Henneguya zschokkei (Gurley) Doslein Lucerne: Myzosoma dujardini Thélohan

Myxobolus ellipsoides Thélohan

Wallen:

Myzobolus oviformis Thélohan Myxobolus mülleri Bütachli Henneguya psorospermica Thélohan

Henneguya texta (Cohn)

Henneguya zschokkei (Gurley) Doslein Henneguya zschokkei (Gurley) Doflein

Gen. et spec. incert. Nufer

2) From Lake connected with Mediterranean Sea

Geneva: Myxobolus sphaeralis Gurley

Henneguya zschokkei (Gurley) Doflein

IX AUSTRIA

A. Myxosporidia of fresh-water fish

1) From Rivers opening into Black Sea

Danube tributaries Chloromyxum thymalli Lebzelter

and Neusiedler: Myxosoma (?) lobatum Nemeczek Myxobolus aeglefini Auerbach

Myxobolus cyprini Doflein Myxobolus rotundus Nemeczek Myxobolus minutus Nemeczek Myxobolus sp. Lebzelter Henneguya acerinae Schröder Henneguya gigantea Nemeczek

2) From Rivers opening into North Sea

Myxosoma dujardini Thélohan Prag:

> Myxobolus ellipsoides Thélohan Myxobolus oculi-leucisci Trojan

Krakau: Myxobolus cyprini Doflein

B. Myxosporidia of marine fish (Adriatic Sea)

Rovigno: Leptotheca agilis Thélohan

Ceratomyxa pallida Thélohan Ceratomyxa appendiculata Thélohan Myxoproteus ambiguus (Thél.) Doflein Chloromyxum leydigi Mingazzini

Sphaeromyxa sabrasesi Laveran et Mesnil

Locality unknown: Gen. et spec. incert. Heckel et Kner

C. Myxosporidian of Amphibia

XI RUSSIA

Vienna: Chloromyxum protei Joseph

X SERBIA

Pergrad (Danube): Henneguya gigantea Nemeczek

A. Myxosporidia of fresh-water fish

Volga (to Caspian

Sea):

Lentos pora multiplicata Reuss Myxobolus volgensis Reuss Myxobolus scardinii Reuss Myxobolus physophilus Reuss Myxobolus macrocapsularis Reuss

Myxobolus sandrae Reuss Myxobolus bramae Reuss Myxobolus cyprinicola Reuss Myxobolus balleri Reuss

Locality unknown:

Don (to Black Sea): Myxobolus sp. Gurley Zschokkella nova Klokacewa

Myxobolus magnus Awerinzew Myxobolus carassii Klokacewa Henneguya kolesnikovi (Gurley) Labbé

B. Myxosporidia of marine fish from Arctic Ocean

Murman coast:

Ceratomyxa ramosa Awerinzew Ceratomyxa drepanopsettae Awerinzew

Myxidium sp. Awerinzew Leptotheca sp. Awerinzew Chloromyxum sp. Awerinzew

B. DISTRIBUTION OF MYXOSPORIDIA IN ANIMALS

The number of host species that harbor Myxosporidia is 237, as will be seen from List III.

Tho two incompletely studied forms are found in Annelida and Insecta, Myxosporidia are the parasites of Vertebrata, especially of Pisces, only few being found infecting Amphibia and Reptilia. They are distributed among these groups of animals as follows:

	Number of host species
Annelida	1
Insecta	1
Pisces	223
Amphibia	8
Reptilia	

Gurley (1894:101-105), Wasielewsky (1896:132-148), Labbé (1899: 133-161) and Auerbach (1910: 36-45; 1911: 471-494) gave lists in which they recorded the names of host species. Wasielewsky arranged the names alphabetically while others listed them according to their systematic order. In the following pages, the writer followed Wasielewsky, i.e., the names of the host species are arranged alphabetically as is supposed to be more convenient in referring to the host than any form presented otherwise.

LIST III. LIST OF HOST SPECIES

Host	Organ Infected	Myxosporidian	Locality
Annelida			
Nais lacustris (N. probo-			
scid ea)	Unknown	Myxobolus sp.	Germany
Insecta			1
Tortrix viridana L. (imago)	Abdominal cav-		ļ
· · · · · · · · · · · · · · · · · · ·	ity	Chloromyxum diploxys	France
Pisces			
Abramis ballerus L	Branchiae	Myxobolus balleri	Russia
A. brama L	"	bramae	"

		<u> </u>	
Host	Organ Infected	Myxosporidian	Locality
	Branchiae	Myxobolus cycloides	Germany
	46	elli psoides	"(?)
	44	exiguus	France
	"	oviformis	France
	"	rotundus	Austria
	Gall-bladder	Sphaerospora masovica	Germany
	Kidney	Myxobolus cyprini	",
	_		Hungary
	Subcut. conn.	gigas	Germany,
	tissue of oper-		Italy
	culum		
A. simba L	Branchiae	cycloides	Germany
	at .	elli psoides	" (?)
	"	oviformis	Germany
Acanthias acanthias L	Gall-bladder	Chloromyxum leydigi	France
A. blainvillei	"	magnum	Africa
Acerina cernua L	Branchiae	Henneguya acerinae	Germany
	", Muscle	creplini .	"
	Conn. tissue of	tenuis	France
	aliment. canal		
	Eye	Myxobolus magnus	Russia
	Muscle	Lepiotheca perlata	France?
Acheilognathus lanceolatum	Gall-bladder.		}
Temm. et Schl	Gall-duct	Zschokkella acheilognathi	Nippon
Alburnus alburnus L	Branchiae	Myxobolus cycloides	Germany
(A. lucidus Heck)	"	dispar	4
	66	elli psoides	Germany?
	"	oviformis	France
	", kidney	obesus	"
	Muscle and	dispar	"
	spleen	•	ļ
	Eye	mülleri	Switzerland
Alosa finta Cuv. var. lacus-			İ
tris Fatio	Kidney	Mitras pora caudata	Italy
Ameiurus melas Raf	Base of spines of	•	
	2nd dorsal fin	Henneguya gurleyi	U. S. A.
Ancylopsetta quadrocellata			
Gill	Gall-bladder	Ceratomyxa undulata	44
Anguilla japonica Temm.	Integument	M yzidium anguillae	Nippon
et Schl	"	Lentos pora dermatobia	- "
A. vulgaris Flemm	Kidney	Myxidium giardi	France
Aphredoderus sayanus Gill.	Subcutaneous		1
-	intermusc. tiss.	Неппедиуа топита	U. S. A.
Apogon rex-mullorum Cuv.	Gall-bladder	Myxidium oviforme	Italy
Argentina silus Nilss	"	procerum	Norway
Ariodes polystaphylodon	Muscle	Chloromyxum quadratum	Africa
Aspius rapax Ag	Branchiae	Myxosoma(?) lobatum	Austria
Aspro asper L	"	Myxobolus mülleri	France
A. zingel Cuv	"	Henneguya acerinae	"



Host	Organ Infected	Myxosporidian	Locality
Atherina hepsetus L	Gall-bladder	Leptotheca hepseti	France
Boirdiella chrysura		Myxoproleus cornulus	U. S. A.
B. rouchus Cuv. et Val	Gall-bladder	Myxidium striatum	Brazil
Barbus barbus L	Kidney, spleen,		
(B. fluviatilis)	intestine, ovary,	Myxobolus pfeifferi	France, Germany
	Inner surface of	squamae	Germany
	Muscle of ven-	cordis	"
	Muscle, kidney	musculi	66
B. plebejus Val		pfeifferi	Italy
B. vulgaris Flem		milleri	Germany
Barilius barna	Under scales	Sphaerospora sp.	Burma
Belone acus Risso	Gall-bladder	Myxidium sphaericum	France
B. belone L	"	Myxidium sphaericum	66
Blennius ocellatus	"	Sphaeromyxa incurvata	Italy
B. gattorugins Brunn	Kidney	Chloromyxum quadratum	"
B. pholis L	Gall-bladder	Myxidium incurvatum	France
	Renal tubules	Sphaerospora divergens	"
Blicca björkna L	Branchiae	Myxobolus cycloides	Germany
	"	elli psoide s	" ?
	"	macroca psularis	Russia
	"	oviformis	Germany
Box boops L	Gall-bladder	Ceratomyxa pallida	France, Italy
B. salpa L	Gall-bladder	Ceratomyxa herouardi	Monaco
, in the second	"	pallida	France,
			Italy
	Kidney	Henneguya neapolitana	Italy
Brevoortia tyrannus Latr	Muscle	Chloromyxum clupeidae	U. S. A.
Brosmius brosme Ascanius	Gall-bladder	Leptotheca longipes	Norway
	44	Sphaeromyxa hellandi	"
Callionymus lyra L	66	Myxidium incurvatum	France, Norway
	Muscle	Chloromyxum quadratum	France
Corossius ourotus L	Branchiae	Leniospora acuta	Nippon
	Kidney	Sphaerospora angulata	"
	44	Mitraspora cyprini	"
	Subcutaneous		1
	tiss. of head	Henneguya miyairii	"
(C. carassius L.)	Body cavity	Myxobolus sp.	Germany
	Branchiae	dispar	"
(7)		Sphaerospora carassii	Nippon
(C. vulgaris L.)	Body cavity,		l
	liver, intestine	Myzobolus carassii	Russia
	Gall-bladder	Zschokkella nova	· "
Carcharhinus limbatus	"	Chloromyxum leydigi	U. S. A.
C. sp	••	Ceratomyxa flagellifera	<u> </u>

Host	Organ Infected	Myxosporidian	Locality		
Carpiodes difformis	Branchiae	Myxobolus discrepans	U. S. A.		
Catostomus commersonii		1	1		
Lac	Gall-bladder	Chloromyxum catostomi	"		
Centronotus gunellus	"	Sphaeromyxa kellandi	Norway		
Cepola rubescens L	"	balbianii	France		
Cestracion tiburo	"	Chloromyxum leydigi	U. S. A.		
	"	Ceratomyxa mesospora-	66		
C. sygaena	"	Ceratomyxa mesospora	"		
	"	recurvala	"		
	"	Chloromyxum leydigi	**		
	"	Leptotheca fusiformis	"		
Chaetodipterus faber	Gall-bladder	Ceratomyxa streptospora	"		
	Urin. bladder	Myxoproteus cordiformis	"		
Chondrostoma nasus L	Branchiae	Myxobolus exiguus	Germany		
	"	Gen. et spec. incert.	Switzerland		
	Tongue	Gen. et spec. incert.	3		
Citharus linguata Gthr	Gall-bladder	Myxidium depressum	Italy		
Clupea harengus Young	"	Ceratomyxa sphaerulosa	Norway		
orn for more many a company	Muscle	Chloromyxum clupeidae	U. S. A.		
C. pilchardus Walb.	1-1 45010		0.0		
(Alosa sardina)	Gall-bladder	Ceratomyxa truncata	France, Ital		
(220030 307007007	"	Sphaeromyxa balbianii	Italy		
Cobitis barbatula L	Kidney	Myxidium barbatulae	France		
200,000	Urin. bladder	Henneguya legeri	"		
C. fossilis L		Hennegayo regers			
Conger conger L.	kidney, spleen	Myxobolus piriformis	Germany		
(Leptocephalus c.)		Gen. incert. congri	Italy		
Coregonus lavaretus L.	Cur. Danador	Com. most a congre	1		
(C. fera)	Branchiae	Henneguya sp.	France?		
(0.)0,0,	" (mucosa)	Myxobolus sphaeralis	Switzerland		
	Muscle	Henneguya kolesnikovi	Russia		
	"	sschokkei	Switzerland		
C. exiguus albellus	", branchia	"	"		
C. wartmanni nobilis	", brancina	66	4		
Coris giofredi Risso	Gall-bladder	Ceratomyxa coris	France		
C. julis L	"	"	"		
C. J	Muscle	Chloromyxum quadratum	"		
	Gall-bladder	Myxidium oviforme	Italy		
Cottus gobis L		Myxobolus mülleri	France		
C. scorpius	Diancinae "	Myxidium sp.	Russia		
Crenilabrus mediterraneus	"	Ceratomyxa inaequalis			
	1		Italy		
C. melops L	Eye	M yxobolus mülleri	Germany		
,	Call bladder	Canatamana accusts	France		
	Gall-bladder	Ceratomyxa arcuata	France		
C A C1 77 3	Kidney	Sphaerospora divergens			
C. \$000 Cuv. et Val	Gall-bladder	Ceratomyxa inaequalis	Italy		
ļ	Kidney	Lentospora asymmetrica	Italy		
		Sphaerospora divergens	France, Ital		
Cyclopterus lumpus L	Gall-bladder	M yxidium inflatum	Norway		



Host	Organ Infected	Myxosporidian	Locality
Cynoscion regalis	Gall-bladder	Myxidium glutinosum	U. S. A.
Catalantaniantan	Urin. bladder, ureters	Sinuolinea dimorpha	"
Cyprinodon variegalus	Subcutaneous tissue Visceral conn.	Myxobolus lintoni	"
	tissue	ca psulatus	"
Cyprinus carpio L	Branchiae	cy prinicola	Russia
	"	dis par *	France.
			Germany
	"	oviformis	France?
	"	toyamai	Nippon
		Myxosoma dujardini	France
	"	Myxobolus koi	Nippon
	Brain	Lentospora encephalina	Germany
	Gall-bladder	Chloromyxum koi	Nippon
	Kidney	Hoferellus cyprini	Germany, France
	" liver, spl.	Myxobolus cyprini	Germany, Hungary
	Kidney	Mitraspora cyprini	Nippon
	"	Sphaerospora angulata	"
C. (Rasbora) daniconius	Subcutaneous		1
Day	intermuscular		ļ
	tissue	Myxobolus sp.	India
.	Muscles	Myxobolus nodularis	India
Dasybatis hastatus	Gall-bladder	Chloromyxum leydigi	U. S. A.
D seline	l ::	Leptotheca scissura	"
D. sabina		Chloromyxum leydigi	"
Drepanopsetta platessoides Fabr	"	Ceratomyxa drepanopsettae	Russia
Erimyson sucetta oblongus		Ceratomyza arepanopsenae	Russia
Lac (Catostomus	Branchiae	Myxobolus globosus	U. S. A.
tuberculatus)	Integument	oblongus	"
Esox lucius L	Branchiae	anurus	Germany
	"	Henneguya lobosa	"
	66	Henneguya psorospermica	France, Germany
	Intestinal wall	Henneguya peri-intestinalis	France, Italy
	Eye muscle, etc.	Henneguya schizura	U. S. A.
	Ovary	Henneguya oviperda	Germany, Switzerland
	Urin, bladder	Myxidium lieberkühni	France, Italy, Canada, U. S. A.,
Fundulus sp	Muscle	Chloromyxum funduli	Germany U. S. A.



			
Host	Organ Infected	Myxosporidian	Locality
F. diaphamus	Muscle	Myxobolus funduli	U. S. A.
F. heteroclitus	Branchiae,		"
	Muscle	Myxobolus funduli	1
	Branchiae	Myxosoma funduli	"
F. majalis	"	Myxosoma funduli	"
	", muscle		"
	Gall-bladder	Myxidium incurvatum	- "
Gadus aeglesinis L	Cartilage	Myxobolus aeglefini	Germany
•	Urin. bladder	Zschokkella kildae	Norway
G. callarias L	Cartilage	M yxobolus aeglefini	Germany
	Gall-bladder	Myxidium oviforme	Norway
	Urin. bladder	Zschokkella hildae	Norway
G. esmarkii Nilss	Cartilage, bone		Ì
	of cranium,		1 _
	eye	M yxobolus aeglefini	Germany, England
G. merlangus L	Gall-bladder	Leptotheca informis	Norway
O. Mersongas D	Cartilage	Myxobolus aeglefini	Germany
G. morrkus L	_	Myxobolus aeglefini	Germany
G. pollachius L		Myxidium gadi	France
G. virens L	"	bergense	Norway
	Urin, bladder	Zschokkella hildae	"
Galeocerda tigrinus	Gall-bladder	Ceratomyxa lunata	U. S. A.
Galeus galeus L. (G. canis)	"	sphaerulosa	France
Gambusia affinis	"	Myxidium incurvatum	U. S. A.
<u></u>	"	phyllium	"
Gasterosteus aculeatus	Kidney (r. t.),		
	ovary	Henneguya media	France
	"	brevis	44
	"	Sphaerospora elegans	", Italy
	Kidney	Henneguya gasterostei	Italy
G. pungitius L	" (r. t.), ovary	brevis	France
	` "	media	"
	j.	Sphaerospora elegans	"
G. spinachiaGobio gobio L.	Gall-bladder	Sphaeromyxa gasterostei	"
(G. fluviatilis)	Fin	Myxobolus mülleri	Germany
(G. jiwaans)	Fin, spleen,	m yaooona muueri	Germany
	kidney, liver	oviformis	France
	Branchiae	cycloides	
Gobius fluviatilis L	Body-cavity		Germany
Gobius paganellus L		Gen. et. spec. incert.	Italy
Heliases chromis Gthr	"	Ceratomyxa arcuata Ceratomyxa arcuata	Italy.
nasaa aa omaa Gun		Сегаютуха отсымо	Monaco
Hippocampus brevirostris			i monaco
Cuv	"	M yxidium incurvalum	Italy
~4 T	"	Sphaeromyxa sabrasesi	France,
		D procesomy sour uses	Italy,
			Hungary
			11ungary



Host	Organ Infected	Myxosporidian	Locality
H. guttulatus Cuv	Gall-bladder	Sphaeromyxa sabrasesi	Fr., Hun.,
Hippoglossoides liman-			Monaco
doides	Urin. bladder	Sphaerospora divergens	Norway
Hippoglossus vulgaris			
Flemm	Gall-bladder	Ceratomyxa drepanopsettae	Norway
	"	ramosa	Russia
Hybognathus nuchalis Ag	Conn. tissue of		1
	the head	Henneguya macrura	U. S. A.
Idus melanotus Heck	Branchiae	Myxobolus ellipsoides	Germany?
	Muscle	Lentos por a multi plicata	Russia
Labeo rokita	Branchiae	Myxobolus rokitae	India
	Fins	Myxobolus seni	"
Labeo niloticus Forsk	?	Myxobolus unicapsulatus	Egypt
Labrus turdus	Gall-bladder	Ceratomyxa linospora	Italy
Leiostomus xanthurus	"	aggregata	U. S. A.
Lepisosteus platystomus	Urinary bladder	Sphaerospora sp.	"
Lepomis cyanellus Raf	Mesentery	Myxobolus mesentericus	"
	Urin. bladder	Henneguya mictos pora	u
	Kidney ~	Mitraspora elongata	"
L. humilis Girard	Ovary	Wardia ovinocua	"
	Urinary bladd.	Henneguya mictospora	"
L. megalotis Raf	Gall-bladder	Chloromyxum trijugum	"
Leuciscus cephalus (Squalis	Air-bladder,		France,
cephalus)	branchiae	Myxobolus mülleri	Germany
• • •	Branchiae	elli psoides	" "
	Gall-bladder	Chloromyxum fluviatile	France
L. lucius L	Branchiae	Myxobolus sp.	Germany?
L. phoxinus L. (Phoxinus	Conn. tiss. of		1
laevis Ag.)	kidney; ovary	Myxidium histophilum	France
	"	Myxobolus mülleri	"
L. rutilus L	Branchiae "	elli psoides mülleri	Germany?
	"	Myxosoma dujardini	France
	Conn. tiss. un-	May 30000 ma Gay ar arms	Trance
	der the mouth		1
	muc. mem-		1
	brane	Myxobolus fuhrmanni	Switzerland
	Opercle, pseu-		- Switzeranza
	do branchiae,		France.
	kidney	cycloides	Germany
	Vitreous body	- Cyaronacas	00
,	of eye	oculi-leucisci	Austria
,	7	Henneguya sp.	Germany
ļ	Heart	Gen. et spec. incert.	"
Leuciscus sp	Branchiae	Myxobolus minutus	Austria
	"	Myxosoma (?) lobatum	"
Lophius budegassa Spin	Gall-bladder	Ceratomyxa appendiculata	Italy,
To Lucian augustance of transmission	~ D	J. J. Jan of Police Committee	**
			France

Host	Organ Infected	Myxosporidian	Locality
L. piscatorius L	Urin. bladder	M yxoproteus ambiguus	France, Italy,
			Hungary
Lota lota L. (L. vulgaris)	Branchiae	Myxobolus mülleri	Germany
	"	Myxobolus cycloides	Germany
	Gall-bladder	Chloromyxum dubium	" , Austria
	Urin. bladder,		
	kidney	mucronatum	France
	Urin. bladder	Myxidium lieberkühni	France,
	44		Germany
	-	Sphaerospora elegans	Germany
Lucioperca lucioperca L (L. sandra Cuv.)	Branchiae ", head, fin,	Henneguya acerinae	", Austria
•	opercle	Myxobolus sp.	3
	44	Henneguya gigantea	Aus., Servia
	"	Gen. et sp. incert.	Austria
· - · · · · · · · · ·	Muscle	Myxobolus sandrae	Russia
L. volgensis Pall	Branchiae, cor-		
	nea, dorsal fin	volgensis	
Melanogrammus aeglefinis	Gall-bladder	Myxidium bergense	Canada
Menticirrhus americanus L.	••	striatum	Brazil
Merluccius merluccius L.	"	G 4 Labeltone	F
(M. vulgaris)	"	Ceratomyxa globulifera	France
	"	Leptotheca elongata Gen. incert. merluccii	", Italy
Micropogon undulatus	"	Ceratomyxa aggregata	U. S. A.
Micropterus salmoides Lac Misgurnus anguillicau-	Urin. Bladder	Henneguya miclospora	U. S. A.
datus Cantor	Branchiae	Myxobolus sp.	Nippon
borns Canton	Gall-bladder	Chloromyxum fujitai	""
	(f	misgurni	· ·
		Myxidium kagayamai	"
	44	Myxobolus misgurni	"
Molva vulgaris Flem	Bone	aeglefini	Austria
	Gall-bladder	Le plotheca informis	Norway
	46	Sphaeromyxa hellandi	"
Motella maculata Risso	46	balbianii	France
M. tricirrata Bl	"	Ceratomyxa arcuata	"
·	"	Leptotheca elongata	", Monaco
	"	Sphaeromyxa balbianii	France
	"	sabrazesi	Monaco
Mugil auratus Risso	Intestine, stom- ach, spleen,		
	pyloric coecum	Myxobolus mülleri	Italy
	Kidney	exiguus	", France
M. capito Cuv	"	exiguus	"
M. cephalus L	Gall-bladder	Myxidium incurvatum	U. S. A.
M. chelo Cuv	Stomach,spleen,		Italy,
	kidney, etc.	M yxobolus exiguus	France

Host	Organ Infected	Myxosporidian	Locality
<u>И. sp</u>	Kidney	Sphaerospora rostrata	Italy, France
Muraena sp	Gall-bladder	Ceratomyxa sp.	Monaco
Mustelus canis Mitch.			
(M. vulgoris)	Gall-bladder	Ceratomyxa sphaerulosa	France
Nerophis aequoreus L	Gall-bladder	Myxidium incurvatum	France
•	Muscle	Chloromyxum quadratum	"
N. annulatus	Gall-bladder	Myxidium incurvatum	66
	"	Sphaeromyxa sabrazesi	Monaco
N. lumbriciformis	"	Myxidium incurvatum	France
Notropis megalops Raf	Subcut. tissue	Gen. et spec. incert.	U. S. A.
N. gilberti J. et M	Muscle	Myxobolus orbiculatus	U. S. A.
N. blennius	"	"	"
N. anogenus	Fins	aurealus	"
· ·	"	Henneguya brachyura	"
Oncorkynchus keta	Under the skin	Henneguya salminicola	Kamtschatka
O. kisutch	"	"	"
	Connective tiss.		
	of body muscle	"	Alaska
O. nerko	Gall-bladder	Chloromyxum wardi	"
Ophidium vasalli		Ceralomyxa arcuala	Monaco
Opsanus lau	Urin. bladder	Sphaerospora polymorpha	U. S. A.
Pagellus centrodontus Del	Gall-bladder	Ceratomyxa arcuata	France, Italy
Paralichthys albiguttus			lary
J. et G	Urin. bladder	navicularia	U. S. A.
_	"	spinosa	"
	"	Leptotheca glomerosa	"
	"	Sinuolinea brachiophora	"
	"	capsularis	"
	"	opacita	"
P. dentatus	Gall-bladder	Ceratomyxa drepanopsettae	"
	Urin. bladder	' navicularia	"
	"	Leptotheca lobosa	"
	"	Sinuolinea capsularis	"
Parasilurus asotus L	Intestinal wall	M yxobolus miyairii	Nippon
Peprilus alepidotus	Gall-bladder	Ceratomyxa monospora	U. S. A.
Perca flavescens	Spleen	Myxobolus sp.	"
	Urin. bladder	Henneguya wisconsinensis	"
P. fluviatilis	Branchiae	• lexia	Italy,
			Germany
	"	Henneguya minuta	"
	"	Myxobolus sp.	Germany
	", operculum	permagnus	"
	"	M yxosoma dujardini	Switzerland
		Henneguya psorospermica	"
Phoxinus (Clinostomus)	Under scales on		
funduloides Girard	ext. surf.	Myxobolus transovalis	U. S. A.
P. laevis	Branchiae	mülleri mülleri	France



Host	Organ Infected	Myxosporidian	Locality			
P. laevis	Kidney, ovary	Myxidium kistophilum	France			
	Urin. bladder	Sphaerospora elegans	Germany			
Phycis blennioides Br	Urin. bladder	Zschokkella kildae	Norway			
P. mediterraneus (P.						
pkycis L.)	. Gall-bladder	Leptotheca polymorpha	France			
Pimelodus sebae Cuv. et	Membrane lin-					
Val	ing branchial	1	1			
	cavity	Henneguya linearis	S. America			
Pimephales notatus Raf	Gall-bladder	Myxobolus notatus	Canada			
Piramulana blocki C. et V.	.] ?	inaequalis	S. America			
Platystoma fasciatum L	Branchiae	Henneguya linearis	"			
Pleuronectes flesus L	Gall-bladder	? Ceratomyxa drepanopsettae	Norway			
P. platessa L		"	"			
	Otic-capsule	Sphaerospora platessae	England			
Pomolobus aestivalis	Muscle	Chloromyxum clupeidae	U. S. A.			
P. mediocris Mitch	"	clupeidas	4			
P. pseudoharengus Young	"	clupeidas	u			
Pseudo pleuronectes amer-			1			
iconus	Gall-bladder	Ceratomyxa acadiensis	Canada			
	"	Myxidium sp.	"			
	Subcutaneous		Ì			
	muscl. tissue	Myxobolus pleuronectidae	U. S. A.			
Pteroplatea maclura						
Le Sueur	Gall-bladder	Chloromyxum leydigi	u			
	"	Leblotheca scissura	4			
Raja asterias	"	Myxidium giganteum	Italy			
R. batis L.	"	Chloromyxum leydigi	France			
	Gall-duct	Myxidium sp.	Germany?			
R. radiata	Gall-bladder	Chloromyxum sp.	Murman			
		Cancer can yan and Cp.	Coast			
R. clavata L	"	Chloromyxum leydigi	France			
R. undulata Lac	"	Chloromyxum leydigi	4			
Rhina squatina L	"	Chloromyxum leydigi	France,			
			Germany			
Rhinobathus sp. (?) Awer	"	Ceratomyxa sp. (?)	Africa			
Rhodeus amarus Bl	Branchiae	Myxobolus cycloides	Germany			
Salmo fontinalis Mitch	Cartilage, pe-		,			
J	richondrium	Lentos por a cerebralis	44			
Scardinius erythrophthal-						
##\$	Branchiae	Myzobolus cycloides	66			
	"	scardinii	Russia			
ļ	"	Myxosoma dujardini	France,			
l			Germany			
l	Gall-bladder	Myxidium macrocapsulare	"			
	Muscle, spleen	M yxobolus dispar	France			
	Air-bladder	permagnus	Germany			
	· m-marifier	pkysophilus	Russia.			
Scatophagus argus	Gall-bladder	Ceratomyza sp. (?)	Africa			
State handles on Eng	Can-Diadici	Coronomyste sp. (r)	THE			



Host +	Organ Infected	Myxosporidian	Locality
Scoliodon terrae-novae	Gall-bladder	Ceratomyxa abbreviata	U. S. A.
	44	attenuata	"
	"	s p hairo p hora	"
	44	laenia	"
	u	Chloromyxum leydigi	"
Scomber scombrus L	44	Leptotheca parva	France, Norway
	Kidney	renicola	France
	?	Gen. et sp. inc.	Germany?
Scorpaena porcus L	Gall-bladder	Ceratomyxa arcuata	France
S. scrofa L	"	Ceratomyxa arcuata	"
5. s 3,0 =	66	Myxidium incurvatum	"
S. sp	66	Leptotheca agilis	", Germany
Scyllium canicula	44	Ceratomyxa sphaerulosa	Monaco
00,000,000,000,000,000,000,000,000,000	cc .	Chloromyxum leydigi	", France
			Germany
S. asterias	46	"	Italy
Sebastes dactylopterus		Leptotheca macros pora	Norway
S. norvegicus		Leptotheca sp.	Eastern
		•	Finmark
S. viviparus H. Kr	.,	Leptotheca macrospora	Norway
Siphonostoma rondeletii		Sphaeromyxa sabrazesi	Monaco
Siphostoma floridas	"	balbianii	U. S. A.
	Urin. bladder	Sinuolinea arborescens	"
S. louisianae		Sphaeromyxa balbianii	"
Solea vulgaris		Myxidium gadi	France
Sparus berda	"	Ceratomyxa (?) spari	Africa.
Sphaeroides maculatus	Urin. bladder	navicularia	U. S. A.
	"	Sinuolinea capsularis	"
	"	Zschokkella globulosa	"
Spinax spinax L	Gall-bladder	Chloromyxum leydigi	Italy, France
Squalis agassisii Heck S. a. Savigny Bona-	Branchiae	Myxobolus mülleri	France
parte	"	Myxobolus mülleri	44
Stenotomus chrysops L	Muscle	Chloromyxum clupeidae	U. S. A.
Stisostedion vitreum Mitch.	1	Gen. et sp. incert.	Canada
Syngnathus acus L	Gall-bladder	Myxidium incurvatum	France
	"	Sphaeromyza sabrazesi	Italy, Monaco
	Muscle	Chloromyxum quadratum	France
S. typhle	Gall-bladder	Myxidium incurvatum	France
Synodontis schall Bl. Schn			
- Jacobson Comme Da Commis	cephalic reg.	Henneguya strongylura	Egypt
	> copulation reg.	Myxobolus inaequalis	S. America
Synodus faetans	Gall-bladder	Ceratomyxa agglomerata	U. S. A.
~ j j 00.0 /N	"	Gmorphs	0. S. A.
Tautogolabrus adspersus	1	отот рыс	1
Walb	Muscle	Chloromannon alcabaidas	
44 9TD	Muscie	Chloromyxum clupeidae	



			
Host	Organ Infected	Myxosporidian	Locality
Thymallus thymallus L	Gall-bladder "	Chloromyxum thymalli Myxobolus sp.	Austria
	Neurilemma (?)	pfeifferi	Germany?
Thysanophris japonicus	Gall-bladder	Sphaeromyxa exneri	Africa
Tinca tinca L. (T. vulgaris).	Branchiae	Myxobolus piriformis	France,
			Germany
	Air-bladder,		
	kidney, etc.	elli psoide s	"
	Gall-bladder	Chloromyxum cristatum	France
	"	Myxidium þfeifferi	Germany
	Kidney	Myxobolus cyprini	Germany,
			Hungary
Torpedo narce Risso	Gall-bladder	Chloromyxum leydigi	France
T. ocellata	"	Chloromyxum leydigi	Germany
T. torpedo L		Chloromyxum leydigi	"
Trachinus draco L	9	Ceratomyxa reticularis	"
	"	Myxidium incurvatum	"
Trachurus trachurus L	Muscle	Chloromyxum quadratum	France,
			Germany
Trutta fario L	Gall-bladder,		1
	Gall-duct	Chloromyxum truttae	France
	Nervous syst.	Myxobolus neurobius	Germany
	Subcutaneous		
•	conn. tiss. at		1
	base of fin	Henneguya nüsslini	"
T. iridea Gibb	Cartilage, peri-		
	chondrium	Lentospora cerebralis	"
T. salar L	,,		1
Ø	Gall-bladder	Myxidium oviforme	Norway
Trygon pastinaca L	Gall-bladder	Chloromyxum leydigi	France
T.1	TT 1.1	Leptotheca agilis	", Italy
Tylosurus marianus T. schismatorhynchus	Urin. bladder Gall-bladder	Chloromyxum granulosum	U. S. A.
Urophycis chuss	Gam-bladder	Ceratomyxa tylosuri acadiensis	Africa
Zoarces angularis	"	acadiensis acadiensis	Canada
Amphibia		acadiensis	1
Bufo lentiginosus	Kidney	Wardia ohlmacheri	U. S. A.
B. marinus L	Gall-bladder	Sphaeromyxa immersa	Brazil
Hyla ourea	Testis, ovary	Myxobolus hylae	Australia
Leptodactylus ocellatus	Gall-bladder	Sphaeromyxa immersa	Brazil
Molge cristata Laur.	Oan-Diaduci	Sphaerom yaa riiniici sa	Diazii
(Triton c.)	"	Chloromyxum caudaium	France
Proteus anguineus L	Kidney	Chloromyxum protei	Austria
Rana esculenta	"	?Wardia ohlmacheri	France?
R. temporaria (R. fusca)	"	?Wardia ohlmacheri	""
Reptilia			}
Emys orbicularis L.			
(Cistudo europaea)	Kidney	Myxidium danilewskyi	Russia,
	•		France

Host	Organ Infected	Myxosporidian	Locality
Lacerta sp Trionyx (Amyda) ganzeli-	Ovarian egg	Gen. et spec. incert.	Italy
Trionyx (Amyda) ganseti-	Kidney	M yxidium mackiei	India
T. spinifera	"	americanum	U. S. A.

C. DISTRIBUTION OF MYXOSPORIDIA IN THE ORGANS OF THE HOST

Altho some species are found in various organs of the host animal, the majority has one or two particular seats of infection. Among the various organs which become infected, the gall-bladder is most frequently infected. The kidney, branchia and urinary bladder have less chances of being parasitized. As to the infection of the reproductive organs of the host, little is known. The male reproductive organ becomes rarely infected, being reported only twice. The female reproductive organ, however, is more frequently infected. The infection of the next generation of the host animal thru the infected ovum which is known to occur in some Microsporidian parasites, has not been reported in Myxosporidia as yet.

LIST IV. ORGANS OF HOST INFECTED BY MYXOSPORIDIA

I. PISCES

1) Integument.—Sphaerospora sp. Southwell et Prashad (under the scales)

Myxobolus seni Southwell et Prashad (fin)

Myxobolus transovalis Gurley (under the scales)

Myxobolus unicapsulatus Gurley (head)

Myxobolus cycloides Gurley (opercle)

Myxobolus inaequalis Gurley (head)

Myzobolus sp. Gurley (opercle, head, fin)

Myxobolus squamae Keysselitz (inner surface of the scales)

Myxobolus volgensis Reuss (fin)

Myxobolus permagnus Wegener (operculum)

Myzobolus aureatus Ward (fin)

Henneguya brachyura Ward (fin-ray)

Henneguya linearis (Gurley) Labbé (membrane lining branchial cavity)

Henneguya gurleyi Kudo (base of spines of dorsal fin)

Henneguya strongylura (Gurley) Labbé (cephalic region)

Lentos pora dermatobia Ishii

2) Connective tissue.—Myxidium anguillae Ishii (subcutaneous)

Myzobolus fuhrmanni Auerbach (under the oral mucous membrane)

Myzobolus oviformis Thélohan (subcutaneous)

Myzobolus lintoni Gurley (subcutaneous)

Myxobolus oblongus Gurley (chiefly of the head)

Myxobolus gigas Auerbach (of operculum, sides and fins)

Myxobolus capsulatus Davis (visceral)

Henneguya kolesnikovi (Gurley) Labbé (interstitial)

Henneguya nüsslini Schuberg et Schröder (at the base of dorsal fin)

Henneguya miyairii Kudo (of the head)

Gen. et sp. incert. Linton (subcutaneous)

3) Muscle.—Leptotheca perlata (Gurley) Labbé

Chloromyxum quadratum Thélohan

Chloromyxum funduli Hahn

Chloromyxum clupeidae Hahn

Lentos pora multiplicata Reuss

Myxobolus notatus Mavor (connective tissue of voluntary muscle)

Myxobolus pfeifferi Thélohan

Myxobolus sandrae Reuss

Myzobolus musculi Keysselitz

Myxobolus funduli Kudo

Myxobolus pleuronectidae Hahn

Myzobolus sp. Southwell (subcutaneous intermuscular tissue)

Myxobolus nodularis Southwell et Prashad

Myxobolus orbiculatus Kudo

Henneguya creplini (Gurley) Labbé

Henneguya monura (Gurley) Labbé

Henneguya zschokkei (Gurley) Doflein

Henneguya salminicola Ward

Gen. et spec. incert. Leydig

4) Eye.—Sphaerospora platessae Woodcock (optic capsule)

Myxobolus oculi-leucisci Trojan (vitreous body)

Myxobolus ellipsoides Thélohan

Myxobolus mülleri Bütschli

Myxobolus aeglefini Auerbach

Myxobolus volgensis Reuss

Myxobolus magnus Awerinzew

Henneguya schisura (Gurley) Labbé (intercellular tissue of eye muscle)

5) Branchiae.—Sphaerospora carassii Kudo

Myzosoma dujardini Thélohan

Myxosoma (?) lobatum Nemeczek

Myxosoma funduli Kudo

Lentospora acuta (Fujita)

Myxobolus piriformis Thélohan

Myxobolus toyamai Kudo

Myxobolus rokitae Southwell et Prashad

Myzobolus dispar Thélohan

Myxobolus ellipsoides Thélohan

Myzobolus exiguus Thélohan

Myxobolus oviformis Thélohan

Myzobolus mülleri Bütschli

Myxobolus globosus Gurley

Myxobolus cycloides Gurley (also pseudobranchiae)

Myxobolus sphaeralis Gurley

Myxobolus anurus Cohn

Myxobolus sp. Gurley

Myxobolus gigas Auerbach

Myxobolus polgensis Reuss

Myxobolus scardinii Reuss

Myxobolus macrocapsularis Reuss

Myxobolus bramae Reuss

Myxobolus cyprinicola Reuss

Myxobolus balleri Reuss

Myxobolus sp. Miyairi

Myxobolus sp. Wegener

Myxobolus permagnus Wegener

Myzobolus rotundus Nemeczek

Myxobolus minutus Nemeczek

Myxobolus funduli Kudo

Myxobolus koi Kudo

Myxobolus discrepans Kudo

Henneguya psoros permica Thélohan

Henneguya texta (Cohn)

Henneguya minuta (Cohn)

Henneguya lobosa (Cohn)

Henneguya creplini (Gurley) Labbé

Henneguya linearis (Gurley) Labbé

Henneguva acerinae Schröder

Henneguya gigantea Nemeczek

Gen. et spec. incert. Heckel et Kner

6) Heart.—Myzobolus cordis Keysselitz (muscle of ventricle and bulbus arteriosus)

Gen. et spec. incert. Leydig (auriculo-ventricular valve)

7) Air bladder.—Myxobolus ellipsoides Thélohan (conn. tiss.)

Myxobolus mülleri Bütschli (conn. tiss.)

Myxobolus physophilus Reuss (surface)

Myzobolus permagnus Wegener

8) Body-cavity (cyst).-Myxobolus sp. Gurley

Myxobolus carassii Klokacewa

Gen. et spec. incert. Leydig

9) Nervous tissue.—Myzobolus neurobius Schuberg et Schröder

Lentospora encephalina Mulsow (blood vessel of brain)

10) Bone, cartilage, perichondrium.—Lentos pora cerebralis (Hoier) Plehn

Myzobolus aeglegini Auerbach Henneguya brachyura Ward (of fin)

11) Stomach, pyloric cecum.—Myzobolus exiguus Thélohan

Myzobolus mesentericus Kudo

Henneguya tenuis Vaney et Conte

12) Liver.-Myxobolus ellipsoides Thélohan

Myzobolus oviformis Thélohan

Myxobolus cyprini Doflein

Myxobolus cordis Keysselitz

Myxobolus musculi Keysselitz

Myzobolus carassii Klokacewa

Myxobolus mesentericus Kudo

13) Gall-bladder.—Leptotheca agilis Thélohan

Leptotheca elongata Thélohan

Leptotheca polymorpha (Thélohan) Labbé

Leptotheca parva Thélohan

Leptotheca hepseti Thélohan

Leptotheca sp. Awerinzew

Lepiotheca macrospora Auerbach

Leptotheca informis Auerbach

Leptotheca longipes Auerbach

Leptotheca fusiformis Davis Leptotheca scissura Davis Ceratomyxa arcuata Thélohan Ceratomyza sphaerulosa Thélohan Ceratomyxa pallida Thélohan Ceratomyxa globurifera Thélohan Ceratomyxa appendiculata Thélohan Ceratomyza truncata Thélohan Ceratomyxa reticularis Thélohan Ceratomyxa inaequalis Doflein Ceratomyxa linospora Doflein Ceratomyxa ramosa Awerinzew Ceratomyza drepanopsettae Awerinzew Ceratomyxa tylosuri Awerinzew Ceratomyxa (?) spari Awerinzew Ceratomyxa sp. (?). Awerinzew Ceratomyxa sp (?). Awerinzew Ceratomyxa acadiensis Mavor Ceratomyxa sp. Georgévitch Ceratomyxa coris Georgévitch Ceratomyxa herouardi Georgévitch Ceratomyxa mesospora Davis Ceratomyxa sphairophora Davis Ceratomyxa taenia Davis Ceratomyxa attenuata Davis Ceratomyxa recurvata Davis Ceratomyxa lunata Davis Ceratomyxa abbreviata Davis Ceratomyxa flagellifera Davis Ceratomyxa agglomerata Davis Ceratomyxa amorpha Davis Ceratomyxa monospora Davis Ceratomyxa streptos pora Davis Ceratomyxa aggregata Davis Ceratomyxa undulata Davis Chloromyxum leydigi Mingazzini Chloromyxum fluviatile Thélohan Chloromyxum truttae Léger (also gall-duct) Chloromyxum cristatum Léger Chloromyxum dubium Auerbach Chloromyxum sp. Awerinzew Chloromyxum thymalli Lebzelter Chloromyxum koi Fujita Chloromyxum magnum Awerinzew Chloromyxum misgurni Kudo Chloromyxum fujitai Kudo Chloromyxum trijugum Kudo Chloromyxum catostomi Kudo Chloromyxum wardi Kudo Sphaerospora masovica Cohn Myxidium incurvatum Thélohan Myxidium sphaericum Thélohan

Myxidium sp. Gurley (only in gall-duct)

Myxidium giganteum Doslein

Myxidium pfeifferi Auerbach

Myxidium inflatum Auerbach

Myxidium bergense Auerbach

Myzidium procerum Auerbach

Myxidium macrocapsulare Auerbach

Myxidium sp. Awerinzew

Myxidium oviforme Parisi

Myxidium sp. Mavor

Myxidium kagayamai Kudo

Myxidium gadi Georgévitch

Myxidium glutinosum Davis

Myxidium phyllium Davis

Myxidium striatum Cunha et Fonseca

Myxobolus misgurni Kudo (few spores only)

Myxobolus sp. Lebzelter (spores only)

Sphaeromyxa balbianii Thélohan

Sphaeromyxa incurvata Doflein

Sphaeromyza sabrasesi Laveran et Mesnil

Sphaeromyxa kellandi Auerbach

Sphaeromyza exneri Awerinzew

Sphaeromyza gasterostei Georgévitch

Zschokkella nova Klokacewa

Zschokkella acheilognathi Kudo (also in gall-duct)

Gen. incert. congri Perugia

Gen. incer. merlucii Perugia

Gen. et spec. incert. Mavor

14) Spleen.—Myxobolus piriformis Thélohan

Myzobolus sp. Kudo

Myxobolus ellipsoides Thélohan

Myxobolus exiguus Thélohan

Myxobolus oviformis Thélohan

Myxobolus pfeifferi Thélohan

Myxobolus cyprini Doflein

Myxobolus cordis Keysselitz

Myxobolus musculi Kevaselitz

Myxobolus mesentericus Kudo

15) Intestine.—Myxobolus exiguus Thélohan

Myxobolus mülleri Bütschli

Myxobolus pfeifferi Thélohan

Myxobolus miyairii Kudo

Myzobolus carassii Klokacewa

Myxobolus mesentericus Kudo

Henneguya peri-intestinalis Cépède

Henneguya tenuis Vaney et Conte

16) Ovary.-Wardia ovinocua Kudo

Sphaerospora elegans Thélohan

Myxidium histophilum Thélohan

Myzobolus pfeifferi Thélohan

Myxobolus mülleri Bütschli

Myxobolus musculi Keysselitz

Henneguya oviperda (Cohn) Henneguya media Thélohan Henneguya brevis Thélohan

17) Kidney.—a) Urinary tubules.—Leptotheca renicola Thélohan

Mitraspora caudata (Parisi) Kudo Mitraspora cyprini Fujita (also in ureter) Mitraspora elongata Kudo Sphaerospora elegans Thélohan Sphaerospora disergens Thélohan Henneguya media Thélohan

Henneguya brevis Thélohan Henneguya gasterostei Parisi

Hoferellus cyprini Doflein

b) Tissue.—Mitraspora elongata Kudo

Chloromyxum quadratum Thélohan

Sphaerospora rostrata Thélohan (Malpighian bodies)

Myzidium kistophilum Thélohan Lentospora asymmetrica Parisi

Myzobolus pfeifferi Thélohan

Myxobolus cyprini Doflein

Henneguya neapolitana Parisi (conn. tiss. of ren. tubules)

Hoferellus cyprini Doflein

c) Seat, unstated.—Chloromyxum mucronatum Gurley

Sphaerospora angulata Fujita Myzidium barbatulae Cépède Myzidium giardi Cépède Myzobolus piriformis Thélohan Myzobolus ellipsoides Thélohan Myzobolus exiguus Thélohan Myzobolus oviformis Thélohan

Myxobolus mülleri Bütəchli

Myxobolus obesus Gurley

Myzobolus cycloides Gurley Myzobolus cordis Keysselitz

Myzobolus musculi Keysselitz

18) Urinary bladder.—Leptotheca lobosa Davis

Leptotheca glomerosa Davis

Ceratomyxa navicularia Davis

Ceratomyxa spinosa Davis

Chloromyxum mucronatum Gurley

Chloromyxum granulosum Davis

Sphaerospora elegans Thélohan

Sphaerospora divergens Thélohan

Sphaerospora polymorpha Davis

Sphaerospora sp. Davis

Sinuolinea dimorpha Davis (also in ureter)

Sinuolinea capsularis Davis Sinuolinea arborescens Davis

Sinuolinea opacita Davis

Sinuolinea brachiophora Davis

Myxidium lieberkühni Bütschli

Zschokkella kildae Auerbach

Zschokkella globulosa Davis

Myzoprotous ambiguus (Thélohan) Doslein Myzoprotous cordiformis Davis Myzoprotous cornutus Davis Henneguya legeri Cépède Henneguya wisconsinensis Mavor et Strasser Henneguya miclospora Kudo

- Gen. et spec. incert. Mavor

 19) Testis.—Myzobolus pfeifferi Thélohan (only spores)
- 20) Mesentery. Myzobolus mesentericus Kudo
- 21) Seat unknown.—Henneguya sp. Gurley (integument?)
 Gen. et. spec. incert. Borne

II AMPHIBIA

- 1) Gall-bladder.—Chloromyzum caudatum Thélohan Séhaeromyza immersa (Lutz) Thélohan
- 2) Urinary tubules of kidney.—Wardia ohlmackeri (Gurley) Kudo Chloromyxum protei Joseph

								TA	BLE	I											
	Integument	Conn. tissue	Muscle	Eye	Gill	Heart	Air-bladd.	Nerv. tiss.	Bone, cartilage	Stomach wall	Liver	Gall-bladd.	Spleen	Intestine wall	Testis	Ovary	Kidney	Body cavit.	Urinary	Mesentery	Seat
Leptotheca	_		1	_	_	_	_			_		11		_	_		1(a)*		2		
Ceratomyxa			l									33	ļ						2		
Myxoproteus																			3		
Wardia																1	1(a)				
Mitraspora																	3(a)				
																	1(b)*				
Chloromyxum			3									14					1(a)				
																	1(b)				
																	1(c)*	1	2		
Sphaerospora	1			1	1					l		1				1	2(a)				
																	1(b)				
1																	1(c)	l	4		
Sinuolinea													l				2(a)		5		
Myxidium		1										18				1	1(b)				
							-										2(c)				
Autorio		11	130														3(a)		1	l	
Sphaeromyxa												7									
Zschokkella												2							2		
Myxosoma					3																
Lentospora	1		1		1			1	1								1(b)	1			
Myxobolus	10	6	9	6	28	1	4	1	1	2	7	2	10	6	2	4	2(b)				1
	-	1				Ī						-	7.		15	-	9(c)	1		1	1
Henneguya	4	3	4	1	8				1	1				2			3(a)		3		1
0 ,	1711								-								1(b)				
Hoferellus																	1(a)				
																	1(b)				
Total	16	10	10	-	41	1	_	2	3	3	-	-	10	8	2	10	39	3	24	1	2

^{*} For (a), (b) and (c), see page 42.

- 3) Testis, oviduct.—Myzobolus kylae Johnston et Bancroft
 - III REPTILIA
- Kidney (ren. tub.).—Myzidium danilewskyi Laveran Myzidium mackiei Bosanquet Myzidium americanum Kudo
- 2) Ovary.—Gen. et spec. incert. Mingazzini

IV INSECTA

1) Abdominal cavity.—Chloromyxum diploxys Gurley

V ANNELIDA

Myzobolus sp. Gurley

The data in this section are summarized on the preceding page (Table I).

D. THE EFFECT OF ENVIRONMENT ON THE ORGANAL DISTRIBUTION OF MYXOSPORIDIA IN HOSTS

Myxosporidia are almost equally distributed among marine and freshwater fishes in regard to the number of species. This is shown in the following table.

TABLE II

_		Number of species	Other hosts						
Genus	found in marine fish	found in fresh- water fish	Rept.	Amph.	Insect	Annelida			
Leptotheca (15)*	15			l					
Ceratomyza (35)			l						
Myxoproteus (3)									
Wardia (2)		1		1					
Mitraspora (3)		3	l						
Chloromyxum (22)		12	l	2	1	l			
Sphaerospora (10).		5							
Sinuolinea (5)			l						
Myxidium (26)		8	3						
	(2 common)	(2 common)							
Sphaeromyxa (7)	6		l·	1					
Zachokkella (4)		2							
Myxosoma (3)	l .	2							
Lentospora (6)		6							
	(2 common)	(2 common)							
Myxobolus (63)	5	56		1		1			
Henneguya (32)		31				.			
Hoferellus (1)		1							
Gen. et spec.	}	ĺ	l						
incert (12)	4	7	1		••••				
TOTAL 237+12	104+4	134+7	4	5	1	1			

^{*} The number in parenthesis denotes the number of species in the corresponding genus.

These genera have certain relations to the organal distribution in the body of the host, which are shown in List IV (page 37) and Table I (on page 43) and which can be put together as follows:

TABLE III

Genus	Number of species found in body- cavity	Number of species found in tissue	Number of species found in both	Seat unknown
Leptotheca (15)	14	1		••
Ceratomyxa (35)				
Myxoproteus (3)	3	<u>.</u>		••
Wardia (2)	1	1		
Mitraspora (3)	2		1 1	••
Chloromyxum (22)		4		
Sphaerospora (10)	4	4	1	1
Sinuolinea (5)				
Myxidium (26)	22	4		
Sphaeromyxa (7)				1
Zschokkella (4)				••
Myxosoma (3)	l	3		••
Lentospora (6)		5		••
Myxobolus (63)		59		2
Henneguya (32)		28		••
Hoferellus (1)		l	1	••
Gen. et spec. inct (12)	1	5		3
TOTAL 237+12	121+4	109+5	3	4+3

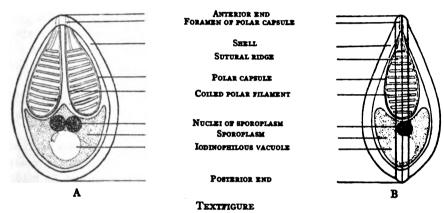
From the facts shown in the above tables, the following conclusions can be drawn.

- 1) The genera Leptotheca (one species in tissue), Ceratomyxa, Myxoproteus, Sinuolinea and Sphaeromyxa (one species in Amphibia) include parasites from the body cavity of marine fish.
- 2) The majority of the genera Lentospora, Myxosoma, Myxobolus (one species in Amphibia), Henneguya and Hoferellus include parasites in tissues of fresh-water fish.
- 3) The genera Chloromyxum, Sphaerospora, Myxidium and Zschokkella include forms that infect the body-cavity as well as the tissue of marine and fresh-water fishes.
- 4) The genus Mitraspora includes three species that parasitize the fresh-water fish.
- 5) The genus Myxidium has three species found in the kidney of reptiles.

- 6) The genera Wardia, Chloromyxum, Sphaeromyxa and Myxobolus include species parasitic in Amphibia.
 - 7) The genus Chloromyxum has one species found in an insect.
- 8) The genus Myxobolus has one species found in an annelid, which was not normally recorded.

THE SPORE

As will be shown later (page 52-55), the spore stage is still the only constant character by which various forms of Myxosporidia are identified from each other. For this reason it is necessary to have a clear conception of the form and structure of the spore and at the same time to define the terms used in the present paper, even tho they have commonly been used heretofore.



DIAGRAMMATICAL FRONT [A] AND SIDE [B] VIEWS OF A MYXOBOLUS SPORE.

(For further explanation see following pages.)

The spore of Myxosporidia is covered by a shell, which is composed of two valves usually symmetrical in form and size that come in contact in the sutural plane. The sutural line is straight in most cases, tho sometimes curved like an S. It is more or less thickened, forming the sutural ridge. The sutural ridge is to be made out clearly in fresh as well as in stained preparations and furnishes important data in regard to the classification of the parasite. The thickness of the shell-valve is usually uniform; in some species (Myxobolus), however, it may differ slightly in different parts of the shell. Besides, in many species of Myxobolus, the shell differentiates a small triangular intercapsular appendix on the inside at the anterior end directed posteriad between two polar capsules.

The form of the spore varies greatly owing to the shape of the shell together with its variously developed appendages; 1) lateral appendages as in Ceratomyxa, 2) anterior processes as in Myxoproteus, 3) posterior processes as in Wardia (fringe-like), Mitraspora (filiform), Hoferellus (spinous), Henneguya (tail-form), etc.

The surface of the shell may be smooth or exhibit various markings. More or less conspicuous ridges varying in form and number in different species, may run parallel to the sutural line, may show a network-like structure or may exhibit short tooth-like processes arising from the sutural ridge and radiating toward the center of each valve. When the ridges are fine, they form delicate striations, arranged usually parallel to the sutural line. Tho these markings are usually easily seen in vivo, they are very often more readily studied in stained preparations.

Inside of the shell are present the polar capsules and sporoplasm. Gurley (1894:120) and Davis (1917:210) used the term "capsules" instead of polar capsules because of the facts that "the situation implied by the the latter (polar capsule) is not constant" (Gurley) and that "they are often not in the position indicated by the term polar capsule" (Davis). The present writer, however, does not agree with these authors and retains the commonly used term, polar capsule, thruout the present paper on the basis of the fact that these polar capsules are situated at or near the more or less attenuated anterior end in the great majority of species or at each end (in Myxidiidae) of the spore, except in the few cases as in Wardia in which they are situated in the central portion and have the foramina at the anterior end of the spore.

The polar capsules may be pyriform or spherical. They are located at or near one end (anterior end) of the spore. In Myxidiidae, one polar capsule is situated at each end, in which case no distinction can be made between the anterior and posterior ends. The end or side opposite to the anterior, is the posterior end of the spore. The number of polar capsules in a spore varies according to the different genera. There is only one polar capsule in the spore of unicapsular Myxobolus, four in Chloromyxum, two in all the other genera. They may be equal or unequal in form and size. When two polar capsules are located at the anterior end, they may be convergent or divergent. Each has a foramen to the outside of the spore thru the shell in or near the sutural line, thru which the polar filament is extruded. The foramen is observable in the fresh condition. Staining will very often show clearly the canals thru the shell. Each polar capsule has an independent foramen.

In the polar capsule exists a coiled polar filament, which in most cases can be recognized without difficulty in the fresh condition. The polar filament is as a rule a more or less extended, probably hollow thread connected with the polar capsule, which is extruded from the spore thru the foramen under the action of the stimulants such as the digestive fluid of the host or certain chemicals. In Sphaeromyxa it is rather short and thick, tapering to a point. The polar filament is coiled around the longest axis of the polar capsule, except in Sphaeromyxa in which it is coiled around an axis perpendicular to the longest axis of the polar capsule.



The sporoplasm occupies the extracapsular cavity at the posterior region of the spore. It is of granular structure with almost always two nuclei. Besides, it has an iodinophilous vacuole mostly round or oval in the spores of the family Myxobolidae. It occurs thruout the spore stage and is the important character of the said family. The contents of the vacuole is probably of glycogenous nature and is stained deeply with iodine. Small refringent fat globules have also been observed in the spore.

Davis (1917:212) proposed to use capsular and postcapsular sides in place of anterior and posterior ends which have most frequently been used and are also used in the present paper. The latter terms can be employed as properly as Davis' terms except in the case of the Myxidiidae, where both terms, strictly speaking, are inapplicable.

The various abnormal spores are very often encountered in several species, the majority of the spores are of typical form, structure and size. In Myxosoma and Myxobolus, the spore sometimes develops a short posterior process, which is highly developed in the spore of the genus Henneguya.

Young spores, generally speaking, are more rounded in form than the mature form, while the mature spores, as a rule, are of definite form, structure and size characteristic to the species. It should, however, be kept in mind that there is a certain amount of variation among these characters.

As is generally recognized, one must mention whether the spores were measured in fresh condition or in fixed and stained state. The fresh spore is generally more or less larger than the mounted one.



DEFINITION OF TERMS USED FOR DESCRIPTIONS

- Anterior end.—The end of the spore where the polar capsules open; in most cases the polar capsules are situated at this end.
- Anterior process.—The spinous process of the shell at the anterior end of the spore of the genus Myxoproteus.
- Breadth.—The larger diameter of the spore measured at right angles to the length or sutural diameter; the shorter diameter thus measured being the thickness.
- Capsulogenous cell.—A small island of protoplasm with a nucleus, in which polar capsule becomes differentiated.
- Cyst.—The vegetative form of more or less conspicuous size in tissues of the host, surrounded usually by a membranous structure composed of the host issue.
- Disporous.—The character of a trophozoite of forming only two spores.
- Foramen.—Opening of the polar capsule thru which the polar filament is extruded.
- Front view.—The view in which length and breadth of the spore are laid horizontally.
- Gemmules.—A small mass of trophozoite separated from the mother body by plasmotomy. Used by Davis (1917). (See page 105.)
- Iodinophilous vacuole.—The vacuole in the sporoplasm of the spore of the family Myxobolidae, the contents of which are stained brownish with iodine.
- Lateral process.—The lateral prolongation of the shell-valve at right angles to the sutural plane.
- Length.—Antero-posterior diameter of the spore in the sutural plane; equivalent to sutural diameter.
- Longitudinal striations.—Fine ridges or thickenings marked longitudinally on the shell of the spore.
- Mesoplasm.—An intermediate layer between ectoplasm and endoplasm, coined by Cohn in the case of Myxidium lieberkühni (see page 107).
- Mictosporous.—The character of the trophozoite of forming a variable number of spores in an individual.
- Monosporous.—The character of the trophozoite of forming a single spore.

 Pans poroblast —Coined by Gurley (1893:408) used here in the same mean-
- Pans poroblast.—Coined by Gurley (1893:408) used here in the same meaning, an enclosed area in the endoplasm of the vegetative form, in which two sporoblasts become differentiated.

Plasmogamy. Fusion of two trophozoites, coined by Doflein (1898).

Plasmotomy.—Division of trophozoite into daughter individuals, coined by Doflein (1898).

Polar capsule.—The pyriform or spherical, hollow body in the spore which forms a polar filament.

Polar filament.—The filament which is coiled inside the polar capsule.

Polysporous.—The character of the trophozoite of forming spores, more than two.

Posterior filament.—Fine posterior appendage of the spore.

Posterior processes.—Posterior differentiations of the shell.

Ridge.—Linear or network-like elevation of the shell of the spore.

Shell.—The envelope of the spore.

Shell-values.—Two valves which compose the shell of the spore.

Sporoplasm.—The protoplasmic mass found inside of the spore (amebula or sporozoite), usually situated in the posterior portion of the spore.

Sutural diameter.—Same as length.

Sutural edge.—The edge of the shell-valves cut by the sutural plane.

Sutural line.—The line on the shell of the spore marked by the sutural plane.

Sutural plane.—The plane on which two shell-valves meet together.

Sutural ridge.—The ridge marking the sutural line.

Tail.—The posterior prolongation of the valves from the median posterior end; it may be a single process or bifurcated.

Thickness.—See breadth.

Trophosoite.—The vegetative or multiplicative stage of a Myxosporidian.

Vegetative form.—Same as trophozoite.

CLASSIFICATION OF MYXOSPORIDIA

The classification of Myxosporidia, was first carried out by Thélohan as early as 1892, who considered rightly that the spore was the only reliable means for the purpose. In 1899 and 1901, Doflein introduced into the classification two Legions, Disporea and Polysporea, and a new family. This plan has generally been followed by various authors in dealing with these protozoa.*

The classification of the said author, however, no longer agrees with our present knowledge of the animals. In the first place, as was pointed out by some authors, for instance Davis (1917:217), it is far from being correct to divide the Myxosporidia into two Legions, Disporea and Polysporea, on the basis of the number of spores formed in each vegetative form, since this differs even in one and the same species as was observed by Léger, Auerbach, Awerinzew, Parisi, Georgévitch, Davis, Kudo and others (see Table IV on page 53).

Auerbach who had observed numerous interesting facts in this group, had adopted Doflein's classification in his splendid work (1910) by simply adding two genera, Zschokkella and Lentospora, to the family Myxidiidae. In the following year (1911), he tried a new classification, on the same basis as Doflein did, by introducing two new Legions besides these two already existing, and by discarding all the families. Thus:

I	Monosporea	a) Genus	Coccomyxa
II	Mictosporea	a) Genus	Zschokkella
	-	b) Genus	Myxoproteus
		c) Genus	Myxidium
		d) Genus	Sphaeromyza
		e) Genus	Chloromyzun
		f) Genus	Sphaerospora
Ш	Disporea	a) Genus	Ceratomyxa
	•	b) Genus	Leptotheca
IV	Polysporea	a) Genus	Myxosoma
	• •	b) Genus	Lentospora
		c) Genus	Myxobolus
		d) Genus	Henneguya
		e) Genus	Hoferellus

As will be distinctly seen from Table IV, the classification not only fails to improve Doflein's classification in bringing together the genera, Myxo-

^{*} Doflein still uses the same classification in his recent work (1916).

proteus, Myxidium and Sphaeromyxa into Mictosporea, and Lentospora and Henneguya into Polysporea, but increases the confusion concerning relationship among the genera.

TABLE IV

Genus	Mono- and di- sporous	Mono- and poly- sporous	Di- sporous	Mono-, di- and poly- sporous	Di- and poly- sporous	Poly- sporous	Unknown
Leptotheca							
15 species		••	8			••	7
Ceratomyxa						,	
35 species	3		23		4		5
Myxoproteus							
3 species	••		2				1
Wardia*							
2 species	••	••				1	1
Mitraspora*							
3 species		••			2	1	
Chloromyxum							
22 species	1	2		2	6	4	7
Sphaerospora							
10 species	1	••	2	1	2	2	2
Sinuolinea*							
5 species			2		2	1	
Myxidium							
26 species	1		2	2	2	9	10
Sphaeromyxa				94			
7 species			••			5	2
Zschokkella							ļ
4 species	1	••		1		1	1
Myxosoma							
3 species		••	••		••	3	
Lentospora							İ
6 species			2			2	2
Myxobolus							1
63 species						54	9
Henneguya							
32 species				1	1	25	5
Hoferellus							1
1 species				l		1	۱

^{*} These three genera are unknown to Auerbach, except two species which were formerly placed in Leptotheca and Sphaerospora.



Parisi (1912) followed Auerbach in his paper dealing with Myxosporidia from Italian waters. Poche (1913) put Auerbach's classification in better form as follows:

```
Order: Myxosporidia
   2 Superfamily Mictosporea
        2 Family Myxidiidae
                     2 Genus Zschokkella
                     3 Genus Myxoproteus
                     4 Genus Myxidium
                     5 Genus Sphaeromyza
                     6 Genus
                              Sphaerospora
        3 Family Chloromyxidae
                     7 Genus Chloromyxum
    3 Superfamily Disporea
        4 Family Ceratomyxidae
                     8 Genus Ceratomyza
                     9 Genus Leptotheca
    4 Superfamily Polysporea
        5 Family Myxosomatidae (Poche)
                    10 Genus Myxosoma
                    11 Genus Lentospora
        6 Family Myxobolidae
                    12 Genus Myxobolus
                    13 Genus Henneguya
                    14 Genus Hoferellus
```

For the same reason given in discussing Auerbach, this, however, is not conformable with the present state of knowledge regarding these protozoa.

It was not until 1917 that the classification of the Myxosporidia approached to a more natural state in the valuable work by Davis (1917: 219-221). He pointed out sharply the unsatisfactory features in Doflein's classification and proposed a different system as follows:

```
Order: Myxosporidia.

Suborder I Myxosporea Davis
Family 1 Ceratomyxidae
Genus 1 Leptotheca
Genus 2 Ceratomyxa
Family 2 Sphaerosporidae Davis
Genus 1 Myxoproteus
Genus 2 Sphaerospora
Genus 3 Sinuolinea
Family 3 Myxidiidae
Genus 1 Myxidium
Genus 2 Sphaeromyxa
Genus 2 Sphaeromyxa
Genus 3 Zschokkella
Family 4 Chloromyxidae
Genus 1 Chloromyxum
```

Suborder II Cystosporea Davis Family 1 Myxosomidae* Davis

> Genus 1 Myxosoma Genus 2 Lentospora

Family 2 Myxobolidae

Genus 1 Myxobolus Genus 2 Henneguya Genus 3 Hoferellus

Thus, Davis selected the form of the spore for the establishment of two suborders and further rearranged the genera into closer positions to show relationship to each other better than any one of the previous authors. He, however, named the suborders according to a secondary character, i.e., the seat of the parasites in the host. According to his definition the trophozoites of the species belonging to Myxosporea are "with few exceptions free living in the body-cavity," while those of Cystosporea "with few exceptions" are tissue parasites.

From TABLE III on page 45, are taken the following data regarding this point:

	Total number of species known	Number of species found in body cavity	Number of species found in tissue	Number of species found in both places	Seat unknown
Myxosporea Cystosporea	132 105	114 7	14 95	2 1	2 2

Thus it appears that the terms Myxosporea and Cystosporea do not seem to be properly used. These may be replaced by terms that denote the first and common character of the suborders.

The suggestions as to the adoption of other characters than the spore for the divisions of Myxosporidia, proposed by Awerinzew (1907:831; 1908:64), Auerbach (1910:161) and Davis (1917:217) can only be applied in the future. At the present time, the characters concerning the vegetative form do not appear to afford a better and more natural basis for the classification of Myxosporidia than those of the spore. Thus from the taxonomic point of view the present situation does not seem to be much improved as compared with that at the end of the last century.

The writer proposes in the following pages a new classification based on the characters of the spore.



^{*} Davis did not notice the establishment of the family Myxosomatidae by F. Poche (1913), including exactly the same genera. See page 54.

Order MYXOSPORIDIA Bütschli 1881 Suborder EURYSPOREA nom. nov.

Largest diameter of the spore at right angles to the sutural plane. One polar capsule on each side of the plane. Sporoplasm with no iodin-ophilous vacuole. Vegetative form found in body cavity (except 2 species). Great majority parasites of marine fish. Monosporous, disporous and polysporous.

Family CERATOMYXIDAE Doflein 1899
With the characters of the suborder.

Genus LEPTOTHECA Thélohan 1895

Shell-valves of spore hemispherical or shortly rounded. 15 species. Disporous (7 unknown). 14 species in body-cavity; 1 in tissue; all in marine fish. Type species: Leptotheca agilis Thélohan.

Genus CERATOMYXA Thélohan 1892

Shell-valves, conical and hollow, attached on the bases; free ends extended, tapering to more or less sharply pointed or rounded ends. Sporoplasm usually does not fill the cavity, but is located asymmetrically in it. 35 species. Disporous (23 species), monosporous and disporous (3 species), disporous and polysporous (4 species) and unknown (5 species). All (except 2 species in urinary bladder) in the gall-bladder of marine fish. Type species: Ceratomyxa arcusta Thélohan.

Genus MYXOPROTEUS Doflein 1898 emend. Davis 1917

Spores roughly pyramidal; with or without distinct processes from the base of the pyramid. 3 species. Disporous (one species unknown). All in urinary bladder of marine fish. Type species: Myxoproteus ambiguus (Thélohan) Doslein.

Genus WARDIA nov. gen.

Spore form of isosceles triangle with two convex sides. Oval in profile. Surface of shell with fine ridges which turn into fringe-like processes at the posterior end. The polar capsules, large and perfectly spherical, situated at the central portion of the spore, opening at the anterior tip. Two species. Polysporous (one species unknown). Tissue parasite (one species) of fresh-water fish and amphibia, both found in Illinois, U. S. A. Type species: Wardia ovinocua nov. spec.

Genus MITRASPORA Fujita 1912 emend. Kudo

Spores spherical or ovoidal. Two polar capsules pyriform, one situated on each side of the sutural plane. Shell longitudinally striated; with or without long and fine filaments projecting posteriorly in a row at right

angles to the sutural plane at the posterior side. 3 species. Disporous and polysporous. All found in kidney of fresh-water fish. Type species: *Mitraspora cyprini* Fujita.

Suborder SPHAEROSPOREA nom. nov.

Spores spherical or subspherical, with two to four polar capsules. Sporoplasm without iodinophilous vacuole. Vegetative form found in body-cavity and tissue. Monosporous, disporous and polysporous. Parasites of marine and fresh-water fish and amphibia.

Family CHLOROMYXIDAE Thélohan 1892*

Spores with four polar capsules. Monosporous, disporous and polysporous.

Genus CHLOROMYXUM Mingazzini 1890

With the characters of the family. 22 species. 18 in body cavity; 4 in tissue. 7 from marine and 12 from fresh-water fish, 2 in amphibia, 1 in insect. Type species: Chloromyxum leydigi Mingazzini.

Family SPHAEROSPORIDAE Davis 1917

Spores with two polar capsules. Monosporous, disporous and polysporous.

Genus SPHAEROSPORA Thélohan 1892

Spores with two polar capsules. Monosporous, disporous and polysporous. 10 species. Body-cavity and tissue. 5 from fresh-water and 5 marine fish. Type species: Sphaerospora divergens Thélohan.

Genus SINUOLINEA Davis 1917

Spores with or without lateral processes. Two polar capsules spherical. Sutural line sinuous. 5 species. Disporous and polysporous. In the urinary bladder of marine fish. Type species: Sinuolinea dimorpha Davis.

Suborder PLATYSPOREA nom. nov.

Sutural plane of the spore coincides with or at an acute angle to the longest diameter. One or two polar capsules. Sporoplasm with or without an iodinophilous vacuole.

Family MYXIDIIDAE Thélohan 1892

Two polar capsules, one at each end. Sporoplasm without any iodinophilous vacuole. Spores fusiform.

Thélohan (1892) used the terms: Chloromyxidées, Myxididées, Myxobolidées, which Gurley (1893) made over into Chloromyxidae, Myxidiidae, Myxobolidae, so that the credit of recognizing and establishing these families belongs to Thélohan.

Genus MYXIDIUM Bütschli 1882

Spores more or less regularly fusiform, with pointed or rounded ends. Polar filaments long and fine. 26 species. Monosporous, disporous and polysporous. 22 in body-cavity; 4 in tissue. 15 in marine and 6 in freshwater fish, 2 in fishes from both waters and 3 in reptilia. Type species: Myxidium lieberkühni Bütschli.

Genus SPHAEROMYXA Thélohan 1892

Spores fusiform, with truncated ends. Polar filament short and thick. Trophozoites large and disc shaped. 7 species. Polysporous (2 unknown). 6 in body-cavity; 1 unknown. 6 in marine fish; 1 in amphibia. Type species: Sphaeromyxa balbianii Thélohan.

Genus ZSCHOKKELLA Auerbach 1910

Spores, semicircular in front view; pointed at ends. Polar capsules large and spherical, opening on the flat edge near the tips. Sutural line usually curved in S-form. 4 species. Monosporous, disporous and polysporous. Body-cavity. 2 from marine and 2 from fresh-water fish. Type species: Zschokkella hildae Auerbach.

Family MYXOSOMATIDAE Poche 1913

Two polar capsules at the anterior end. Sporoplasm without iodinophilous vacuole.

Genus MYXOSOMA Thélohan 1892

Spores ovoidal, flattened and more or less elongated. 3 species. Polysporous. Tissue parasites. 2 in fresh-water and 1 in marine fish. Type species: Myxosoma dujardini Thélohan.

Genus LENTOSPORA Plelin 1905

Spores similar to Myxobolus in form. Sporoplasm without any iodinophilous vacuole. 6 species. Disporous and polysporous (2 unknown). 1 in marine and 3 in fresh-water fish, 2 from fishes in both waters. Type species: Lentospora cerebralis (Hofer) Plehn.

Family MYXOBOLIDAE Thélohan 1892

Spores with one or two polar capsules at the anterior end, with or without posterior processes. Sporoplasm with an iodinophilous vacuole. Majority polysporous in fresh-water fishes.

Genus MYXOBOLUS Bütschli 1882

Spores ovoidal or ellipsoidal; flattened. One or two polar capsules at the anterior end. Shell without posterior process. 63 species. Polysporous (9 species unknown). 59 species in tissue; 4 unknown. 5 in marine and 56 in fresh-water fish, 1 in annelid and 1 in amphibia. Type species: Myxobolus mülleri Bütschli.

Genus HENNEGUYA Thélohan 1892

Spores more or less globular or ovoidal. Two polar capsules at the anterior end. Posterior end of the shell-valves prolonged into more or less extended processes, which unite and form a tail in the median line. 32 species. Polysporous, disporous and monosporous. 28 species in tissue and 4 in body-cavity. In fresh-water fish, except one. Type species: Henneguya psorospermica Thélohan.

Genus HOFERELLUS Berg 1898

Spores pyramidal, with two posterior processes from the lateral faces. 1 species. Polysporous. Tissue and body-cavity of fresh-water fish. Type and only species: *Hoferellus cyprini* Doflein.

DESCRIPTIONS OF GENERA AND SPECIES

Suborder EURYSPOREA nom. nov.

The definition of the suborder is recorded on page 56.

Family CERATOMYXIDAE Doflein

1899	Ceratomyxidea	Doflein	1899 : 378
1901	Ceratomyxidae	Doflein	1901 : 182

The characters of the family are described on page 56.

Genus LEPTOTHECA Thélohan

1895 Leptotheca	Thélohan	1895 : 331
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The characters of the genus are described on page 56.

Type species: Leptotheca agilis Thélohan.

LEPTOTHECA AGILIS Thélohan

[Figs. 1 to 5]

1892	Ceratomyxa agilis	Thélohan	1892:962
1895	Leptotheca agilis	Thélohan	1895:332
189 8	Leptotheca agilis	Doflein	1898: 294, 297

Habitat: Gall-bladder of Trygon pastinaca L. and Scorpaeona sp.; France, Rovigno, Napoli.

Vegetative form: Form generally elongated. Anterior end rounded where a mass of fat globules is found, while the posterior end terminates in a point. Size not exceeding 85μ by 20 to 25μ . The posterior part is sometimes divided into a certain number of lobes. In the protoplasm, the globules are clearly seen. Pseudopodia are localized at the anterior portion of the body. They are long, 40 to 50μ in length, filiform and very active in moving from back toward front, just like the motion of oars. Disporous.

Spore: Slightly elongated. Dimensions: sutural diameter 6 to 7μ , breadth 11 to 12μ .

LEPTOTHECA ELONGATA Thélohan

[Fig. 6]

1895	Lepioiheca elongaia	Thélohan	1895:332
1898	Leptotheca elongata	Doflein	1898:312
1917	Leptotheca elongata	Georgévitch	1917b : 99-106

Habitat: Gallbladder of Merluccius merluccius L. (M. vulgaris) and Motella tricirrata; Marseille, Banyuls, Le Croisic, Napoli, Monaco.

Vegetative form: Form variable. Many individuals show, however, a very characteristic form. It is elongated and has the length of about 120μ . The anterior end is enlarged into a disc-shaped depression, on the edge of which, the branched pseudopodia are formed. The body gradually narrows itself toward the posterior end. Also club-shaped, etc. The short lobose pseudopodia show no movement like that of oars.

Georgévitch's form: Young forms, oval or rounded, are attached to the epithelial cells of the bladder with a long filiform pseudopodium at the free end. Such forms often agglomerate in great number.

Spore: Form similar to the spore of Leptotheca agilis. Dimensions on the average: Sutural diameter 12 to 15μ , breadth 18 to 20μ .

LEPTOTHECA POLYMORPHA (Thélohan) Labbé

1895	Leptotheca elongata	Thélohan	1895 : 332-333
1899	Leptotheca polymorpha	Labbé	1899:88

Habitat: Gall-bladder of *Phycis mediterraneus* (P. phycis L.); Banyuls. Vegetative form: Form extremely polymorphous, with three main types. 1) Somewhat regularly club-shaped, with lobose pseudopodia, sometimes filiform at one end. 2) Irregular as is the case with *Ceratomyxa truncata*, with long (25μ) ectoplasmic processes, which are motionless or very slow in motion. Lobose pseudopodia are formed actively. 3) More or less rounded with bristle-like filose pseudopodia. Intermediary forms are also found. Often many individuals unite together. The protoplasm is much different from other forms, i.e., more homogeneous and compact. Granules are hardly visible on account of vacuolar appearance.

Spore: Dimensions: sutural diameter 10 to 12μ , breadth 18 to 20μ , length of polar filament 40μ .

LEPTOTHECA PARVA Thélohan

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1895	Lepiotheca parva	Thélohan	1895 : 333
1912	Lepiotheca parva	Auerbach	1912 : 42-43

Habitat: Gall-bladder of Scomber scombrus L.; Marseille, Le Croisic, Le Vivier-sur-mer, Kristiansund, Stavanger, Bergen.

Vegetative form: Form ordinarily rounded, spherical or subspherical. Often club-shaped. Size not larger than 12 to 15μ in diameter. Protoplasm finely granular. Pseudopodia lobose.

Spore: Small, more or less elongated, curved in arch-form. Dimensions: sutural diameter 3 to 4μ , breadth 8 to 10μ .

LEPTOTHECA RENICOLA Thélohan

1895 Leptotheca renicola Thélohan 1895 : 333

Habitat: Urinary tubules of the kidney of Scomber scombrus L.; Marseille, Le Croisic.

Vegetative form: Small. No marked character.

Spore: Globular. Form similar to the spore of Sphaerospora. Dimensions: sutural diameter 8μ , breadth 10μ .

LEPTOTHECA HEPSETI Thélohan

1895 Leptotheca kepseti

Thélohan 1895 : 334

Habitat: Gall-bladder of Atherina hepsetus L.; Marseille. Of rare occurrence; Thélohan observed it but once.

Vegetative form: Not described.

Spore: Form triangular with rounded angles. Dimensions: sutural diameter 7 to 8μ , breadth 12 to 15μ .

LEPTOTHECA PERLATA (Gurley) Labbé [Fig. 8]

1883		Balbiani 1883 : 201, 204
1894	Chloromyxum (Sphaerospora) perlatum	Gurley 1894: 272
1899	Leptotheca perlata	Labbé 1899 : 88

Habitat: Acerina cernua L.; France (?).

Vegetative form: Not described.

Spore: Elliptic. Two small polar capsules converging. Dimensions not given.

LEPTOTHECA sp. Awerinzew [Figs. 16, 17]

1908 Leptotheca sp.

Awerinzew 1908: 51, 52

Habitat: Gall-bladder of Sebastes norvegicus; Eastern Finmark? Vegetative form: Rounded form with clear differentiation of protoplasm into ectoplasm and endoplasm. Plasmotomy occurs.

Spore: Undescribed. No figure.

LEPTOTHECA MACROSPORA Auerbach [Fig. 9]

1909	Leptotheca macrospora	Auerbach	1909:70-71
1910	Leptotheca macrospora	Auerbach	1910ь : 768-769
1910	Leptotheca macrospora	Auerbach	1910c : 167
1912	Leptotheca macrospora	Auerbach	1912:42-43

Habitat: Gall-bladder of Sebastes viviparus H. Kr. and S. dactylopterus; Bergen, Kristiansund (May, September).

Vegetative form: Trophozoites spherical with the average diameter of 26 to 30μ . Homogeneous ectoplasm layer exhibits somewhat active ameboid movements. Endoplasm, granular in living specimen, is rather sharply distinguishable from the ectoplasm and contains large nuclei.

Spore: Size large. Form resembles to that of Leptotheca parva. Dimensions: sutural diameter and thickness 13μ , breadth 26μ . Polar capsules short oval, with a length of 5.2μ , length of polar filament about 130μ (KOH). In the second host, a few normal and numerous abnormal spores with three or four polar capsules were observed.

LEPTOTHECA INFORMIS Auerbach

[Fig. 10]

1910 Leptotheca informis Auerbach 1910b: 770-771 1912 Leptotheca informis Auerbach 1912: 42-44

Habitat: Gall-bladder of *Molva vulgaris* Flem. and *Gadus merlangus*; Bergen, Tjomo.

Vegetative form: Young trophozoites with somewhat long and narrow pseudopodia formed of hyaline ectoplasm; movements active. The protoplasm is differentiated into ectoplasm and endoplasm. When stained, two large (7 to 9μ) and two small (3 to 4μ) nuclei were observed in an individual, 27μ long excluding the pseudopodia. Sporulating trophozoites are generally round and each forms two spores, which are developed independently to each other (i.e., not in ordinary pansporoblast). Auerbach observed centrosomes in the nuclei of larger type in division. Disporous.

Spore: Large and heavily built. Greatly curved. Sutural line fairly well marked. Polar capsules round. Dimensions: sutural diameter 10μ , breadth 18 to 20μ , thickness 9μ , diameter of polar capsules 3 to 4μ . Sporoplasm contains two nuclei, 3.5 to 4μ in diameter.

LEPTOTHECA LONGIPES Auerbach

[Fig. 11]

1910 Leptotheca longipes Auerbach 1910b: 771 1912 Leptotheca longipes Auerbach 1912: 42-43

Habitat: Gall-bladder of Brosmius brosme Asc.; Bergen (May).

Vegetative form: Trophozoites elongated or rounded. Only few pseudopodia which are very long. Small forms with a very long process, were observed in large numbers; length of the body being 10μ , while the process was 60μ long. Endoplasm contains nuclei of various sizes. Disporous.

Spore: Form similar to that of *Leptotheca informis*, though smaller. Dimensions: sutural diameter 8 to 9μ , breadth 12 to 14μ , thickness 8μ , diameter of polar capsule 2.5μ .

LEPTOTHECA FUSIFORMIS Davis

[Fig. 12]

1917 Leptotheca fusiformis Davis 1917: 222

Habitat: Gall-bladder of Cestracion zygaena; Beaufort (July).

Vegetative form: Pyriform, tapering gradually toward the posterior end, which usually terminates in a long, slender process; colorless and transparent. Progressive movements rapid. Endoplasm granular, the granules being more abundant at the anterior end. The average size of full-grown individuals: 50μ by 13μ . Disporous.

Spore: Elliptical in front view; fusiform in side view. Sutural plane slightly oblique to the longest diameter, the line forming a marked ridge. Polar capsules open on opposite sides of the spore. Sporoplasm finely granular, confined to the central part of spore. Dimensions: sutural diameter 9μ , breadth 16μ , polar capsule 4.5μ long, length of polar filament 30μ .

LEPTOTHECA SCISSURA Davis

[Fig. 13]

1917 Leptotheca scissura

Davis

1917:222

Habitat: Gall-bladder of Dasybatis hastatus and Pteroplatea maclura Le Sue; Beaufort (July, August).

Vegetative form: Young form elongated, with long attenuated posterior process; usually slightly constricted just posterior to rounded anterior end, which bears numerous long, filiform pseudopodia. Progressive movement rapid. Ectoplasm distinguishable at the anterior end. Endoplasm usually filled with small, clear, colorless spherules, which become larger and yellowish as the body increases in size. Each spherule contains one to several dark-brown granules, which increase in size and number and finally collect in an irregular clump at the centre of spherule. The larger individuals are usually flattened dorso-ventrally. The posterior end is divided into long slender processes, presenting sometimes a network caused by the fusion of two or more adjacent processes. Full-grown forms: length 125 to 150μ , breadth 20 to 25μ . The longest observed, 195μ by 16μ . Disporous.

Spore: Elliptical in front view; somewhat flattened along the posterior side. Sutural line distinct and at right angles to the longest diameter. Polar capsules have foramina at some distance from the capsular margin. Sporoplasm finely granular, nearly filling both valves. Dimensions: sutural diameter 11μ , breadth 22μ , diameter of polar capsule 4μ .

LEPTOTHECA LOBOSA Davis

[Fig. 14]

1917 Leptotheca lobosa

Davis

1917:223

Habitat: Urinary bladder of Paralichthys dentatus L.; Beaufort (July). Vegetative form: Usually spherical which may form a large rounded pseudopodium composed of ectoplasm. Body colorless and transparent to translucent. Ameboid movements very slow. Ectoplasm contains coarse granules, which are of uniform size and very distinct. Endoplasm

less granular and more transparent than ectoplasm, containing numerous large, yellow, fat globules, which are abundant in large forms. Diameter up to 24μ . Disporous.

Spore: Elliptical in front view; valves slightly tapering but rarely alike. Sutural line forming a sinuous ridge. Polar capsules open at some distance from the anterior margin. Sporoplasm nearly filling both valves. Free spores are often seen to remain united at the sutural line. Dimensions: sutural diameter 9 to 10μ , breadth 16 to 18μ ; diameter of polar capsule 3μ .

LEPTOTHECA GLOMEROSA Davis

[Fig. 15]

1917 Leptotheca glomerosa

Davis 1917: 223

Habitat: Urinary bladder of Paralichthys albiguttus; Beaufort.

Vegetative form: Rounded or somewhat irregular in shape, with short lobose pseudopodia. Body transparent and colorless. Ameboid movements slow. Ectoplasm hyaline, forming a distinct outer layer. Endoplasm finely granular, with numerous small fat globules varying in size. Almost entire body is used for spore formation. Diameter of rounded sporulating trophozoite about 11 μ . Disporous.

Spore: Approximately cylindrical; valves rounded at ends. The coiled polar filament not visible in the polar capsule. Sutural line at right angles to the longest diameter. Sporoplasm finely granular, fills the extracapsular cavity of spore. Dimensions: sutural diameter 4.5μ , breadth 9μ , diameter of polar capsule 2μ .

Genus CERATOMYXA Thélohan

1892	Ceralomyxa	Thélohan	1892 : 169, 171, 175
1805	Ceratomora	Thélohen	1805 - 334

The characters of the genus described on page 56.

Type species: Ceratomyxa arcuata Thélohan.

CERATOMYXA ARCUATA Thélohan

[Figs. 18 to 22]

1892	Ceratomyxa arcuata	Thélohan	1892a : 1091
1895	Ceratomyxa arcuata	Thélohan	1895 : 335-336
1899	Ceratomyxa arcuata	Labbé	1899:90
1912	Ceratomyxa arcuata	Parisi	1912 : 290-291
1913	Ceratomyxa arcuata	Jameson	1913 : 2
1916	Ceratomyxa arcuata	Georgévitch	1916a: 3

Habitat: Gall-bladder of Pagellus centrodontus C. et V., Crenilabrus melops L., Motella tricirrata Bl., Ophidium vasalli, Gobius paganellus L., Heliases chromis Gthr.; Scorpaena scrofa L., S. porcus L.; France (Marseille, Banyuls, Concarneau, Roscoff), Italy (Napoli, summer), Monaco (May).

Vegetative form: Polymorphous; generally club-shape, pseudopodia localized at the broad end; the other end cylindrical or terminating in a sharp point. Some other different forms. Pseudopodia, always localized, lobose pointed at the extremities. Ectoplasm hyaline and thin. Endoplasm contains fat globules and particular elements, mostly large refractive globules, which seem to disappear in the sporulating individuals. Dimensions (maximum): length 35 to 40μ , breadth 12 to 15μ , pseudopodia about 10μ long. Disporous.

Spore: Arch form. Shell valves equal. Sporoplasm occupies the extracapsular cavity of the spore. The length varies rather considerably according to the development of the lateral processes, which are occasionally acuminated or very short. Often extremities are rounded. Dimensions (Thélohan); breadth 20 to 30μ , sutural diameter 5 to 8μ . Parisi's measurements: breadth 25 to 31μ , sutural diameter 5.5 to 6μ , length of polar capsules 3.5 to 4μ , length of polar filaments 25μ .

Remarks: The writer agrees with Parisi in eliminating Labbé's two subspecies (1899:90), as they are too arbitrary.

CERATOMYXA SPHAERULOSA Thélohan

[Figs. 23 to 24]

1892	Ceratomyxa sphaerulosa	Thélohan	1892:171
1895	Ceratomyxa sphaerulosa	Thélohan	1895 : 334-335
1909	? Ceratomyxa sphaerulosa	Auerbach	1909:80
1912	Ceratomyxa sphaerulosa	Auerbach	1912:4,45
1916	Ceratomyxa sphaerulosa	Georgévitch	1916a:3

Habitat: Gall-bladder of Musiclus canis Mitch. (M. vulgaris), Galeus galeus L. (G. canis), Clupea harengus, Scullium canicula Cuv.; St-Valery-en-Caux, Roscoff, Bergen, Monaco (May).

Vegetative form: Form more or less definite among the adults. Generally elongated. Both ends slightly attenuated. Wide in the middle part of the body. Lobose pseudopodia at one of the extremities. Others more massive or more or less regularly spherical, in which case the pseudopodia are formed from the whole surface. Spherical form does not exceed 50 to 60μ in diameter. Other forms 90 to 100μ by 30 to 40μ (largest). Young forms colorless and are more variable than the adults. Protoplasm homogeneous and finely granular. Adult form, on the contrary, yellowish or greenish yellow. The endoplasm is filled with small (5μ in diameter) spheres, in the centre of which 5 to 6 small granules, yellowish brown or greenish in color, are present. Disporous.

Spore: Remarkably large. Polar filament can be seen in vivo (easily extruded by KOH, ether, etc.) Sporoplasm occupies one of the shell-valves, while a small mass of very pale looking substance is seen in the other. Dimensions: sutural diameter 10 to 12μ , breadth 90 to 100μ , subspherical polar capsule 6 to 7 by 5μ , sporoplasm 12 to 15 by 8 to 9μ .

CERATOMYXA PALLIDA Thélohan

1895	Ceratomyxa pallida	Thélohan	1895 : 336-337
1898	Ceratomyxa pallida	Doflein	1898:341
1916	Ceratomyxa pallida	Georgévitch	1916b : 2, 3

Habitat: Gall-bladder of Box boops L. and B. salpa L.; Marseille, Villefranche, Rovigno, Monaco (May).

Vegetative form: Ordinarily spherical not exceeding 16 to 20μ in diameter. Many individuals often found in massive groups. Pseudopodia lobose and mostly short. Protoplasm extremely pale with fine granules.

Spore: Dimensions: sutural diameter 5μ , breadth 25 to 30μ .

CERATOMYXA GLOBULIFERA Thélohan [Fig. 25]

1895 Ceratomyza globulifera Thélohan 1895 : 338

Habitat: Gall-bladder of Merluccius merluccius L. (M. vulgaris); Marseille, Banyuls.

Vegetative form: Polymorphous. Elongated into long branches, including endoplasm. Endoplasm contains small refractive globules.

Spore: Elongated. Shell-valves unequal, one being longer and finer than the other. Dimensions: sutural diameter 10μ , breadth 50μ .

CERATOMYXA APPENDICULATA Thélohan [Fig. 26]

1892	Ceratomyxa appendiculata	Thélohan	1892a: 963-964
1895	Ceratomyxa appendiculata	Thélohan	1895:337
1898	Ceratomyza appendiculata	Doflein	1898 : 300, 311

Habitat: Gall-bladder of Lophius piscatorius L., L. budegassa Spin.; Roscoff, Le Croisic, Marseille, Banyuls, Napoli, Rovigno.

Vegetative form: Extremely polymorphous. Young form spherical, spatulaform, club-shape, etc. In adult form, the main thick part of the body, in which spore formation takes place, forms 1 to 6 long prolongations, twice or three times longer than the main part of the body. Pseudopodia lobose, filiform or elongated with enlargements. Disporous.

Spore: Lateral prolongations of shell-valves well developed. Dimensions: sutural diameter 5 to 7μ , breadth 50μ .

CERATOMYXA TRUNCATA Thélohan [Fig. 27]

	<u> </u>	-	
1895	Ceratomyxa truncata	Thélohan	1895:336
1912	Ceratomyza truncata	Parisi	1912:289-290

Habitat: Gall-bladder of Clupea pilchardus Walb. (Alosa sardina); Marseille, Villefranche, Napoli (August, September).

Vegetative form: Polymorphous. Ordinarily more or less rounded, with lobose pseudopodia. Pseudopodia long and often shows very active movements. Endoplasm very finely granular, contains small fat globules which are found in irregular mass or in a circular form. Disporous.

Spore: Valves are short and truncate. Sporoplasm occupies the whole cavity. Spores with three valves are frequently encountered. Dimensions: breadth 25μ , 5μ in sutural diameter. According to Parisi, spores with two shell-valves are rather few (10%), while those with three (70%) and four shell-valves (20%) prevail in number! Dimensions: breadth 20 to 30μ , length of the polar filament 45μ .

CERATOMYXA RETICULARIS Thélohan [Fig. 28]

1895 Ceratomyza reticularis Thélohan 1895: 337-338

Habitat: Gall-bladder of Trachinus draco L.; Banyuls.

Vegetative form: Extremely polymorphous. Generally spherical or club-shaped. Well developed trophozoites have the similar form as in *C. appendiculata*. Endoplasm highly reticular, with refringent fluid.

Spore: Shell valves are short and truncate, one of which is curved to the rear. Dimensions: sutural diameter 12 to 15μ , breadth 45 to 50μ .

CERATOMYXA INAEQUALIS Doflein [Fig. 29]

1898 Ceratomyza inaequalis Doflein 1898: 284-285

Habitat: Gall-bladder of Crenilabrus mediterraneus and C. pavo; Napoli.

Vegetative form: Form usually club-shaped. Protoplasm in active motion, is differentiated distinctly into ectoplasm and endoplasm. Body yellowish brown by the presence of granules in endoplasm. Inactive formation of pseudopodia. Ameboid movements or progressive movements by means of the posterior process. Size: 20 to 40μ by 5 to 10μ in average. Length of the posterior process up to 30μ . After spore formation, only two nuclei remain in protoplasm, which apparently degenerate later. Disporous.

Spore: Elliptical, somewhat flattened. Massive. Very transparent. Both ends round, but unequally built, i.e., one end is club-shaped. Polar capsules are somewhat round and are bound to the shell by protoplasmic bridges. The polar filament is not seen in fresh spores. Dimensions: sutural diameter 6μ , breadth 31μ , diameter of the polar capsule 2.5 to 3μ , length of polar filament is half breadth of the spore (diluted nitric acid).

CERATOMYXA LINOSPORA Doflein

[Figs. 30 to 31]

1898 Ceratomyxa linos pora

Doflein

1898:285

Habitat: Gall-bladder of Labrus turdus; Napoli.

Vegetative form: Club- or spindleshape. Protoplasm highly granulated. Body whitish grey, though very transparent. Pseudopodia very fine and only formed at the anterior end of the body. Size: 30 to 35μ by 16 to 18μ . Disporous.

Spore: Form symmetrical with long thread-like lateral processes. In sporoblast, the processes are wound around the spore. It is twice as long as the breadth of the spore. Polar capsules large and spherical pyriform. Dimensions: total breadth 50μ , breadth of the main part of the spore 10 to 12μ , sutural diameter 5μ , length of lateral process 20μ . "Polar filament was too fine to be measured."

CERATOMYXA RAMOSA Awerinzew

[Figs. 32 and 33]

1907 Ceratomyxa ramosa Awerinzew 1907: 831–834
*1908 Ceratomyxa ramosa Awerinzew 1908: 60–66

Habitat: Gall-bladder of Hippoglossus vulgaris Flemm.; Kjellebjord, Murman coast.

Vegetative form: Form irregular ameboid, owing to the presence of peculiar pseudopodia. The middle part of the body is enlarged into an ellipsoidal form, where nuclei and sporoblasts are present. From this part two, rarely one or three processes are formed, which branch out several pseudopodia of different length. The finer portions of pseudopodia anastomose each other and form a characteristic and remarkable network. Differentiation of protoplasm is not very distinct. Ectoplasm is not well developed, the covering the entire surface of the body as a thin layer. Endoplasm slightly vacuolated and granular, forms the greater part of the body. Disporous and polysporous.

Spore: Form and size (?) resemble C. arcuata. Slightly curved toward the posterior side. Valves usually unequally built, one being longer than the other. Sporoplasm almost always asymmetrically situated in the shell. Polar capsules on each side of the sutural plane and of the plane perpendicular to the sutural plane, cutting the spore into two equal parts. Young spores in development ellipsoidal to kidney bean shape. Dimensions: sutural diameter 12 to 20μ , breadth 50 to 80μ .

^{*}Professor J. Zeitlin has kindly translated some part of the paper, for which the writer expresses his thanks.

CERATOMYXA DREPANOPSETTAE Awerinzew [Figs. 34 to 39]

1907	Ceratomyxa sp.	Awerinzew	1907:832-833
1908	Ceratomyxa drepanopsettae	Awerinzew	1908 : 1-41, 45-47
1909	Ceratomyxa drepanopsettae	Awerinzew	1909 : 74-112
1912	Ceralomyxa drepanopsettae	Auerbach	1912 : 44-45
1918	Ceratomyxa drepanopsettae	Kudo	1918 : 14-15

Habitat: Gall-bladder of Pleuronectes platessa, P. flesus, Drepanopsetta plattessoides, Hippoglossus vulgaris, Hippoglossoides limandoides and Paralichthys dentatus; Murmankuste, Kabelvaag, Rorvik, Tjomo, Woods Hole (August, September).

Vegetative form: Polymorphous. Usually very much elongated and slender forms. Protoplasm differentiated. Endoplasm coarsely granular. Pseudopodia lobose and filiform (2 to 3μ), with which the trophozoites attach themselves to the epithelium of the bladder. Disporous.

Spore: Curved toward the posterior side. Shell with rounded ends. Valves almost always unequally built. Dimensions: breadth 50 to 80μ . Auerbach's form: Form variable. Size: sutural diameter about 12 to 14μ , breadth about 56μ , diameter of polar capsule about 4 to 6μ , length of the cavity in which the sporoplasm is located about 34μ . Kudo's form: Variable. Sutural diameter 8 to 10μ , average breadth 64μ , diameter of polar capsule 6μ .

CERATOMYXA TYLOSURI Awerinzew [Figs. 40 and 41]

1913 Ceratomyxa tylosuri Awerinzew 1913a: 153

Habitat: Gall-bladder of Tylosurus schismatorhynchus; Lorenço Marques, Delagoa Bay (Africa).

Vegetative form: Large, irregular, disc-like or large ameboid, with blunt lobose pseudopodia and highly granular protoplasm.

Spore: Large. The anterior edge arch-shape, while the posterior edge has two small horns which are located symmetrically to the sutural line. Polar capsules elongated and are separated from binuclear sporoplasm by a special membrane. Rarely spore with three polar capsules. Dimensions breadth 124 to 140μ , sutural diameter 40 to 45μ , thickness 25 to 30μ .

CERATOMYXA (?) SPARI Awerinzew [Figs. 42 and 43]

1913 Ceratomyxa (?) spari Awerinzew 1913a : 153-154

Habitat: Gall-bladder of Sparus berda; Lorenço Marques, Delagoa Bay (Africa).

Vegetative form: Large (100 to 120μ), disc-form ameboid, containing a large number of enclosures and granules of different size. In one case, a number of this form, carrying no spore, underwent budding, which resulted in forming spherical forms of various size, some of which divided again into 2 to 6 parts (Plasmotomy?). Monosporous and disporous.

Spore: More or less curved. Two polar capsules lie closely together on each side of the sutural plane. Ends of shell-valves are rounded. Dimensions: breadth 50 to 60μ , sutural diameter 12 to 15μ , thickness 12 to 15μ , polar filament very long (length not given).

Remarks: Awerinzew thinks this is the intermediate form between Leptotheca and Ceratomyxa.

CERATOMYXA sp. (?) Awerinzew

1913 Ceratomyxa sp. (?) Awerinzew 1913a: 154

Habitat: Gall-bladder of Scatophagus argus; Delagoa Bay (Africa). Vegetative form: Small, disc-form ameboid (25 to 35μ), containing two spores of indistinct contour, on account of incomplete formation of the shell. Two spores, apparently, developed in one pansporoblast. Disporous.

Spore: Form could not exactly be made out. Polar capsules were arranged like those of other Ceratomyxa.

CERATOMYXA sp. (?) Awerinzew

1913 Ceratomyxa sp. (?) Awerinzew 1913a: 154-155

Habitat: Gall-bladder of Rhinobathus Awer. (?); Lorenço Marques (Africa).

Vegetative form: Irregular shape. Endoplasm highly granular. In the epithelial layer of the gall-bladder numerous, spherical cysts (30 to 35μ) were found. Two spores are formed in one pansporoblast. Disporous.

Spore: Cylindrical with broad and slightly rounded ends. Dimensions: sutural diameter 16 to 19μ , breadth 70 to 80μ , thickness 16 to 19μ .

CERATOMYXA ACADIENSIS Mavor

[Figs. 44 to 47]

 1915
 Ceratomyxa acadiensis
 Mavor
 1915 : 27-30

 1916
 Ceratomyxa acadiensis
 Mavor
 1916 : 551-574

Habitat: Gall-bladder of *Urophycis chuss* (trophozoites are attached to undetermined Myxosporidia, see p. 176), *Zoarces angularis*, *Pseudo-pleuronectes americanus*; New Brunswick (Canada) (July to September).

Vegetative form: Polymorphous. Typically club-shaped with very long tail, or irregularly stellate. Pseudopodia show rigidity. Sometimes

clumps of protoplasm along their length, which are connected by thin hyaline filaments of ectoplasm. Differentiation of protoplasm is usually observable at the anterior region. Dimensions: length, excluding tail, 12 to 15μ , breadth 10 to 20μ , tail up to 60μ . Disporous.

Spore: Wide, short and slightly compressed dorso-ventrally, with very long fine lateral filaments. Polar capsules spherical. Polar filament invisible *in vivo*. Dimensions: breadth 40 to 50μ , sutural diameter 7 to 8μ , diameter of polar capsule 3 to 4μ , length of polar filament 70μ , length of lateral filaments 250 to 300μ .

CERATOMYXA sp. Georgévitch

1916 Ceratomyxa sp.

Georgévitch 1916a: 3

Habitat: Gall-bladder of Muraena sp.; Monaco (May).

Vegetative form: No description. Spore: No description. No figure.

CERATOMYXA CORIS Georgévitch

[Fig. 48]

1916 Ceratomyxa coris
 1917 Ceratomyxa coris
 Georgévitch
 1917a: 1-20

Habitat: Gall-bladder of Coris julius, C. giofredi; Villefranche (March, June).

Vegetative form: Various forms, club-shape, spherical or elongated, with lobose or filiform pseudopodia. Disporous and rarely Polysporous.

Spore: More or less ellipsoidal. Lateral prolongations of the shell-valves short and truncate. Sutural line straight. Sporoplasm, elongate, rounded, elliptical, fills a part of the extracapsular cavity of the spore. Polar capsules rounded, almost spherical, not converging. Dimensions not given.

Remarks: Georgévitch observed (1917: Fig. 30) that spores of Glugea marionis occurred in disporous trophozoite of Ceratomyxa coris, which he thought to have happened accidentally by plasmogamy of these two Cnidosporidia. The above mentioned figure, however, strongly suggests that G. marionis may be leading parasitic life in the trophozoite of C. coris.

CERATOMYXA HEROUARDI Georgévitch [Fig. 49]

1913	Leptotheca (?) sp.	Jameson	1913 : 2
1916	Ceratomyxa herouardi	Georgévitch	1916a : 5–8
1916	Ceratomyxa herowardi	Georgévitch	1916b : 717-19, 983-985
1917	Ceratomyxa herouardi	Georgévitch	1917:375-399

Habitat: Gall-bladder of Box Salpa L.; Monaco (May).

Vegetative form: Polymorphous. Elongated with same breadth or tapering to one end; club-shaped with roundish enlargements. Young trophozoites spherical or pyriform. Pseudopodia long and narrow or broad and bi- or multi-lobate. Body colorless both in the young and the adult. Protoplasm homogeneous and finely granular. Disporous and polysporous. Spores are found inside of the endoplasm and in the roundish buds, ordinarily two spores being formed in each bud. Number of buds on one trophozoite varies. Plasmotomy by budding and division.

Spore: Elongated elliptic. Polar capsules spherical and large. Sutural plane cuts the spore into exactly equal two parts. Two nuclei in sporoplasm are rather small and are always in one of the shell-valves. Dimensions not given.

Remark: The form, mentioned by Jameson in the same seat, host and locality, that "has something of the appearance of a Leptotheca" and that is also "almost certainly neither of the two Myxosporidia—Ceratomyxa pallida and Henneguya neapolitana . . . ," is probably identical with the present form.

CERATOMYXA MESOSPORA Davis [Fig. 50]

1917 Ceratomyxa mesospora Davis 1917: 223-224

Habitat: Gall-bladder of Cestracion zygaena, C. tiburo; Beaufort (July). Vegetative form: Pyriform, elongate, with long, slender posterior process. Numerous filiform pseudopodia at anterior end. Progressive movements rapid. Body colorless. No sharp demarcation between ectoplasm and endoplasm. Endoplasm finely granular and filled with small, colorless, homogeneous spherules. Spherules absent at anterior end. Size: total length 70 to 85μ, length exclusive of posterior process 50 to 75μ, breadth 20 to 25μ. Disporous.

Spore: Greatly elongate, each valve forming a slightly tapering cone, rounded at the apex. Valves not compressed. Sutural plane forming an acute angle with the longest diameter. Polar capsules conspicuous. Coiled polar filaments very distinct. Polar capsules are remarkable in that they are asymmetrically situated, one being always located in the widest part of the spore, while the other being a little to one side. Sporoplasm asymmetrically situated, sometimes being entirely confined to the larger valve. Dimensions: breadth 50 to 65μ , sutural diameter about 8μ , diameter of polar capsule 4.5μ , length of polar filament 90μ .

Remarks: Similar to C. sphaerulosa Thél. and occurs with C. recurvata Davis in the same organ.

CERATOMYXA SPHAIROPHORA Davis [Fig. 51]

1917 Ceratomyxa sphairophora Davis 1917: 224

Habitat: Gall-bladder of Scoliodon terrae-novae; Beaufort.

Vegetative form: Pyriform, elongate. Numerous fine filiform pseudo-podia at anterior end. Progressive movements rapid. Body colorless and transparent. Ectoplasm clear and homogeneous. Structure of endoplasm highly variable, in majority of trophozoites filled with transparent homogeneous spherules. Small fat globules at the anterior end. In some sporulating individuals, the endoplasm shows vacuolated structure without any spherules, usually, however, sporulating trophozoites exhibit well-defined spherules. The spherules or vacuoles, as the case may be, are separated by a thin layer of distinctly granular endoplasm containing numerous rod-shaped or rounded, colorless bodies, which in their appearance are strikingly like small bacteria tho they are not bacteria, as they fail to take Giemsa stain. Size of sporulating trophozoites 100 to 110μ by 25μ . Disporous.

Spore: Shell-valves greatly elongated, tapering gradually toward the ends. Long, attenuated ends of valves hollow and so fragile that it is almost impossible to find an example in which they are not more or less distorted. Sutural plane perpendicular or only slightly oblique to the longest diameter. Polar capsules are spherical and large; slightly convergent, opening some distance apart on the anterior side. Coiled polar filament distinct. Sporoplasm confined to large, central part of spores, but extending farther into one valve than the other. Dimensions: total breadth 115 to 140μ , sutural diameter about 12μ , diameter of polar capsules 6μ , length of polar filament 75μ .

CERATOMYXA TAENIA Davis [Figs. 52 and 53]

1917 Ceratomyxa taenia

Davis

1917:224-225

Habitat: Gall-bladder of Scoliodon terrae-novae; Beaufort.

Vegetative form: Similar to those of C. sphairophora Davis, and no character has been found by which they may be distinguished. Sporulating trophozoites can be easily distinguished on account of the very different appearance of the spore and their different arrangement within the trophozoites. The spores of this species are situated, as is usually the case in Ceratomyxa, with the greater part of the spore parallel to the long axis of the trophozoite, only a part of one valve being bent back along the rest of the spore. Size: sporulating trophozoites length 80μ , breadth 25μ . Disporous.

Spore: Valves greatly elongated. Shell very thin, the membrance on opposite sides of each valve being in contact for about two-thirds of its length, forming a thin ribbonlike structure; basal third of each valve only slightly compressed; terminal ribbonlike portion of each valve usually twisted so that plane of ribbon is at right angles to the main part of the spore. Polar capsules small, pyriform to spherical and convergent. Coiled polar filament indistinct. Sutural plane perpendicular to the longest

diameter. Sporoplasm finely granular, filling the basal third of each valve, sometimes extending farther into one valve than the other. Dimensions: breadth 140 to 150μ , breadth of central portion 45μ , sutural diameter 6μ , diameter of the polar capsules 3μ .

CERATOMYXA ATTENUATA Davis [Fig. 54]

1917 Ceratomyxa attenuata

Davis

1917:225

Habitat: Gall-bladder of Scoliodon terrae-novae; Beaufort (July).

Vegetative form: Elongate, pyriform, with long, tapering posterior process; at anterior end numerous long filiform pseudopodia. Progressive movements rapid. Ectoplasm distinct only at anterior end. Endoplasm filled with small, refractive, yellowish or brownish granules, which are uniformly distributed throughout the trophozoite. Between the brownish granules, the endoplasm is clear and colorless, not granular, except at extreme anterior end where it contains a clump of small fat globules. Size of full-grown trophozoites 100 to 120 by 27 μ . Disporous.

Spore: Valves greatly elongated; a symmetrical, one valve being about 15μ shorter than the other and ending abruptly; the longer valve tapering gradually to a point. About midway of each valve, is a thin septum; external to the septum the valves are empty. Polar capsules are large, opening on the anterior margin. Coiled polar filaments distinct. Sutural plane oblique to longitudinal axis, usually forming a ridge. Sporoplasm asymmetrically situated in central part of the spore. Dimensions: breadth 115μ , sutural diameter 9μ , diameter of polar capsules 4.5μ , length of polar filament 60μ .

CERATOMYXA RECURVATA Davis [Figs. 55 and 56]

1917 Ceratomyxa recurvata

Davis

1917:225-226

Habitat: Gall-bladder of Cestracion zygaena; Beaufort (July).

Vegetative form: Pyriform with long, slender posterior process. Body colorless. Actively motile, forming filiform pseudopodia of ectoplasm at anterior end. Endoplasm colorless and granular, filled with large, homogeneous spherules. Full-grown trophozoites 130 to 175μ , length of the main body about 100μ . Spores are developed singly from distinct sporoplasts and not necessarily in pairs. Disporous and polysporous (up to 10 spores, 6 and 8 are common numbers).

Spore: Valves greatly curved toward the posterior side, usually symmetrical, but occasionally one may be much more incurved than the other. Valves circular in cross section at the base but toward the ends greatly

flattened. Ends of valves sharply pointed. Polar capsules large, opening at some distance from the anterior margin. Coiled polar filaments distinct. Sporoplasm finely granular usually extending farther into one valve than the other. Dimensions: breadth between points of greatest curvature about 16μ , sutural diameter 8 to 9μ , diameter of polar capsules 4.5μ .

CERATOMYXA LUNATA Davis

[Figs. 57 to 60]

1917 Ceratomyxa lunata

Davis

1917 : 226-227

Habitat: Gall-bladder of Galeocerda tigrinus; Beaufort (August).

Vegetative form: Pyriform, rounded after being on the slide for some time. Progressive movements slow Endoplasm filled with large, homogeneous spherules, which are usually colorless, sometimes light yellow. At extreme anterior end, the endoplasm contains numerous small fat globules. Disporous.

Spore: Considerably variable in size and form. The larger and more typical are more or less crescent-shaped; symmetrical; valves curved toward rear, terminating in more or less rounded ends. Polar capsules large and open on opposite sides of spore. Coiled polar filament distinct. Sporoplasm finely granular, symmetrically situated in spore. Smaller spores differ from large ones chiefly in size; valves are much shortened and have a greater curvature, with more distinctly rounded ends. Dimensions: breadth 30μ (longest 38μ), sutural diameter 9μ , diameter of polar capsules 4μ , length of polar filament 37μ . Small forms: breadth 15μ , sutural diameter 7μ , diameter of polar capsules 3μ .

CERATOMYXA ABBREVIATA Davis

[Fig. 61]

1917 Ceratomyxa abbreviata

Davis

1917:227

Habitat: Gall-bladder of Scoliodon terrae-novae; Beaufort (August).

Vegetative form: Elongate, pyriform, with usually a very long, slender posterior process. Body colorless. Progressive movements rapid. Distinct differentiation of protoplasm, posterior process usually composed of ectoplasm (rarely endoplasm may extend into it for a short distance). Pseudopodia, short, tapering or filiform at anterior end. Dimensions: length up to 90μ , breadth 10 to 12μ , diameter of rounded sporulating trophozoites about 27μ . Disporous.

Spore: Roughly crescent-shaped; sutural diameter exceptionally great in comparison with the breadth. Ends of valves rounded, slightly asymmetrical. Shell exceptionally tough and resistant to reagents. Polar capsules large, prominent and open on opposite sides of spore. Sporoplasm finely granular, confined entirely to one valve. Dimensions: breadth 17μ , sutural diameter 14μ , diameter of polar capsules 4.5μ .

CERATOMYXA FLAGELLIFERA Davis [Fig. 62]

1917 Ceratomyxa flagellifera

Davis

1917:227

Habitat: Gall-bladder of Carcharhinus sp?; Beaufort (July).

Vegetative form: Pyriform, short, tapering toward the posterior end, sometimes dividing into a number of long, slender, transparent processes. Extremely long filiform pseudopodia, developed at anterior end, can be seen to sweep slowly back like a whiplash until they come to lie by the side of the body. Progressive movements slow. Ectoplasm clear, transparent, forming a distinct layer at anterior end. Endoplasm in large trophozoites filled with large numbers of rod-shaped, bacteria-like bodies, which are more abundant in the anterior half than in the posterior. Endoplasm in younger trophozoites, with much less or without any bacteria-like bodies, shows a vacuolated structure. Size up to 115 to 120μ in length and 40 to 45μ in breadth. Disporous.

Spore: Valves greatly elongated, conical, with rounded ends. Sutural ridge well marked. Polar capsules large, opening on opposite sides of spore. Coiled polar filament very distinct. Sporoplasm granular, symmetrically situated, but extending only a short distance into each valve. Dimensions: breadth 118μ , sutural diameter 12μ , diameter of polar capsules 6μ .

CERATOMYXA AGGLOMERATA Davis [Fig. 63]

1917 Ceratomyxa agglomerata

Davis

1917:228

Habitat: Gall-bladder of Synodus foetans; Beaufort.

Vegetative form: Pyriform, usually with long, slender, posterior process. Body colorless and transparent. Actively motile, moving by means of characteristic wavelike movements of the ectoplasm, from which are projected numerous short, conical to filiform pseudopodia. Pseudopodia travel back along sides of body for about one-third its length and then disappear, new ones being continually formed at the anterior end. Ectoplasm distinguishable at anterior end. Endoplasm clear, very transparent, usually homogeneous, sometimes finely granular. Large numbers of fat globules usually present. Size of sporulating trophozoites 38μ by 12μ . Disporous.

Spore: Asymmetrical, one valve being smaller and more attenuated than the other; larger valve compressed. Polar capsules spherical. Coiled polar filaments indistinct. Sporoplasm filling nearly entire smaller valve, but only extending a short distance into the larger one. Dimensions: breadth 24 to 28μ , sutural diameter 5μ , diameter of polar capsules 3μ .

CERATOMYXA AMORPHA Davis [Fig. 64]

1917 Ceratomyxa amorpha

Davis

1917:228

Habitat: Gall-bladder of Synodus foetans; Beaufort.

Vegetative form: Rounded or irregular in shape, with short lobose pseudopodia; not pyriform; slowly ameboid. Body colorless. Ectoplasm well developed, forming a distinct layer; transparent, finely granular. Endoplasm granular, with large numbers of small fat globules scattered through it or aggregated into one or two large clumps (difference between the present form and *C. agglomerata*). Disporous.

Spore: Asymmetrical; crescent-shaped; valves short, conical, somewhat compressed. One valve distinctly smaller and more conical than the other. Sutural ridge perpendicular to longitudinal axis. Polar capsules large, opening at some distance from the anterior side. Coiled polar filaments distinct. Sporoplasm granular, asymmetrically situated, being chiefly confined to smaller valve. Dimensions: breadth 27μ , sutural diameter 11μ , diameter of polar capsules 4μ .

CERATOMYXA MONOSPORA Davis

[Figs. 65 to 67]

1917 Ceralomyxa monospora

Davis

1917:228-229

Habitat: Gall-bladder of *Peprilus alepidotus*; Beaufort. Abundantly present in June, much less in July, being entirely absent in the bladder at the end of the month.

Vegetative form: Pyriform, with a slender posterior process and one to several filiform pseudopodia at anterior end. Body colorless and transparent. Movements very slow. No clear differentiation between ectoplasm and endoplasm, the entire body being composed of a clear, finely granular protoplasm. Fat globules more abundant in larger individual, which are aggregated into small clumps. Size of vegetative trophozoites up to 24μ in length and 15μ in width. Monosporous form much smaller than disporous. Monosporous and disporous. Nearly entire substance of trophozoite is used up in spore formation.

Spore: Crescent-shaped. Valves cylindrical, tapering toward the end, which is rounded and compressed. Curvature of valves varies. One valve is more attenuated than the other. Sutural ridge perpendicular to the longest diameter. Polar capsules large. Sporoplasm usually asymmetrically situated. Dimensions: breadth 18 to 25μ , sutural diameter 5 to 6μ , diameter of polar capsules 3μ .

Remarks: This species is evidently very close to *C. pallida* Thél. Similar form was found in *Prionotus evolans* (gall-bladder), which showed somewhat larger trophozoites and spores than *C. monos pora*.

CERATOMYXA STREPTOSPORA Davis [Figs. 68 and 69]

1917 Ceralomyza strepiospora

Davis

1917:229

Habitat: Gall-bladder of Chaetodipterus faber; Beaufort (June, but not in July).

Vegetative form: Pyriform, colorless and transparent. A few conical, filiform, wavelike pseudopodia at anterior end. Ectoplasm recognizable at anterior end. Endoplasm finely granular, with a few, small, fat globules, filled with transparent, homogeneous spherules. Size: 48 by 12μ to 60 by 9μ . Disporous.

Spore: Compressed valves greatly elongated, with rounded ends. Sutural ridge. Polar capsules spherical. Coiled polar filament indistinct. Sporoplasm finely granular, entirely filling both valves. Dimensions: breadth 34 to 39μ , sutural diameter 4μ , diameter of polar capsules 3μ .

CERATOMYXA AGGREGATA Davis [Fig. 70]

1917 Ceratomyxa aggregata

Davis

1917:229

Habitat: Gall-bladder of Leostomus xanthurus, Micropogon undulatus; Beaufort (July).

Vegetative form: Form rounded to somewhat irregular in shape, rarely pyriform; slowly ameboid. Body colorless and transparent. No clear differentiation of protoplasm. Endoplasm finely granular, containing numbers of small fat globules. Sporulating trophozoites show a tendency to collect in groups composed of a large number of individuals so closely associated that it is often impossible to make out the individual outlines. Size of full-grown form 18μ by 14μ . Disporous.

Spore: Crescent-shaped; valves much elongated, tapering toward the ends, which are compressed. Polar capsules spherical and opaque. Sporoplasm granular, situated symmetrically in the spore cavity. Dimensions: breadth about 50μ , sutural diameter 6 to 7μ , diameter of polar capsule 3.5μ .

CERATOMYXA UNDULATA Davis [Fig. 71]

1917 Ceratomyxa undulata

Davis

1917:230

Habitat: Gall-bladder of Ancylopsetta quadrocellata Gill.; Beaufort (June to August).

Vegetative form: Pyriform, sometimes fusiform, tapering toward posterior end. Movements rapid. Body colorless. Ectoplasm observable at anterior part, constantly undergoes rapid, wavelike undulating movements and extrudes fine conical or filiform pseudopodia. Pseudopodia

are formed very rapidly and vary in length. After reaching a considerable length the pseudopodia usually travel posteriorly along sides of body and then disappear. Endoplasm very transparent, often vacuolated, containing numerous small fat globules. Size of full-grown trophozoite: 25μ by 10 to 12μ in average. Disporous.

Spore: Crescent-shaped. Valves cylindrical, not compressed, ends rounded, one valve being somewhat longer and more conical than the other. Polar capsules convergent. Coiled polar filaments distinct. Sporoplasm granular, asymmetrically situated, sometimes being almost confined to more conical valve. Dimensions: breadth 22 to 24μ , sutural diameter 6μ , diameter of polar capsules 3μ .

CERATOMYXA NAVICULARIA Davis

[Figs. 72 and 73]

1917 Ceratomyxa navicularia

Davis

1917:230

Habitat: Urinary bladder of Paralichthys dentatus, P. albiguttus, Sphaeroides maculatus; Beaufort (June to August).

Vegetative form: Rounded or slightly irregular in shape, never pyriform. Body colorless. Very slow ameboid. No distinct ectoplasm. Entire trophozoite finely granular, containing a few small fat globules. Nearly entire body is used up in the formation of spores. Diameter about 17μ . Disporous.

Spore: Variable in shape and size. Symmetrical or asymmetrical, often boat-shaped, slightly compressed dorso-ventrally, with rounded ends. Polar capsules convergent, shows polar filament indistinctly. Sporoplasm finely granular, extending into both valves, but usually somewhat farther into one than the other. Dimensions: breadth 14 to 22μ (average 16μ), sutural diameter 5 to 7.5μ (average 6μ), diameter of polar capsules 2μ .

CERATOMYXA SPINOSA Davis [Fig. 74]

1917 Ceratomyxa spinosa

Davis

1917:230

Habitat: Urinary bladder of Paralichthys albiguttus; Beaufort.

Vegetative form: Rounded or slightly irregular in shape, with short, lobose pseudopodia; slowly ameboid. Body colorless and transparent. Distinct differentiation of protoplasm along the entire surface, ectoplasm forming outer layer. Endoplasm faintly granular, with numerous small fat globules. Monosporous and disporous.

Spore: Central portion greatly enlarged; ovoid, with very long tapering process extending out from each end. Sutural line perpendicular to the longest diameter. Polar capsules large and spherical. Sporoplasm finely

granular, chiefly located in one valve, extending into the other only a short distance beyond the capsule. Dimensions: breadth 80μ , breadth of enlarged central portion 13μ , sutural diameter 7μ , diameter of polar capsules 4μ .

Genus MYXOPROTEUS Doflein emend. Davis

1898	M yxo proteus	Doflein	1898:287
1917	M yxo proleus	Davis	1917 : 219

The characters of the genus are described on page 56. Type species: Myxoproteus ambiguus (Thélohan) Doflein.

MYXOPROTEUS AMBIGUUS (Thélohan) Doflein [Figs. 75 to 80]

1895	Myxosoma ambiguum	Thélohan	1895 : 3 44
1898	Myxoproteus ambiguus	Doflein	1898 : 287-288

Habitat: Urinary bladder of Lophius piscatorius; Le Croisic, Rovigno, Napoli.

Vegetative form: Polymorphous. Color milky white. Protoplasm is filled with numerous granules and fat globules. Pseudopodia, short, pointed lobose. Plasmogamy and plasmotomy take place. Many small individuals formed apparently by plasmotomy, often, make up groups. Disporous, polysporous?

Spore: Almost pyramidal, with anterior processes. Two very large polar capsules at the anterior end. The distance between these capsules is equal to or greater than the diameter of the capsules. Sporoplasm with two nuclei. Dimensions: length 25μ , breadth 18 to 20μ , diameter of polar capsules 7μ .

MYXOPROTEUS CORDIFORMIS Davis [Figs. 81 to 83]

1917 Myxoproteus cordiformis Davis 1917: 231

Habitat: Urinary bladder of Chaetodipterus faber; Beaufort (June, July).

Vegetative form: Rounded; very slowly ameboid, usually forming a single, short, lobose pseudopodium. Body colorless and transparent. Ectoplasm not distinct. Entire trophozoite finely granular, with a few fat globules. Rarely vacuolar. Small trophozoites often show a single large, central vacuole. Rounded sporulating trophozoites 18μ in diameter. Disporous.

Spore: Heart-shaped in front view, with peculiar wing-like expansions on each side which contain remains of parietal cells. Sutural plane oblique in position. Capsulogenous cells distinct. Sporoplasm finely granular, fills the extracapsular cavity of the spore. Dimensions: length 12μ , breadth 10 to 11μ , thickness 6μ , polar capsules 3 to 4μ in diameter.

MYXOPROTEUS CORNUTUS Davis [Fig. 84]

1917 Myxoproteus cornulus Davis 1917: 231

Habitat: Urinary bladder of Bairdiella chrysura; Beaufort.

Vegetative form: Somewhat elongated or irregular in shape, with short lobose pseudopodia; slowly ameboid. Differentiation of protoplasm clear. Ectoplasm well developed, hyaline; in rounded individuals forming a distinct layer around the body. Endoplasm opaque, contains coarse refringent granules varying in shape, and a few fat globules. In contracted rounded resting condition, endoplasm becomes condensed, while ectoplasm appears more abundant. Rounded trophozoites up to 27μ in diameter. Disporous.

Spore: Heart-shaped, with two anterior processes. Shell very thick. Polar capsules large, opening some distance apart. Coiled polar filament distinct. Sporoplasm finely granular, with a few small fat globules, fills the extracapsular cavity of the spore. Dimensions: sutural diameter exclusive of the processes 9μ , breadth 12μ , length of processes 5μ , diameter of polar capsules 3μ .

Genus WARDIA nov. gen.

The characters of the genus are described on page 56.

Type species: Wardia ovinocua nov. spec.

WARDIA OVINOCUA nov. spec. [Figs. 85 to 95]

Habitat: Ovum and connective tissue of ovary of *Lepomis humilis* Girard;* Salt Fork, Ill. (November). Only one fish, 6.5 cm. long with normal appearance, was found to be infected.

Vegetative form: Trophozoites form cysts visible to the naked eyes as white spherical spots in the pink-colored ovary. Four cysts present. The cyst (Figs. 85 and 86), in section, shows a circular form surrounded by several layers of hypertrophied nurse cells and connective tissue, in which many large blood vessels are present. Protoplasm is not clearly differ-

*Professor F. Smith of the Department kindly identified all the fish that were collected in the vicinity of Urbana and mentioned in this paper as hosts, for which the writer wishes to express his appreciation. entiated into ectoplasm and endoplasm, the whole protoplasm showing granulated reticular structure. Cysts contained numerous fully developed spores and a small number of spores in developmental stages, which suggested the fact that two spores rise from each pansporoblast. The parasite is also found in the state of diffuse infiltration in the connective tissue around the cyst. Diameter of cysts 316 to 445μ in sections. Polysporous.

Spore: In front view, isosceles triangular form, two sides of which usually convex, with more or less attenuated anterior end (Figs. 87, 90, 92); in profile, ellipsoidal (Fig. 88); and oval viewed from the anterior end (Fig. 89). Sutural plane at right angles to the longest diameter (Figs. 87 and 89). Shell comparatively thin except at the anterior end and has many fine network-like ridges on the surface. These ridges are hardly observable on fresh spores on account of their fine form and the conspicuously large polar capsules lying in the spore. When stained, however, they are not only made distinctly visible, but the prolongation of each ridge from the posterior edge which forms about 1µ long fringe-like structure is also more clearly recognized (Figs. 90-95). Two large and spherical polar capsules located in the central portion of the spore. Coiled (5 to 6 times) polar filament extremely distinct. The openings of polar capsules at the anterior end. Sporoplasm comparatively small, finely granular, without any vacuole, contains two small nuclei, when stained. Dimensions in vivo: sutural diameter 9 to 10μ , breadth 10 to 12μ , thickness 6μ , diameter of the polar capsule 4μ , length of polar filament 35 to 45μ .

WARDIA OHLMACHERI (Gurley) Kudo

[Figs. 96 and 97]			
1893	M yxos poridian	Ohlmacher	1893 : 561-567
1893	Chloromyxum ohlmacheri	Whinery	1893 : 660-662
1894	Chloromyxum (Sphaerospora)	
	oklmackeri	Gurley	1894 : 267-272
1895	? Leptotheca ranae	Thélohan	1895 : 383
1899	Lebtotkeca oklmackeri	Labhé	1890 · 87

Habitat: Urinary tubules of kidney of *Bufo lentiginosus* Shaw and kidney of *Rana esculenta* and *R. temporaria* (*R. fusca*); Sycamore, De Kalb county, Ill.

Vegetative form: Not found.

Spore: Transversely elliptic. Sutural plane perpendicular to the longer axis of the spore. A well defined undulate-parallel longitudinal striation on the shell. Sutural ridge comparatively well marked. Two polar capsules lying side by side, occasionally only one. Dimensions: sutural diameter 6μ , breadth 8μ , diameter of polar capsule 3 to 3.5μ , length of polar filament 6 to 8 times the breadth of spore (48 to 64μ).

Remarks: This form is apparently very much different from any species of genus Leptotheca, in the general form, form of polar capsules,



striations on the shell and the habitat. Tho the form of the spore is different from the type species of the genus Wardia and nothing is known about the vegetative form, the presence of large spherical polar capsules in the central portion of the spore, the striations on the shell and the occurrence of the same nature, i.e., from fresh waters in the close-by localities, show its nearer relationship to the genus Wardia than to the genus Leptotheca. Hence it is placed here provisionally.

Genus MITRASPORA Fujita emend. Kudo

1912 Mitraspora

Fujita

1912:259-260

The characters of the genus are described on page 56.

Type species: Mitraspora cyprini Fujita.

MITRASPORA CYPRINI Fujita

[Figs. 98 to 104]

1912 Mitraspora cyprini

Fuiita

1912:259-260

Habitat: Renal tubules of kidney and ureters of Cyprinus carpio L. and Carassius auratus L.; Sapporo (winter), Tokio (March).

Vegetative form: Fujita's only description is as follows: "The sporoblast contains generally three or four spores." The present writer observed a similar form in the ureter and the renal tubule of the kidney of *Cyprinus* carpio L., in Tokio. The observations are as follows: Trophozoites small ameboid (Fig. 98). Body colorless. Movements tardy. Differentiation of protoplasm imperfect. The hyaline ectoplasm recognizable at one side of the body, where lobose pseudopodia are formed (Figs. 98-99). Endoplasm granular with vacuoles and brownish granules, which become larger as the body grows. Size 10 to 40μ . Disporous (Kudo) and polysporous (?, Fujita).

Spore: Fujita's descriptions are as follows: Form resembles monk's hood, slightly attenuated at its anterior end. Shell uniformly thin, except at two points of the truncated posterior end. Each shell valve has eight distinct striations which run longitudinally and turn into long cilia up to 5.8μ long, planted in a single row at the posterior end of the spore. Two polar capsules at the anterior end. The nucleus is obscure and no vacuole is present. Dimensions: length 10 to 13μ , breadth 5μ , polar capsules 3.8μ by 2μ , length of polar filament 15μ (weak glycerine). The writer observed the following facts: More rounded with rounded anterior end in front and side views. Shell more or less thick along the entire posterior margin. Striations on shell, variable in number. Sporoplasm granular, without any vacuole, exhibits two nuclei when stained. Posterior filaments 5 to 6 in number and 5 to 6μ long, being absent in some spores. Dimensions in vivo: length 10μ , breadth 8 to 9μ , thickness 6 to 8μ , polar capsule 4μ by 1.5 to 2μ , length of polar filament 35 to 40μ .

Remarks: The Fujita does not describe the vegetative form and there are some differences in the form and size of the spore between the forms, the writer does not find out any objection against the union of the above mentioned two forms.

MITRASPORA CAUDATA (Parisi) Kudo [Figs. 105 to 107]

1910	Sphaerospora caudata	Parisi	1910 : 253-254
1912	Sphaerospora caudata	Parisi	1912:289
1913	Sphaerospora caudala	Parisi	1913 : 396-402

Habitat: Renal tubules of kidney of Alosa finta Cuv. var. lacustris Fatio; Lake Como.

Vegetative form: Rounded or variously elongated owing to the movements. Protoplasm is distinctly differentiated into ectoplasm and endoplasm. Ectoplasm, hyaline and homogeneous, forms slowly moving lobose pseudopodia. Endoplasm granular, contains yellow globules and fat granules. Disporous and polysporous.

Spore: Subspherical in front view; oval in profile; anterior end being more rounded than the posterior end. Shell rather thick, longitudinally striated. In front view, the posterior end enlarged into a quadrangular form, which appears as a small spine in side-view and which projects backward long and fine filaments, usually six in number. Two well developed polar capsules open on each side of the sutural plane. Polar filament coiled 5 to 6 times. Sporoplasm without any iodinophilous vacuole. Dimensions: external length 10 to 11μ , internal length 7 to 9μ , length of polar capsules 4 to 4.5μ , length of polar filament up to 48μ , length of posterior filaments up to 28μ .

MITRASPORA ELONGATA nov. spec.

[Figs. 602 to 621]

Habitat: In the urinary tubules and tissue of kidney of Lepomis cyanellus; Crystal Lake, Urbana, Ill. From June to July, all the fish examined, 36 in number and 10 cm. in average length, were found to be infected. Other fish such as Lepomis pallidus and Lepomis humilis, caught at the same time, were free from the infection. Early in June, the number and size of the parasites in a host body were rather small and only a small number of spores could be recognized in the fresh state with the addition of potassium hydrate solution. The growth of the parasite was rather remarkable during the hot weeks in the latter part of June and July so that every fish caught on July 17th showed a heavy infection, exhibiting small whitish pustules over the surface of the organ. During June, vegetative forms and spores were found in the lumen of the urinary tubules,

altho some contained the parasitic masses in the tissue. About the middle of July, the parasite forms conspicuous cysts in the tissue thruout the organ. The cyst may or may not be covered by a thick layer of connective tissue from the host. Aside from this hypertrophy, the host did not show any pathological change which could be recognized.

Vegetative form: Youngest trophozoite found in the urinary tubule is multinucleate, rounded, and of from 20 to 50μ in diameter. The protoplasm is not differentiated, the entire body is finely granular or coarsely reticular in structure. In the protoplasm are to be seen nuclei and sporoblasts at different stages of development. The union of two propagative cells similar to that of Myxobolus toyamai produces a small body which developes into a single sporoblast and ultimately into a single spore (Figs. 605-613). In later stages, the trophozoite reaches a size of 200μ in diameter showing many stages of spore formation and mature spores, surrounded by thick layers of connective tissue from the host. Polysporous.

Spore: Elongated oblong with pointed anterior and truncated posterior extremities. The width is often greatest at the middle of the polar capsules, the posterior portion is much narrower than the anterior. Nearly circular in the cross-section thru the polar capsules. The shell is thin. the sutural line being faintly marked in fresh state. It generally is obliquely located in relation to the capsules. The shell also shows fine longitudinal striations, 14 to 16 in number, on each valve. The sutural line as well as the striations are best seen in spores stained with Heidenhain's iron hematoxylin. Two polar capsules elongated pyriform, mostly equal in size, occupy the anterior half of the spore. Abnormal situations of the polar capsules are sometimes observed (Fig. 619). The coiled polar filament is faintly visible in fresh spores. It is spirally coiled along the wall of the polar capsule without any central axis. This fact was clearly observed in stained section as is shown in Figs. 620 and 621. The filament has seven or eight windings, thus agreeing with the actual length of the extruded polar filament. The polar filament was extruded under the action of potassium hydrate solution. The extrusion takes place even in some spores which were treated with Schaudinn's fixative and kept in 95 per cent alcohol for three months (see the similar observations on Myxobolus discrepans on page 157). The sporoplasm is finely granular and transparent. When stained, it shows two nuclei in the center or near the posterior part of the body. Dimensions of preserved spores: length 15 to 17μ , breadth 5 to 6μ , thickness 4.5 to 5.5μ , polar capsule 7.5μ by 2μ , length of polar filament 40 to 50µ.

Suborder SPHAEROSPOREA nom. nov. The definition of the suborder is recorded on page 57.



Family CHLOROMYXIDAE Thélohan

1892	Chloromyxidées	Thélohan	1892:173
1895	Chloromyxidées	Thélohan	1895 : 344

The characters of the family are described on page 57.

Genus CHLOROMYXUM Mingazzini

1890	Chloromyxum	Mingazzini	1890:160
1892	Chloromyxum	Thélohan	1892:173-176
1895	Chloromyxum	Thélohan	1895 : 344

The characters of the genus are described on page 57. Type species: Chloromyxum leydigi Mingazzini.

CHLOROMYXUM LEYDIGI Mingazzini [Figs. 108 to 113]

1851		Leydig	1851 : 225-234
1852		Leuckart	1852:435
1854		Lieberkühn	1854 : 352
1890	Chloromyxum leydigi	Mingazzini	1890 : 160-164
1892	Chloromyxum leydigi	Thélohan	1892 : 166, 169-170
1894	Chloromyxum leydigi		
	Chloromyxum incisum	Gurley	1894: 259-260
1895	Chloromyxum leydigi		
	Chloromyxum incisum	Thélohan	1895 : 345-346
1898	Chloromyxum leydigi	Doflein	1898 : 292, 310, etc.
1912	Chloromyxum leydigi	Erdmann	1912 : 149-162
1916	Chloromyxum leydigi	Georgévitch	1916a:3
1917	Chloromyxum leydigi	Davis	1917:236-237
1917	Chloromyxum leydigi	Erdmann	1917:276-321
1918	Chloromyxum leydigi	Georgévitch	1918 : 182-189

Habitat: Gall-bladder of Rhina squatina L., Spinax spinax L., Scyllium canicula, S.asterias, Raja batis L., R. clavata L., R. undulata Lac., Torpedo narce Ris., T. marmorata, T. ocellata, T. torpedo L., Acanthias acanthias L., Trygon pastinaca L., Dasybatis hastatus, D. sabina, Pteroplatea maclura Le Sueur, Scoliodon terrae-novae, Cestracion zygaena, C. tiburo, Carcharhinus limbatus; Roscoff, Concarneau, Marseille, Banyuls, Rovigno, Heligoland, Beaufort, Monaco (May), Napoli, Genova. Erdmann observed the species at Naples from March to August. She noticed mixed infection with Ceratomyxa reticulata and especially with Leptotheca parva. Georgévitch studied the parasite at Monaco from February to April.

Vegetative form: Polymorphous, being spherical, oval or irregular. The change of the form rather rapid under favourable conditions. Differentiation of protoplasm distinct. Ectoplasm with pseudopodia of various form, i.e., lobose, filiform or intermediary; short, pointed or branched. Endoplasm alveolar, filled with yellowish granules. Doflein observed the plasmotomic multiplication of young trophozoites. Polysporous. Erdmann's observations (1917) may be summarized as follows: Ameboid.

Color of the body greenish to dark green. The protoplasm is clearly differentiated into ectoplasm and endoplasm. The ectoplasm is hyaline and covers the entire surface of the body. It appears as a fine fibrous structure when fixed with Bouin's solution. The endoplasm contains besides nuclei, two kinds of spherules; one smaller and yellowish "color-carriers" and the other larger and light to dark greenish reserve bodies. The color-carrier is in part composed of myelin, while the reserve body is of glycogenous nature. The infection was studied experimentally per os: young trophozoites appeared in 3 to 5 days which continued to 10th day, various trophozoites were seen in 13 to 19 days, and sporulating individuals were first recognized in 39 days after the infection. The trophozoite multiplies in number either by fission or by budding. It usually contains enclosures which seem to be degenerating erythrocytes. Mictosporous.

Spore: Ovoidal. Shell-valves show wide edge at sutural plane, which is attenuated at the anterior end and forms a quadrilateral process at the posterior end, from which a row of cilia grows. Shell-valves have ridges (6 to 7), which run parallel to the posterior margin. The striations may vary considerably. Four polar capsules at the anterior end. Dimensions: length 8μ . Erdmann gave the following dimensions: Spores from Torpedo marmorata and T. ocellata: length 6 to 9μ , breadth 5μ , polar capsule 3μ by 2μ . Those from Scyllium asterias: length 8 to 9μ , breadth 6μ , polar capsule 2μ by 1μ . Those from Raja batis: length 7 to 8μ , breadth 5μ , polar capsule 2μ by 1μ . Length of polar filament 20 to 30μ (absolute alcohol warmed up to 40° C.).

CHLOROMYXUM CAUDATUM Thélohan [Fig. 114]

1895 Chloromyxum caudatum Thélohan 1895 : 346

Habitat: Gall-bladder of Molge cristata Laur.; Vicinity of Rennes.

Vegetative form: Body yellowish with lobose pseudopodia. Protoplasm finely granular.

Spore: Oval or spheroidal. Shell enlarged at the anterior part, having a simple or bifurcated tail-like process, as in Henneguya, at the posterior end. Dimensions: total length 18μ , length 8μ , breadth 6 to 7μ , length of tail 10μ .

CHLOROMYXUM QUADRATUM Thélohan [Figs. 115 to 117]

1891		Pfeiffer	1891:111
1893		Pfeiffer	1 89 3 : 81
1895	Chloromyxum quadratum	Thélohan	1895 : 347
1912	Chloromyxum quadratum	Parisi	1912:289
1913	Chloromyxum quadratum	Awerinzew	1913a : 155
1913	Chloromyxum quadratum	Fermor	1913 : 199

Habitat: Muscle of Syngnathus acus L., Trachurus trachurus L., Nerophis aequorens L., Callionymus lyra L., Coris julis L., Ariodes polystaphylodon, kidney of Blennius gattorugine Brunn; Helder, Roscoff, Concarneau, Marseille, Beira (Africa), Napoli (summer).

Vegetative form: Not described by any of these authors.

Spore: Quadrangular pyramid with curved edges and roundish angles. Four polar capsules at the anterior end. Dimensions: length 6μ , breadth 5μ , length of polar filament 8 to 10μ .

CHLOROMYXUM FLUVIATILE Thélohan

[Fig. 118]

1892	Chloromyxum fluviatile	Thélohan	1892 : 173-176
1895	Chloromyxum fluviatile	Thélohan	1895 : 346 ⁻

Habitat: Gall-bladder of Leuciscus cephalus L.; Paris.

Vegetative form: Young trophozoites colorless; adults yellowish. Form highly variable. Active change of the form of body. Clear differentiation between ectoplasm and endoplasm. Ectoplasm usually recognizable at one end of the body where lobose pseudopodia are formed. Size reaches 25 to 30μ . Polysporous.

Spore: Spherical, generally small. Sutural ridge fairly well marked. Dimensions: 7 to 8μ in diameter.

CHLOROMYXUM MUCRONATUM Gurley

[Figs. 119 to 122]

1854		Lieberkühn	1854:352-353
1879		Leuckart	1879 : 248
1882		Bütschli	1882 : Pl. 38 : 17
1883		Balbiani	1883 : 201, 203
1893	Chloromyxum mucronatum	Gurley	1893 : 419
1894	Chloromyxum mucronatum	Gurley	1894 : 264, 265
1908	Chloromyxum mucronatum	Auerbach	1908:456
1909	Chloromyxum mucronatum	Auerbach	1909a:71

Habitat: Urinary-bladder and kidney of Lota lota L.; Bodensee, other localities not mentioned.

Vegetative form: Spherical, elliptical or irregular. Size up to 75μ in diameter. Protoplasm containing irregularily scattered fat-like globules.

Spore: Sharp-contoured; subglobular, mucronate anteriorly. Dimensions: length 8μ .

CHLOROMYXUM DIPLOXYS (Gurley) Thélohan [Figs. 123 to 125]

1866		Balbiani	1866 : 600-602
1867		Balbiani	1867 : 275, 276, 335
1882		Bütschli	1882 : 590
1890		Pfeiffer	1890 : 559
1892		Henneguy et Thélohan	1892 : 587
1893	Cystodiscus? diploxys	Gurley	1893 : 411
1894	Cystodiscus? diploxys	Gurley	1894 : 281
1895	Chloromyxum diploxys	Thélohan	1895 : 347

Habitat: Abdominal cavity of Tortrix viridana L. (imago);

Vegetative form: Trophozoites form spherical cysts, 230 to 400μ in diameter. Cyst membrane rather thick. Protoplasm containing brownish granules, and fat-like globules (red with iodine).

Spore: Elliptic or slightly flattened. Sutural line straight, forming a ridge. Two polar capsules at each end.

CHLOROMYXUM PROTEI Joseph

1905	Chloromyxum protei	Joseph	1905 : 450-451
1907	Chloromyxum protei	Joseph	1907:398-412

Habitat: Renal tubules of kidney of Proteus anguineus L.; Vienna.

Vegetative form: Generally rounded or sausage form. No clear differentiation between ectoplasm and endoplasm. Movements slow. Probable occurrence of plasmotomy by budding and division. Size: $40 \text{ to } 45\mu \text{ by } 28 \text{ to } 40\mu$.

Spore: Spherical. Shell finely striated parallel to the sutural line. Four polar capsules each with an independent opening. Dimensions: 10 to 13μ in diameter, polar capsules 4 to 6μ long. The polar filament appears to be rather short.

CHLOROMYXUM TRUTTAE Léger [Fig.126]

1906 Chloromyzum trutiae Léger 1906: 267-270

Habitat: Gall-bladder and gall-duct of Trutta fario L.; Dauphiné.

Vegetative form: Ameboid form. Elongated. Form resembles an Amoeba limax of about 40μ in length. Roundish or irregularly contoured, with small pseudopodia. Ovoidal or spherical, 25 to 40μ in diameter without any visible pseudopodia (resting state). Body colorless, clear and hyaline. Very active movements which last for several hours after the death of the host. Broad and obtuse pseudopodia well developed at the anterior end of the body. Endoplasm alveolar, contains variable numbers of nuclei, which are seen in vivo, refractive bodies and chromatic granules. Monosporous(?) and polysporous.

Spore: Spherical. Four polar capsules of different size. Shell-valves marked with parallel ridges. Dimensions: 8 to 9μ in diameter.

CHLOROMYXUM CRISTATUM Léger [Figs. 127 and 128]

1906 Chloromyxum cristatum

Léger

1906: 270-272

Habitat: Gall-bladder of Tinca vulgaris Cuv.; Grenoble.

Vegetative form: Ordinarily massive, with oval or round contours, without noticeable pseudopodium. Ectoplasm hyaline. Endoplasm granular and colorless. Average diameter of the adults about 20μ . Monosporous, rarely disporous.

Spore: Spherical or subspherical. Ten marked ridges run anteroposteriorly on each shell-valve, so that it presents a cog-wheel form in cross section. Four polar capsules at the anterior end, one pair being smaller than the other. Sporoplasm with two nuclei. Dimension: 10 to 11μ .

CHLOROMYXUM DUBIUM Auerbach [Figs. 129 to 133]

 1908
 Chloromyxum dubium
 Auerbach
 1908 : 456-459

 • 1910
 Chloromyxum dubium
 Auerbach
 1910c : 177

Habitat: Gall-bladder of Lota vulgaris Cuv.; Bodensee (April to September).

Vegetative form: Spherical or rounded. Rarely irregular forms. Protoplasm is differentiated distinctly into ectoplasm and endoplasm. Ectoplasm very thin, forms pseudopodia which move slowly. Endoplasm granular, contains fat globules. Majority of the trophozoites appear to live floating in the bile, while some are attached to the epithelium of the bladder. Disporous and polysporous.

Spore: Spherical, with four polar capsules. Each shell valve has longitudinal ridges, variable in number (6 ridges are found on the drawing), which run parallel to the sutural line. Four polar capsules of nearly same size and convergent. Sporoplasm finely granular with two nuclei. Dimensions: diameter 10.8μ , length of polar capsule 3.6μ .

CHLOROMYXUM sp. Awerinzew [Fig. 134]

1908 Chloromyxum sp.

Awerinzew

1908: 43, 47, 48

Habitat: Gall-bladder of Raja radiata; Murman coast?.

Vegetative form: Form rounded. The protoplasm is distinctly differentiated into ectoplasm and endoplasm. Ectoplasm hyaline and comparatively abundant in quantity compared with endoplasm, forms lobose pseudopodia of active movements. Endoplasm vacuolated, contains enclosures. Between the two layers, a thin layer of protoplasm, reticular in structure and stained deeply with hematoxylin, is present.

Spore: No figure.

CHLOROMYXUM THYMALLI Lebzelter

1912 Chloromyxum thymalli Lebzelter 1912: 295-296

Habitat: Gall-bladder of Thymallus thymallus L.; Vienna?

Vegetative form: Irregular form, 33 to 35μ long in average. Endoplasm contains fat globules which stain brown with carmine. Trophozoites attached to the epithelium. In average, 6 spores formed in each individual. Intracellular stage in the epithelial cell is supposed. Polysporous.

Spore: Spherical. Shell structure similar to C. protei, but ridges are more developed and exhibit somewhat wavy courses. Polar capsules of equal size. Dimensions: 9 to 9.5μ in diameter, polar capsules 3μ .

CHLOROMYXUM KOI Fujita

[Fig. 135]

1913 Chloromyxum koi

Fujita

1913:257-259

Habitat: Gall-bladder of Cyprinus carpio L.; Sapporo (Nippon).

Vegetative form: Spherical, with greatest diameter up to 50μ , containing 1 to 3 spores. Each spore is situated in a clear space surrounded by a membranous envelope (sporoblast?), around which there is some finely granular matter (endoplasm?).

Spore: Spherical, exhibiting a somewhat angular contour at the anterior end. Shell thick and has well marked ridges on the surface, i.e., 4 to 5 circular ridges and on both sides of these ridges, two more ridges each bent in a loop-like manner, so that the outline of spore in cross section, is very much like of a toothed wheel with nearly equidistant teeth, 16 to 18 in number. Four polar capsules, two slightly larger than the other two. Dimensions: length 16μ , breadth 10μ , length of polar capsule 4μ , length of polar filament 64μ .

CHLOROMYXUM MAGNUM Awerinzew [Figs. 136 to 138]

1913 Chloromyxum magnum Awerinzew 1913: 155-156

Habitat: Gall-bladder of Acanthias blainvillei;* Algoa Bay, East London, Lüderitzbucht (Africa).

Vegetative form: Ameboid. Body yellowish by the presence of large yellowish granules in endoplasm. Often round or rosary form. Pseudopodia sometimes absent, so that the trophozoites move like Amoeba limax with a cluster of small, hairy pseudopodia at the posterior end. In larger form, small round pseudopodia, composed of homogeneous ectoplasm, are formed. Plasmotomy by budding, was often observed. Usually polysporous, rarely monosporous.

Spore: Elongated spherical form. Four polar capsules at the narrow, anterior end. Sporoplasm with two nuclei. Dimensions: length 40 to 48μ , breadth 30 to 38μ , length of polar capsules 12 to 15μ .

* Misprinted in Awerinzew's paper as blainvilei.

CHLOROMYXUM FUNDULI Hahn [Figs. 139 and 140]

1915 Chloromyzum funduli

Hahn

1915:205-206

Habitat: Muscle of Fundulus sp.; Woods Hole. In one fish.

Vegetative form: Hahn made observations on few fresh and stained smears. According to him, it is clear that the staining was abnormal. It is hard to quote this here as he used different terms without giving any definition. The reader is advised to consult Hahn's paper.

Spore: Form slightly resembles that of *Choloromyxum quadratum*. Posterior end rounded, the anterior portion narrow and truncated at the tip; optical cross-section thru the posterior part of the polar capsules, circular. Four polar capsules at the anterior end. Dimensions: height (length) 6μ , breadth and thickness 7.5μ respectively.

Remarks: As to the comparison of the present species with Chloro-myxum clupeidae Hahn, see p. 94.

CHLOROMYXUM MISGURNI Kudo

[Figs. 141 to 146]

1916 Chloromyxum misgurni

Kudo

1916:6-7

Habitat: Gall-bladder of Misgurnus anguillicaudatus Cantor; Tokio (September).

Vegetative form: Round or irregular. Semicircular when viewed from side. From the flat surface, many fine root-like, filiform pseudopodia are extruded. No clear differentiation between ectoplasm and endoplasm. Endoplasm alveolar. Trophozoites always found attached to the lining epithelial cells. Size up to 50μ by 20μ . Polysporous (6 to 8 spores), rarely disporous.

Spore: Spherical, slightly attenuated at the anterior end. Sutural line straight and forms a ridge. Fine longitudinal striations run parallel to the sutural line. Four polar capsules at the anterior end. Sporoplasm finely granular, has two nuclei of equal size. Dimensions: length 8 to 9μ , breadth 6 to 7μ , thickness 5 to 6μ , length of polar capsule 2 to 3μ , of polar filament 28 to 35μ (KOH).

Remarks: The host is often infected at the same time by *Chloromyxum* fujitai, the trophozoites of which can be distinguished from the present form by the structure and the floating habit in the bile. Spores in the two species are decidedly different in form, structure and size.

CHLOROMYXUM FUJITAI Kudo

[Figs. 147 to 152]

1916 Chloromyxum fujitai

Kudo

1916:7-9

Habitat: Gall-bladder of Misgurnus anguillicaudatus Cant.; Tokio, (5% of the fish examined in September, found infected).

Vegetative form: Round or irregular. No clear differentiation of protoplasm. Endoplasm highly vacuolated. Ectoplasm being hardly distinguishable. Size up to 40μ in diameter. Trophozoites float in the bile in almost all cases. Disporous and polysporous (up to 8 spores).

Spore: Spherical, often attenuated at the anterior end. Sutural line not straight. Shell very thick, shows thick ridges running longitudinally on the surface. In optical cross section, the spore presents an outline like a cog-wheel with 20 to 22 ridges. The thickness of ridges varies regularly; the thickest ones being located on two lines where a plane perpendicular to sutural plane cuts the shell longitudinally, others decreasing in thickness as they approach the sutural line. Four polar capsules at the anterior end. Sporoplasm with two nuclei. Dimensions: length 10 to 12μ , breadth 8 to 10μ , polar capsules 2 to 3μ , length of polar filament 23 to 30μ (KOH).

CHLOROMYXUM CLUPEIDAE Hahn [Figs. 153 to 156 and 562 to 565]

1900	Sporozoa	Tyzzer	1900 : 66-68
1901	Sporozoa	Linton	1901:438
1910	Chloromyxum sp.	Auerbach	1910 : 178
1917	Chloromyzum clupeidae	Hahn	1917 : 13-19

Habitat: Body musculature of Clupea harengus, Pomolobus pseudoharengus, P. aestivalis, P. mediocris, Brevoortia tyrannus, Stenotomus chrysops, Tautogolabrus adspersus; Woods Hole.

Tyzzer mentioned in his paper and also in a letter to the writer that he collected the material in August of 1900 and that he found the infection occurred only among young fish. Hahn also called attention on the latter fact.

Vegetative form: Hahn's observations are as follows:

Clusters of spores ("pseudocysts") are spindle-shaped, especially when young, usually lying between the bundles of muscle fibres. Color white or creamy. Larger ones usually "in pocket just beneath the integument." Schizogonic multiplication probably exists. Parasites hard to stain, anilin dyes being unable to stain at all. Large form (probably composed of many individuals) 890μ by 30μ .

Tyzzer described as follows: Cysts up to 1 to 2 mm. in length, lying between the muscle fibres of the myotomes, surrounded at times by membraneous connective tissue. The parasites also occur in diffused infiltration.

Linton found two cysts, 1.74mm. by 1.16mm. and 1.16mm. by 0.58mm. and also diffused state between the fibrillae.

The writer's observation on slides prepared by Dr. Tyzzer* is as follows: Two cysts in sections; one almost spherical, 480μ by 430μ , sur-

*The writer had recently the opportunity of examining the slides of the parasites prepared by Dr. Tyzzer, which occasion he appreciated very much. As a result of this, the writer became convinced of the identity of forms observed by Tyzzer, Linton and Hahn, tho he could not examine the latter authors' specimens.



rounded by several layers (about 10μ thick) of connective tissue of the host, the other oval, 120μ by 110μ . The staining sufficed to reveal only indistinct structure of the parasites. The homogeneous ectoplasm surrounds the entire surface of the body as a uniform, but very thin layer. Endoplasm granular, filled with spores of remarkably identical stages of development. Isolated spores, also, occur in the muscle bundle in the state of diffused infiltration. Polysporous.

Spore: Hahn describes it as follows: Low conical pyramid with round base; square with bulging sides. No indication of valves in the spore shell. Dimensions: height (length) 5μ , breadth and thickness 7μ , polar capsule 2μ by 1μ .

Linton's form: squarish in outline with rounded corners, 7μ in diameter.

Tyzzer describes his form as follows: Quadrilateral in anterior end view; oval in side view. The four corners are a little protuberant and are directed slightly forward. Shape varies considerably in different species of host. The corners of the spore from Stenotomus chrysops, are greatly drawn out, exhibiting stellate form. Four polar capsules radiating from the anterior extremity toward the four corners. Shell shows four furrows radiating from the anterior extremity outwards to the side. Sporoplasm occupies extracapsular cavity. Polar filaments are extruded under the action of acetic acid. Dimension: breadth 7 to 7.5μ .

The writer's observations are as follows:

Spores in fixed and decolorized smears. In smear, most of the spores are seen lying on the base exposing the anterior end view toward the observer's eyes, a few lying with the sutural diameter parallel to the surface of the slide. Form quadrilateral with corners more or less drawn out in anterior end view; oval, with concave posterior side in front view (Figs. 562 to 564). Shell apparently thin but was not clearly separated from the sporoplasm which is finely granular and fills the extracapsular cavity of the spore. Four polar capsules of nearly same size and pyriform. Coiled polar filament indistinct. When stained, the polar capsules stained deeply. It is remarkable to see almost all of the spores exhibit four deeply stained nuclei of capsulogenous cells, which in ordinary case disappear as the spore matures. Dimensions: height (length) 4 to 4.75μ , breadth and thickness 5.4 to 6.5μ , polar capsule about 1.5μ by 0.75μ .

Remarks: Thus the forms of Tyzzer, Linton and Hahn had better be treated as one and the same species. As to the distinction of *Chloromyxum funduli* and the present species, the writer is unable to make it clear as he could not examine the preparation of the former species and especially as he observed some intermediate forms between these two forms in Dr. Tyzzer's preparations of the present species.



CHLOROMYXUM GRANULOSUM Davis [Figs. 157 and 158]

1917 Chloromyzum granulosum Davis 1917: 237

Habitat: Urinary bladder of Tylosurus marianus; Beaufort (July, August).

Vegetative form: Elongated when first placed on the slide, but soon becomes contracted and motionless; progressing by very slow ameboid movements. Ectoplasm usually undistinguishable, being noticed only in a few individuals which had formed one or two short, lobose pseudopodia of hyaline ectoplasm. Body colorless to light yellow. After being on the slide for some time rounded trophozoites often became surrounded by a distinct ectoplasmic layer. Entire body usually coarsely granular, the granules varying greatly in size and shape; sometimes indistinctly vacuolated. Fat globules also present. Size of rounded trophozoites about 30μ . Disporous and polysporous.

Spore: Spherical, with four distinct ridges on the posterior half of each valve converging toward the anterior end. Sutural ridge distinct. Polar capsules pyriform and convergent. Dimensions: diameter 7μ , polar capsules 2μ .

Remarks: Trophozoites from some fish were all colorless, while the larger trophozoites from others were distinctly yellow.

CHLOROMYXUM TRIJUGUM nov. spec. [Figs. 159 to 182]

Habitat: Gall-bladder of Lepomis megalotis Raf.; Stony Creek, and Homer Park, Ill. (November). The parasite was only found in this species, Lepomis humilis and L. cyanellus seined at the same time being free from the infection. Six specimens, three from each of the above mentioned localities, harboured abundantly both free spores and trophozoites of various stages of development in the bile. The fish, from 6.5 to 10.5cm long, were normal in external appearance and the bladders did not show any particular abnormality, compared with those of other fish, as is usually the case.

Vegetative form: Trophozoites float usually free in the bile, younger forms are most frequently attached to the epithelium of the bladder. Form extremely polymorphous, manifesting various shapes such as, almost circular, rounded, oval, elongated or irregular, which is chiefly due to the active extrusion and retraction of the pseudopodia from the body surface. Body is highly transparent and colorless in both the young and the adult. The differentiation of protoplasm into ectoplasm and endoplasm, is distinctly visible in vivo as well as in stained preparations, especially in larger forms (Figs. 159 to 165). The endoplasm presents an alveolar structure without

any enclosure except the nuclei and various stages of spore formation (Figs. 161, 165, 168 to 171), the alveolar network being smaller at the periphery than in the center. The ectoplasm is a hyaline, transparent and homogeneous layer, free from any course granulation in fresh conditions. It shows, however, a very fine reticular structure in stained preparations. The pseudopodia are of two kinds in form, always, formed of ectoplasm alone: the filose and bristle-like form, sometimes branching and protruding from the entire surface or from a localized part of the body, vary in length from 0.5 to 4μ according to the size of the individual (Figs. 159, 161, 164). This form developed, sometimes, into a thicker form with two to four branched finer processes. The blunt, lobose pseudopodium formed at a localized part of the body is well recognizable in larger individuals. Frequently the filose and the lobose pseudopodia are formed on a trophozoite at the same time. The movements of the blunt pseudopodia were striking in some specimens. At the beginning of the observation, ten minues after the bile was removed from the host, two club-shaped pseudopodia (Figs. 161 to 163) which were extruded from a trophozoite, the largest diameter of which being 20µ, moved very actively in the semicircular area changing their forms, showing maximum length of 20µ. In about thirty minutes, they were retracted and from the same place, a short, oval-shaped pseudopodium was seen to be extruded, which remained in the same position for some time without great change of form (Fig. 164). In another case, a trophozoite with a very broad and rounded pseudopodium extruded actively two to three rounded smaller processes at its extremity (Figs. 165 to 167). After fifteen minutes the pseudopodium was retracted, the ectoplasm forming a uniformly thick layer around the endoplasm. The observations were done at room temperature in hanging drop preparations, sealed with vaseline and paraffin, by using comp. oc. 12 and apo. imm. ob. 2mm., which caused no mechanical pressure upon the parasites. The change of form and especially that of pseudopodia, was clearly observed for one hour and twenty minutes under the above mentioned conditions after the bile was removed from the host. The trophozoites when kept for sixteen hours at room temperature, underwent degeneration and disintegrated, setting free the spores which were formed in them.

No active multiplication by plasmotomy, was observed in vivo. In fixed preprations, however, forms that suggested the occurrence of the process in the present myxosporidian, were recognized. As was stated before, the pseudopodia are always formed of the ectoplasm and as each portion of these dividing forms has many nuclei, the author is inclined to record the presence of plasmotomy in the present form.

Size varies greatly. The monosporous form 10μ by 14μ , disporous 15μ by 25μ and polysporous 30μ by 50μ , the largest individual, developing and containing more than 200 spores, was 300μ by 50μ .



Spore: Generally circular in front view; oval in side view. Shell comparatively thick, consequently the coiled polar filament is frequently indistinct. Sutural ridge straight and distinct. Each valve has a thick straight, sometimes slightly zigzag-form ridge that runs parallel to the sutural line, so that in side view, three distinct ridges encircling the spore are recognized (Figs. 177 and 180). From each of these two ridges, eight to twelve short ridges are directed toward the center of each valve, which can distinctly be observed on the spores stained with Heidenhain's iron hematoxylin (Figs. 179 and 180). They can be seen as faint markings rising from the margin directed toward the center of the spore, in front view of fresh spores. Four pyriform polar capsules of slightly different size open their foramina independently at the anterior end of the spore (Figs. 178 and 181). The sporoplasm, granular and finely reticular, shows almost always two nuclei when stained. Dimensions in vivo: length and breadth 8 to 10µ, thickness 5 to 7μ , polar capsules 3 to 5μ by 2 to 3μ , length of polar filament 32 to 40μ (H₂O₂, KOH).

Remarks: In carefully made smear of the bile, a number of empty spores which had been seen in fresh hanging drop preparations, and often spores, in which the sporoplasm with two elongated nuclei seemed to leave the shell (Fig. 182), were recognized. As this particular spore was found close to a thicker mass of the wall of the gall-bladder in the smear, it can hardly be thought that the mechanical pressure during the preparation lead to the mission of the sporoplasm from the spore. It is possible, on the other hand, to think that this is one of the cases of the germination of the spore in the host in which they were developed, as was reported by the author in Nosema bombycis Nägeli (Kudo, 1916).

CHLOROMYXUM CATOSTOMI nov. spec. [Figs. 560 and 561]

Habitat: Gall-bladder of Catostomus commersonii Lac.; Salt Fork, Urbana, Ill. (October). Four fish, from 8 to 14cm.; apparently normal.

Vegetative form: Form usually rounded, with filiform pseudopodia. Majority attached to the epithelium, a few being free in the bile. Body colorless. Protoplasm is not well differentiated. Endoplasm occupying the entire body is of granular structure with vacuoles and refringent spherules. Size: from 15 to 35μ . When kept for 16 hours in a refrigerator, the trophozoites liberated the spores. The number of spores in each trophozoite is usually 2 or 3, rarely 5 to 6. Active plasmotomic multiplication observed when examined. Spores were comparatively small in number, while the trophozoites were attached abundantly to the epithelium of the gall-bladder. Disporous and polysporous.

Spore: Form approximately spherical in front view; oval in profile. Shell with very fine striations which run parallel to the sutural ridge that is fairly well marked. Rounded polar capsules almost of same size, have



independent openings at the anterior end. Coiled polar filament indistinct. Abnormal spores with five polar capsules are sometimes seen. Dimensions of fixed spores: length 8μ , breadth 7μ , thickness 5 to 6μ , polar capsules 2 to 2.5μ by 1.5μ .

CHLOROMYXUM WARDI nov. spec. [Figs. 632 to 642]

Habitat: In the gall-bladder of *Oncorhynchus nerka*: Klutina Lake, Alaska (August). A single gall-bladder collected and preserved in formol by Professor Ward, was found to harbor the present species. The study was done on preserved material and on stained smear preparations.

Vegetative form: Young trophozoites (Fig. 632) show ameboid form, and are mostly multinucleated. The protoplasm is not well differentiated either in unstained material or in stained specimens. It is granulated thruout the body, and is vacuolated at places. The smallest form measured was 18μ in largest diameter. The shape of the body suggests its possession of ameboid movements when alive, altho the writer could not examine fresh specimens. Large trophozoites in which the spore formation had partly been completed are generally rounded with reticular protoplasm. Size varies to some extent. The trophozoite shown in figure 633 contains six mature spores and is 23μ in largest diameter. The largest one found was 38 by 30μ , showing ten spores and nuclei. Each spore appears to develop independently from a single sporoblast. Disporous and polysporous.

Spore: Rounded pyramidal in front view (Figs. 640 and 641); circular in transverse section (Fig. 638). The shell is thickened near the posterior margin (Figs. 640 and 641). Sutural line is not straight, the ridge being fairly distinct. The striations on the shell vary to a considerable extent (Figs. 634 to 637 and 639). Four polar capsules at the anterior end, mostly unequal in size and shape. The coiled polar filament is invisible in formol material. Potassium hydrate solution does not cause its extrusion in the preserved spores. The sporoplasm is finely granular with two nuclei. Dimensions of unstained preserved spores: diameter 7.5 to 9μ , polar capsule 3 by 2.5μ .

Remarks: The writer was able to study forty specimens of gall-bladder of Alaskan fishes, chiefly of salmon, which have been collected by Professor Henry B. Ward, during the summer of 1919, for which he wishes to express his deepest appreciation. The examination of these specimens showed that myxosporidia were found only in one of the gall-bladders, and that specimen presented a fairly heavy infection of the present species.

Family SPHAEROSPORIDAE Davis

1917 Sphaerosporidae Davis 1917: 219

The characters of the family are described on page 57.



Genus SPHAEROSPORA Thélohan

1892 Sphaerospora Thélohan 1892:167

The characters of the genus are described on page 57. Type species: Sphaerospora divergens Thélohan.

SPHAEROSPORA DIVERGENS Thélohan

[Figs. 183 to 186]

1895	Sphaerospora divergens	Thélohan	1895 : 339-340
1912	Sphaerospora divergens	Parisi	1912:289
1912	Sphaerospora divergens	Auerbach	1912 : 41 -4 2

Habitat: Urinary tubules of kidney of Blennius pholis L., Crenilabrus melops L., C. pavo Cuv. et V, and urinary bladder of Hippoglossoides limandoides; Concarneau, Roscoff, Napoli (July), Smalfjorden.

Vegetative form: Rounded discoidal or spherical or more or less elongate. Ectoplasm transparent, without real pseudopodium. Movements extremely slow. Endoplasm, granular, contains fat globules and small yellowish granules. Size of sporulating individuals: 65μ by 55μ , 60μ by 25μ , 60μ by 20μ , etc. Polysporous (Thélohan); monosporous, and disporous (Auerbach).

Spore: Spherical. Shell with fine striations. Two polar capsules divergent; coiled polar filament visible in fresh state. Sporoplasm fills the extracapsular cavity of the spore. Dimensions: 10μ in diameter, often 10μ by 12μ , the larger diameter coinciding with sutural plane, thickness 8μ (Auerbach), polar capsules about 4μ long, length of polar filament 20 to 25μ .

SPHAEROSPORA ELEGANS Thélohan [Figs. 187 and 188]

(8		
	Thélohan	1890 : 193-209
Sphaerospora elegans	Thélohan	1892 : 167-175
Chloromyxum (Sphaerospora	:)	
elegans	. Gurley	1894:266
Sphaerospora elegans	Thélohan	1895 : 338-339
Sphaerospora elegans*	Auerbach	1909a:71
Sphaerospora elegans	Parisi	1912:289
	Chloromyxum (Sphaerospora elegans Sphaerospora elegans Sphaerospora elegans*	Sphaerospora elegans Thélohan Chloromyxum (Sphaerospora) elegans Gurley Sphaerospora elegans Thélohan Sphaerospora elegans* Auerbach

Habitat: Renal tubules of kidney, connective tissue of ovary and urinary bladder of Gasterosteus aculeatus L., G. pungitus L., Lota vulgaris Cuv., Phoxinus laevis L.; Paris, Bretagne, Karlsruhe, Lake Garda.

Vegetative form: Rounded or slightly elongated, not exceeding 20 to 25μ in diameter. Protoplasm homogeneous, very finely granular, contains numerous refractive globules, probably of fatty nature. Pseudopodia lobose. Movements slow. Disporous.

^{*}Misprinted as Sphaeromyxa elegans.

Spore: Spherical, somewhat attenuated at the anterior end. Sutural ridge present, terminating in a small projection at each end of the spore. Two polar capsules spherical. Coiled polar filament not visible in fresh state. Dimensions: diameter 10μ in average, sutural diameter about 11μ .

SPHAEROSPORA ROSTRATA Thélohan [Fig. 189]

1895 Sphaerospora rostrata

Thélohan 1895: 339

Habitat: Malpighian bodies of kidney of Mugil sp.; Roscoff, Le Croisic, Le Vivier-sur-mer, Marseille, Banyuls.

Vegetative form: Not described.

Sphore: Subspherical. Shell shows deep longitudinal striations which end in sharp spinous edges at the posterior end. Sutural ridge well marked. Anterior part shows enlargement of quadrangular lamella, which is spinous in side view. Dimensions: 10 to 12μ in diameter, sutural diameter 1 to 2μ longer, length of polar filament 40μ .

Remarks: The parasites cause the degeneration of the Malpighian bodies.

SPHAEROSPORA MASOVICA Cohn

[Figs. 190 to 192]

1902 Sphaerospora masovica

Cohn 1

1902:628-632

Habitat: Gall-bladder of Abramis brama L.; Mauersee.

Vegetative form: Polymorphous, due to active movements. Transparent and colorless, while in motion. Endoplasm highly granular, contains yellowish enclosures. Ectoplasm hyaline, forms a narrow layer around the body, occasionally developing into a blunt lobose pseudopodium. Pseudopodia of two kinds; lobose and filose, also intermediate forms. Filiform pseudopodia are formed and retracted more slowly than the lobose. Plasmotomy is of probable occurrence. Two spores are formed in each pansporoblast. Size variable: 10μ (with no spore), 18μ (with sporoblasts), 29μ (with 4 sporoblasts), 38μ (with 22 sporoblasts). Disporous(?), polysporous.

Spore: Spherical. Sutural ridge well marked. Polar capsules and sporoplasm are comparatively small, the former convergent. By warming the spore, polar filament is extruded and at the same time two filaments ("starren Fäden") are made visible at the anterior part of the sutural plane. Sporoplasm with two nuclei, no vacuole being present. Dimensions: diameter 8μ , length of polar filament 38μ , length of sutural filament 14μ .

Remarks: Cohn did not observe free spores in the gall-bladder. He, however, saw many free spores, separated from each other, in the intestine, concluding that the body and pansporoblast membrane of trophozoites, are destroyed in the intestine, setting the spores free.

SPHAEROSPORA PLATESSAE Woodcock [Figs. 193 and 194]

1904 Sphaerospora platessae

Woodcock

1904:59-60

Habitat: Otic-capsule of Pleuronectes platessa L.; England.

Vegetative form: Cysts opaque masses about 1mm. in diameter. The cartilage was greatly hypertrophied. Polysporous (presumably).

Spore: Spherical. Shell unornamented. Two polar capsules. Sporoplasm with several refractive granules, but without any vacuole. Dimensions: diameter 8 to 9μ , length of polar filament about 70μ .

Remarks: Woodcock placed this species provisionally in the genus as he could not examine any fresh material, but had studied smears only.

SPHAEROSPORA ANGULATA Fujita

[Figs. 195 to 197]

1912 Sphaerospora angulata

Fuiita

1912:261-262

Habitat: Kidney of Cyprinus carpio L., Carassius auratus L.; Sapporo (Nippon).

Vegetative form: Only description: "The number of the spore in the sporoblast is in this case always less than in the others, rarely exceeding two."

Spore: Somewhat triangular, with convex sides, oval in sideview. Slightly pointed at the mid-posterior margin of the spore. Shell very thin, faintly marked with concentric striations. Two oblong polar capsules are of unequal size. Dimensions: length 7 to 8μ , breadth 6 to 7μ , thickness 5μ , length of largest polar capsule 3.8μ , length of polar filament twice as long as that of the spore.

SPHAEROSPORA POLYMORPHA Davis

[Figs. 198 and 199]

1917 Sphaerospora polymorpha

Davis

1917:231-232

Habitat: Urinary bladder of Opsanus tau; Beaufort (June, July).

Vegetative form: Elongate, but never very irregular in shape. Slowly ameboid. Body colorless. Ectoplasm clearly seen in younger forms, forming one to several large lobate pseudopodia, which in turn extrude several short, conical pseudopodia. In larger forms, ectoplasm is, often, recognizable only at ends of pseudopodia, which in such cases are composed chiefly of endoplasm. Endoplasm granular, vacuolated in some smaller forms, but in larger individuals vacuoles are indistinct or absent; small fat globules abundant in large forms; numbers of rounded sporoblast cells can be distinctly seen. Size of large trophozoites 35μ by 50μ . Disporous and polysporous (polysporous forms rarely contain many spores at the same time).

Spore: Spherical, sometimes slightly compressed infero-superiorly. Sutural ridge; on each side are a number of concentric striations extending around each valve parallel to sutural line. Polar capsules pyriform and large. Coiled polar filaments indistinct. Sporoplasm finely granular. Dimensions: diameter 7 to 10μ , (8μ in average), polar capsules 4 to 5μ by 2 to 2.5μ .

SPHAEROSPORA sp. Davis

1917 Sphaerospora sp.

Davis

1917:213

Habitat: Urinary bladder of Lepisosteus platystomus; Gainesville, Fla.

Vegetative form: No description.

Spore: Not described.

SPHAEROSPORA sp. Southwell et Prashad

1918 Sphaerospora sp.

Southwell and

Prashad

1918 : 347-348

Habitat: Under the scales of Barilius barna; from the vicinity of the Ruby Mines, Burma (June).

Vegetative form: The cysts occurred in very large numbers, one under each scale.

Spore: Authors' description: "The poor condition of the material did not allow of a complete account of its structure, but the bicapsulate, rounded structure of its spores places it undoubtedly in the genus Sphaerospora Thélohan."

SPHAEROSPORA CARASSII nov. spec. [Figs. 200 to 204]

Habitat: Gill filament of Carassius carassius L.; Tokio (February).

Vegetative form: Trophozoites small ameboid in groups or in diffused condition in the connective tissue of the gill filament. No cyst formation. The number of trophozoites in groups is generally small. The largest group found in sections was 96μ by 36μ , the macroscopical examination always failing to trace the parasites. The trophozoites, 10 to 20μ long, with poorly differentiated protoplasm and usually reticular endoplasm without any particular enclosure (Fig. 200). Ameboid movements not observed. Schizogonic multiplication rapid, each of the daughter individuals developing into two spores. Disporous. Other sporous characters could not be determined.

Spore: Spherical in front and side views, the form variable to some extent (Figs. 201-203). Shell smooth. Sutural ridge fairly distinct. Two polar capsules, broadly pyriform, of equal size and convergent, located at

the anterior end, one on each side of the sutural plane. Coiled polar filament highly distinct (5 to 6 times) in vivo. Sporoplasm granular, shows two nuclei when stained; no vacuole of any nature. Dimensions in vivo: diameter 8 to 13μ , polar capsules 4 to 5μ by 2.5 to 3.5μ , length of polar filament 35 to 40μ (KOH or pressure).

Remarks: No species of the genus, has ever been found in the branchiae. The characters of the spore, however, compel the writer to place the form in the present genus.

Genus SINUOLINEA Davis

1917 Sinuolinea

Davis

1917:219

The characters of the genus are described on page 57.

Type species: Sinuolinea dimorpha Davis.

SINUOLINEA DIMORPHA Davis [Figs. 205 to 213]

 1916
 Sphaerospora dimorpha
 Davis
 1916: 333-377

 1917
 Sinuolinea dimorpha
 Davis
 1917: 232-233

Habitat: Urinary bladder and ureter of Cynoscion regalis; Beaufort.

Vegetative form: Disporous and polysporous trophozoites differ distinctly from each other. Disporous trophozoites irregular, colorless, transparent and show slow movements. When attached to the epithelium, rounded with one to several pseudopodia. Differentiation of protoplasm distinct. Occasionally endoplasm contains one or more erythrocytes. Average diameter of full-grown form 25 to 30μ .

Polysporous form: when attached to the bladder epithelium, the free end is drawn out into a long, cylindrical process, covered with numerous short, hairlike ectoplasmic processes. While not movable, these processes are readily absorbed and reformed. When the trophozoite is detached from the epithelium, the larger end gives rise to numerous conical or arborescent pseudopodia, by means of which the trophozoite moves slowly. Endoplasm extends into the proximal portion of large pseudopodia. It is granular and vacuolated, contains numerous fat globules, refractive granules, yellowish crystals (hematoidin?) and erythrocytes in various stages of disintegration. Endoplasm also contains gemmules, each composed of outer layer and finely granular central portion. Size varies greatly: up to 575μ by 90μ .

Spore: Spherical. Sutural ridge well marked. Polar capsules large and spherical. Sporoplasm forms a rounded granular mass. Dimensions: diameter 15μ , diameter of polar capsules 4.5μ , length of polar filament 27 to 35μ .

SINUOLINEA CAPSULARIS Davis

[Figs. 214 to 216]

1917 Sinuolinea capsularis

Davis

1917:233

Habitat: Urinary bladder of Paralichthys albiguttus, P. dentatus, Spheroides maculatus; Beaufort (July, August).

Vegetative form: Rounded to irregular shape. Body colorless or light yellow. Progressive movements slow. Pseudopodia large branched or arborescent, formed entirely of ectoplasm. Ectoplasm transparent and usually granular, merging gradually with the endoplasm. Endoplasm contains numberous fat globules. In large trophozoites, gemmules are observed. The gemmules are more finely granular and more transparent than the surrounding protoplasm and are practically identical with the small, free trophozoites. Trophozoites containing several gemmules are usually rounded and motionless and appear to be more or less degenerate. Disintegration of such trophozoites were actually observed. Sporulating trophozoites were rare and were never seen to contain gemmules. Size up to 40μ in diameter. Disporous and polysporous(?).

Spore: Spherical, sometimes slightly elongated. Sutural plane much twisted on its axis. Sutural ridge very distinct. Polar capsules and capsulogenous cells large occupying more than one-half of the cavity of spore. Coiled polar filament distinct. Sporoplasm granular contains numerous fat globules. Dimensions: diameter 12 to 14μ , diameter of polar capsules 4.5μ , length of polar filament 50μ .

SINUOLINEA ARBORESCENS Davis

[Figs. 217 and 218]

1917 Sinuolinea arborescens

Davis

1917:233

Habitat: Urinary bladder of Siphostoma floridae; Beaufort

Vegetative form: Rounded or irregular. Body colorless or light yellow. Actively ameboid, forming large arborescent pseudopodia of ectoplasm. Ectoplasm well developed, hyaline and homogeneous. Endoplasm coarsely granular, sometimes containing a few fat globules. Larger trophozoites are less active and the ectoplasm less distinct. In sporulating trophozoites the ectoplasm may entirely disappear, the entire trophozoite consisting of a coarsely granular mass. Diameter of rounded sporulating trophozoites 75μ . Polysporous.

Spore: Rounded, in front view, slightly elongated in the anterior end view. Polar capsules large. Sutural ridge prominent, makes a characteristic S-shaped turn on the anterior end. Coiled polar filaments distinct. Dimensions: length 15μ , breadth 12μ , diameter of polar capsules 5μ .

SINUOLINEA OPACITA Davis [Fig. 219]

1917 Sinuolinea opacita

Davis

1917:234

Habitat: Urinary bladder of *Paralichthys albiguttus*; Beaufort (August). Vegetative form: Rounded or slightly irregular. Body colorless and opaque. Movements slow. Pseudopodia short lobose. Ectoplasm not distinct, except around ends of pseudopodia, where it forms a thin hyaline layer. Endoplasm opaque, finely granular, with numerous greenish-yellow fat globules varying greatly in size. Diameter of rounded sporulating trophozoites 22μ, exceptionally large trophozoites 100μ. Disporous.

Spore: Nearly spherical, with flattened, lateral appendages extending from the posterior side. Sutural plane slightly twisted on its axis. Sutural ridge distinct. Polar capsules large. Coiled polar filament distinct. Sporoplasm finely granular, containing several comparatively large fat globules. Dimensions: diameter 12 to 13μ , diameter of polar capsules 4μ .

SINUOLINEA BRACHIOPHORA Davis [Fig. 220]

1917 Sinuolinea brachiophora

Davis

1917:234

Habitat: Urinary bladder of *Paralichthys albiguttus*; Beaufort (August only in one fish).

Vegetative form: Rounded to somewhat irregular. Body colorless. Ectoplasm hyaline. Endoplasm granular, with numerous large fat globules. Disporous.

Spore: Nearly spherical, with a long lateral appendage from each valve. These appendages are empty except at extreme distal end, which contains a granular mass, probably the remains of the parietal cell. Sutural plane slightly oblique to longitudinal axis. Sutural ridge distinct. Polar capsules and capsulogenous cells large, occupying more than half of cavity of spore. Sporoplasm finely granular. Dimensions: length exclusive of appendages 9 to 11μ , length of appendages 18 to 22μ , breadth of spore 9μ , diameter of polar capsules 3.5μ .

Remarks: Davis mentions that in many respects this species is very similar to S. opacita, which occurs in the same host.

Suborder PLATYSPOREA nom. nov.

The definition of the suborder is recorded on page 57.

Family MYXIDIIDAE Thélohan

 1892
 Myxidides
 Thélohan
 1892: 173, 175

 1893
 Myxidiidae
 Gurley
 1893: 412

The characters of the family are described on page 57.

Genus MYXIDIUM Bütschli

1882 Myxidium

Bütschli

1882: Pl. 38

The characters of the genus are described on page 58. Type species: Myxidium lieberkühni Bütschli.

MYXIDIUM LIEBERKÜHNI Bütschli [Figs. 221 to 240]

1854		Lieberkühn	1854 : 5-6, 349
1879		Leuckart	1879 : 246
1881		Bütschli	1881 : 638-648
1882	Myxidium lieberkühni	Bütschli	1882 : 593-595
1883	•	Balbiani	1883 : 201-202, 274-275
1891	Myxidium lieberkühni	Pfeiffer	1891 : 20, 91, 105, 127
1894	Myxidium lieberkühni	Gurley	1894 : 283-289
1895	Myxidium lieberkühni	Thélohan	1895 : 3 4 0
1895	Myxidium lieberkühni	Cohn	1895 : 5-36
1898	Myxidium lieberkühni	Doflein	1898 : 229, 341
1902	Myxidium lieberkühni	Prenant	1902a : 200-217
1902	Myxidium lieberkühni	Laveran and	
		Mesnil	1902 : 4 69-4 72
1906	Myxidium lieberkühni	Léger and He	sse 1906: 720
1909	Myxidium lieberkühni	Auerbach	1909a:71
1912	Myxidium lieberkühni	Schröder	1912:326-327
1912	Myxidium lieberkühni	Parisi	1912:286
1916	Myxidium lieberkühni	Mavor	1916a : 66-68
1916	Myxidium lieberkühni	Mavor	1916b : 373-378
	-		

Habitat: Urinary bladder of Esox lucius L., Lota lota L. (L. vulgaris); France, Canada (Georgian Bay), U. S. A., (Wisconsin, Lake Mendota), Italy (Lago Maggiore, Lago di Como, Milano), Germany.

Vegetative form: Form variable with lobose or immovable filiform pseudopodia. Clear differentiation of protoplasm. Cohn described third layer of protoplasm (mesoplasm). Endoplasm yellowish in older trophozoites, contains yellow globules, fat globules and hematoidin crystals. Size varying with age up to a maximum length of 300μ by a breadth of 136μ (Bütschli). Plasmotomous multiplication active. Cohn described budding of larger forms, while Laveran et Mesnil observed only the division of smaller forms. Each pansporoblast develops into two spores. Polysporous.

Spore: Elongated fusiform. Shell with longitudinal striations. Polar capsule at each end of the spore. The longer axis of polar capsules coincides with that of spore. Dimensions: length 18 to 20μ , width 5 to 6μ . Mavor's measurement: polar capsules 5μ by 2.5 to 3μ , length of polar filament 40 to 45μ .

MYXIDIUM INCURVATUM Thélohan

[Figs. 241 to 251]

1892	Myxidium?incurvatum	Thélohan	1892a: 1093-1094
1895	Myxidium incurvatum	Thélohan	1895 : 341
1912	Myxidium incurvatum	Parisi	1912:286-287
1912	Myxidium incurvatum	Auerbach	1912:4,39
1916	Myxidium incurvatum	Georgévitch	1916 : 90 -9 1
1917	Myxidium incurvatum	Davis	1917 : 234-235

Habitat: Gall-bladder of Nerophis aequoreus L., N. annulatus, N. lumbriciformis, Blennius pholis L., Callionymus lyra L., Fundulus majalis, Gambusia affinis, Hippocampus brevirostris, Mugil cephalus, Scorpaena scrofa L., Syngnathus acus L., S. typhle; Roscoff, Concarneau, Marseille, Banyuls, Napoli, Bergen, Monaco, Beaufort (July).

Vegetative form: Thélohan describes as follows: Trophozoites usually small, sometimes reaching a considerable size. Pseudopodia lobose. Protoplasm pale and finely granular with refractive globules. Disporous. According to Parisi and Davis rarely monosporous. Georgévitch observed apparently the polysporous form.

Parisi's form: ectoplasm hyaline, endoplasm granular. Monosporous form 25μ long.

Davis's form: lobose pseudopodia, occasionally being drawn out into a long process. Many trophozoites often cling together closely. Diameter of rounded disporous forms about 13 to 15μ , that of monosporous forms about 10 to 11μ .

Spore: Thélohan's description is as follows: Irregular fusiform. Longest axis curved into S-form, both ends sharply pointed and directed toward opposite directions. Polar capsule opening on opposite side of the spore, in some spores the axis of the polar capsules being parallel to each other. Dimensions: length 8 to 9μ , breadth 4 to 5μ , length of polar filament 10 to 15μ .

Parisi gave the following dimensions: length 10 to 12μ , breadth 5 to 6μ , length of polar capsule 3μ , length of polar filament 28μ .

According to Georgévitch, young spores are not curved (Fig. 245).

Davis's form; Polar filaments when extruded in HCl remained tightly coiled. Dimensions: length 8 to 9μ , width 5 to 6μ , diameter of polar capsule about 3μ .

Remarks: As are shown in figures, Davis's form seems to be somewhat different from the European forms.



MYXIDIUM SPHAERICUM Thélohan [Fig. 252]

1895 Myxidium sphericum (corr.

sphaericum) T

Thélohan 1895: 341-342

Habitat: Gall-bladder of Belone acus (Belone belone L.); Banyuls, Le Vivier-sur-Mer.

Vegetative form: Trophozoites spherical or subspherical, not exceeding 20 to 22μ in diameter with lobose pseudopodia formed from the entire surface. Endoplasm granular, contains small refractive granules. Disporous.

Spore: Form similar to M. incurvatum, but much greater. Coiled polar filament distinctly visible in fresh spore. Dimensions: length 15 to 20μ , width 7 to 8μ , length of polar filament 60μ (KOH).

MYXIDIUM HISTOPHILUM Thélohan

[Fig. 253]

1895 Myxidium histophilum Thélohan 1895: 341

Habitat: Connective tissue of kidney and ovary of Leuciscus phoxinus L. (Phoxinus laevis Ag.); France.

Vegetative form: Small mass.

Spore: Fusiform, being compressed at the middle part. Shell with longitudinal striations. Length of the spore 15μ .

MYXIDIUM sp. Gurley [Fig. 254]

1851		Leydig	1851 : 226, 234
1852		Leuckart	1852:436
1894	Myxidium? sp. incert.	Gurley	1894:290
1899	Myxidium sp.	Labbé	1899:92

Habitat: Gall-duct of Raja batis L. Vegetative form: No description. Spore: Not described. One figure.

MYXIDIUM DANILEWSKYI Laveran

[Figs. 255 to 257]

1887		Danilewsky	1887:35
1897	Myxidium danilewskyi	Laveran	1897 : 725-726
1898	M vxidium danilewskvi	Laveran	1898 : 27-30

Habitat: Kidney of Emys orbicularis L.; France.

Vegetative form: Form elongated, circular in cross-section, tapering toward the ends. Body of greenish color, occupying the lumen of the renal tubules of the kidney. Body bent along the cavity of the tubule. Endo-

plasm granular, ectoplasm covering the entire surface of the body as a thin layer. Each pansporoblast develops two spores. Polysporous.

Spore: Elongated fusiform, similar to M. lieberkühni, but much smaller. Polar capsule at each end, extrudes filament under the action of nitric acid. Sporoplasm granular with one nucleus. Dimensions: length 12μ , breadth 3 to 4μ .

MYXIDIUM GIGANTEUM Doflein

[Fig. 258]

1898 Myxidium giganteum

Doflein

1898: 285-286

Habitat: Gall-bladder of Raja asterias; Napoli.

Vegetative form: Rounded trophozoites. Lobose pseudopodia with slow movement, show remarkable dimensions. Posterior portion forms "Stemm-pseudopodien." Small form club-shaped. Endoplasm is of yellowish color. Diameter of large form 500μ , of medium sized 200μ , small individuals $70-90\mu$, quite young ones, polymorphous 8 to 40μ . Larger individual up to 700μ by 180μ . Many trophozoites form a cyst-like motionless stage, in which many individuals seem to be covered with a common gelatinous envelope. Each pansporoblast forms two spores. Polysporous.

Spore: Elongated. Fusiform in front view; in side view, one valve arch-form, the other being flat. Transparent. Two polar capsules, one at each end. Coiled polar filament is clearly seen in larger polar capsules. Dimensions: length 28μ , breadth 8μ , polar capsules 8μ by 4μ .

MYXIDIUM BARBATULAE Cépède

1906 Myzidium barbatulae Cépède 1906 : 67 1906 Myzidium barbatulae Cépède 1906a : 15-16

Habitat: Kidney of Cobitis barbatula L.; Isère.

Vegetative form: Trophozoites form cysts. Form and size vary greatly. Average size: 400 to 500 μ in length and 200 μ in breadth.

Spore: Irregular fusiform. Polar capsule at each end of the spore. Shell longitudinally striated, number being variable. Dimensions: length 12 to 15μ , breadth about 6μ , polar capsules 5μ by 2.5 to 3μ .

MYXIDIUM GIARDI Cépède

[Figs. 259 to 261]

 1906
 Myxidium giardi
 Cépède
 1906a : 16; 1906b : 170-173

 1908
 Myxidium giardi
 Cépède
 1908 : 93-95

 1908
 Myxidium giardi
 Cépède
 1908a : 8

Habitat: Kidney of Anguilla vulgaris Flem.; Wimereux (August).

Vegetative form: Subspherical white cysts, 800 to 900μ in diameter, surrounded by a thick (up to 30μ) membrane, composed of the connective tissue of the host.

Spore: Irregular fusiform, greatly enlarged at the middle portion. Plane of symmetry of the spore coincides with the sutural plane. Shell thick with 9 to 11 longitudinal striations on each valve, which are more clearly seen on spores stained with iron hematoxylin. Polar capsule at each end. Coiled polar filament distinct. Sporoplasm finely granular with two nuclei and refringent globules. Dimensions in vivo: length 9 to 10μ , width 5 to 5.6μ , thickness 4.75 to 5μ , polar capsules 3.5μ by 2μ .

MYXIDIUM PFEIFFERI Auerbach

[Figs. 262 to 265]

1908	Myxidium pfeifferi	Auerbach	1908 : 459-464
1910	Myxidium þfeifferi	Auerbach	1910c : 171-172

Habitat: Gall-bladder of Tinca vulgaris Cuv.; Karlsruhe.

Vegetative form: Observations in sections. More or less flattened, disc-form, often enrolled. The ectoplasm finely granular, without large pseudopodia. It is not usually distinguishable from the endoplasm which is highly alveolar and contains numerous nuclei, but no enclosures.

Spore: Form varies to some extent. Similar to Myxidium lieberkühni; slightly curved. Shell with fine longitudinal striations. Polar capsules two, one at each end. Polar filament is extruded by adding one drop of water to the smear of the spore, which had been dessicated for 24 hours. Sporoplasm with one or two nuclei, in one case with four small nuclei, which is thought to be an abnormal. Dimensions: length 13 to 18μ , breadth 5.2 to 5.8μ , length of polar capsule 5.2 to 6μ , length of polar filament 45 to 54μ .

MYXIDIUM INFLATUM Auerbach

[Fig. 266]

1909	Myxidium inflatum	Auerbach	1909 : 72-74
1909	Myxidium inflatum	Auerbach	1909a: 31
1910	Myxidium inflatum	Auerbach	1910c : 172
1912	Myxidium inflatum	Auerbach	1912:39

Habitat: Gall-bladder of Cyclopterus lumpus L.; Bergen (September). Vegetative form: Extremely polymorphous. Rounded, spherical, or much elongated. Ameboid movements very active. Differentiation of protoplasm is sharp and clear, which is best observed in individuals in motion; highly hyaline ectoplasm forms very long lobose pseudopodia, into which granular endoplasm flows in slowly. Size variable. Rounded large form 44 to 45 μ in diameter. Fully grown spores are set free from the mother trophozoite in comparatively short time. Spore formation similar to that of Myxidium bergense. Disporous and polysporous (5 spores in maximum).

Spore: Very broad compared with the length. The longitudinal axis is curved in S shape. Polar capsule situated in opposite way at each end of the spore. Dimensions: length 20.8 to 23.4μ , breadth 13 to 15.6μ , polar capsules 7.8μ , length of polar filament 90 to 100μ (KOH).

MYXIDIUM BERGENSE Auerbach [Fig. 267]

1908		Keysselitz	1908:289
1909	? Myxidium sphaericum	Auerbach	1909 : 75-76
1910	Myxidium bergense	Auerbach	1910 : 61
1910	Myxidium bergense	Auerbach	1910c : 172
1912	Myxidium bergense	Auerbach	1912 : 18-39
1915	M vzidium bersense	Mayor	1915 : 30-31

Habitat: Gall-bladder of Gadus virens L., G. aeglefinis, G. merlangus, Pleuronectes platessa and Sebastes viviparus, Melanogrammus aeglefinis; Norway (Bergen), Canada (St. Andrew, July to September).

Vegetative form: Rounded or elongated, as the result of formation of various pseudopodia. Trophozoites partly free, partly attached to the epithelium of the bladder. Size up to 54μ in diameter. Pseudopodia of two kinds: lobose and long filose, sometimes slightly branched. Mavor observed a cyst-like stage under certain conditions, which, he thinks, may be due to some exceptional conditions of the parasite. Plasmogamy. Monosporous, disporous and polysporous.

Spore: Fusiform. Main axis curved into S shape. Form, roughly speaking, very much similar to that of M. sphaericum Thél. Dimensions: length 16.2 to 19μ , breadth 7 to 9μ , length of polar capsules 5.4μ , length of polar filament about three times as that of spore.

MYXIDIUM PROCERUM Auerbach [Fig. 268]

1910	Myxidium procerum	Auerbach	1910 : 61 -62
1910	Myxidium procerum	Auerbach	1910c : 172-173
1912	Myxidium procerum	Auerbach	1912:4.39

Habitat: Gall-bladder of Argentina silus As.; Bergen.

Vegetative form: Not observed.

Spore: Greatly elongated and narrow. Sporoplasm with one or two nuclei. Dimensions: length 21.6 to 25.2 μ , breadth 3.6 to 4 μ , length of polar capsule 7.2 μ .

MYXIDIUM MACKIEI Bosanquet [Figs. 269 to 271]

1910 Myxidium mackiei Bosanquet 1910: 436-438

Habitat: Renal tubules of kidney of Trionyx ganzeticus; Bombay. Observations on three slides.

Vegetative form: The largest trophozoite 160μ by 27μ . No distinction between ectoplasm and endoplasm could be drawn, except in a few individuals in which there was a cyst-wall. Spores are formed in pairs. Protoplasm with two kinds of nuclei, some vesicular, others smaller and compact. Polysporous.

Spore: Fusiform with rather pointed ends. Shell finely striated. Two comparatively small polar capsules, one at each end. Sporoplasm with one or two nuclei, contains, often, two large vacuoles. Dimensions: length 16μ (a few 17μ), breadth 5μ (many broader than this).

Remarks: The discoverer, J. P. Mackie mentioned that the parasites did not appear to excite any reaction in the tissue of the host, the animal's health being unaffected.

MYXIDIUM MACROCAPSULARE Auerbach

[Figs. 272 and 273]

1910 Myxidium macrocapsulare Auerbach 1910: 440-441

Habitat: Gall-bladder of Scardinius erythrophthalmus L.; Karlsruhe. Vegetative form: Not observed.

Spore: Elongated elliptical when viewed at right angles to sutural plane. Shell somewhat thick with longitudinal striations parallel to the sutural line. In side-view, both ends pointed in diagonally opposite directions. Polar capsules are comparatively large, one at each end, opening at the sharply pointed end. Dimensions: length 10 to 12μ , breadth 6μ , polar capsules 3 to 4μ .

Remarks: No pathological change. Bile was clear.

MYXIDIUM sp. Awerinzew [Figs. 274 to 276]

1908	Myxidium sp.	Awerinzew	1908 : 33, 43, 45, 55
1909	Myxidium sp.	Awerinzew	1909 : 76, 78, 80, 81
1911	Myxidium sp.	Awerinzew	1911:199-204

Habitat: Gall-bladder of Cottus scorpius; Aleksandrowsk, North Sea. Vegetative form: Trophozoites are small. The protoplasm is differentiated into ectoplasm and endoplasm in some specimens. Very active formation of filiform pseudopodia of various length. Degenerating trophozoites, with one or two empty spaces are often noticed. Each spore is formed independently from each other. Monosporous, disporous and polysporous (with three spores).

Spore: Form similar to *Myxidium incurvatum*. Young spores not curved. Dimensions: length 20 to 35μ , breadth 10 to 15μ .

MYXIDIUM DEPRESSUM Parisi [Figs. 277 and 278]

1912 Myxidium depressum

Parisi

1912:287

Habitat: Gall-bladder of Citharus linguatula Gthr.; Napoli (August). Vegetative form: Not observed.

Spore: Fusiform with greatly attenuated extremities in front view; flattened and curved in S-form in profile. The axis of polar capsules parallel to each other. Coiled polar filament visible *in vivo*. Sporoplasm with two nuclei, occupies the extracapsular cavity of the spore. Dimensions: length 12 to 14μ , breadth 5.5 to 6μ , thickness 2.5 to 3μ , polar capsules 5.5 to 6μ by 2.3μ , length of polar filament 30μ .

MYXIDIUM OVIFORME Parisi

[Figs. 279 and 280]

 1912
 Myxidium oviforme
 Parisi
 1912: 287-288

 1912
 Myxidium oviforme
 Auerbach
 1912: 39

Habitat: Gall-bladder of Apogon rex mullorum Cuv., Coris julis Gthr., Gadus callarias L., Trutta salar L.; Napoli (August), Norwegian coast.

Vegetative form: Unobserved by Parisi. Auerbach's observation is as follows:

Trophozoites, small ameboid, usually spherical. Size 10 to 12μ in diameter. Monosporous (probably).

Spore: Oval with rounded extremities, slightly pointed at the foramina of polar capsules. Shell with numerous fine striations running longitudinally. Polar capsules being often invisible, opening a little above and below of the hypothetical horizontal plane. Sporoplasm fills the extracapsular cavity of the spore, leaving little space at the extremities of the polar capsules. Dimensions: length 11μ , breadth 8 to 8.5μ , polar capsules 4.5μ by 3μ , length of polar filament 30 to 35μ . Auerbach's measurements: length 12 to 13μ , breadth 8 to 9μ , polar capsules about 4μ long.

MYXIDIUM ANGUILLAE Ishii

[Figs. 281 to 284]

1915 Myxidium anguillas

Ishii

1915:372-382

Habitat: Integument of the side of the body of Anguilla japonica Temm. et Sch.; Schizuoka, Nippon (October). Number of the cysts visible to unaided eye, 10 and 9 on the left and the right side respectively.

Vegetative form: Trophozoites form white and sharply contoured cysts. Cysts, spherical or oval, surrounded by a membranous connective tissue (about 2μ thick) of the host. Protoplasm is clearly differentiated into ectoplasm and endoplasm. Diffuse infiltration also occurs. Size measured along the skin, 1.2 to 2mm. in diameter; in sections 1.174mm. by 0.658mm.

Spore: Form similar to *Myxidium pfeifferi* Auerbach, but rather straight fusiform, rarely slightly bent. In many spores the shell tapers to a sharp point at each end. Shell striated longitudinally, 22 in all (2 sutural ridges?). Two polar capsules, one at each end. Sporoplasm usually with two nuclei. Dimensions: length 9.1μ , breadth 2.8μ , length of polar capsule 3.5μ .

MYXIDIUM sp. Mavor [Figs. 285 and 286]

1915 Myxidium sp.

Mavor

1915: 32

Habitat: Gall-bladder of *Pseudopleuronectes americanus*; New Brunswick (Canada), of rare occurrence.

Vegetative form: Observations in smears are as follows: Spheroidal, with numerous long pseudopodia on one side, which suggests the attachment of the trophozoite to the bladder. Trophozoites without any spore. Pansporoblasts spherical, 15 to 16μ in diameter.

Spore: Spindle shaped. The long axis being slighly bent in S-form. Two pear shaped polar capsules, one at each end of spore. Coiled polar filament visible in fresh state. Dimensions: length 14 to 15μ , breadth 6 to 7.5μ , polar capsules 4μ by 2.5μ , length of polar filament 90 to 95μ (ammonia water).

MYXIDIUM GADI Georgévitch [Figs. 287 to 290]

1916	Myxidium gadi	Georgévitch	1916 : 88-89
1917	Myxidium gadi	Georgévitch	1917c : 797-799
1919	Mvzidium zadi	Georgévitch	1919:251-289

Habitat: Gall-bladder of Gadus pollachius, Solea vulgaris Quens; Roscoff (September).

Vegetative form: Highly polymorphous. Spherical or oval. Large forms fill up the bladder. Ectoplasm hyaline and transparent, forming one long or many short lobose pseudopodia. Endoplasm colorless and finely granular, contains more or less large numbers of nucleus. Monosporous, disporous and polysporous.

Spore: Fusiform with attenuated ends. Young spores more attenuated than the fully grown forms. The main axis of the spore coincides with the longitudinal axis of the polar capsules, with slight deviation. Two nuclei of the sporoplasm, are always smaller than those of the shell-valves or of polar capsules. Dimensions: length 6 (?) to 14μ , breadth 4 to 6μ .

MYXIDIUM GLUTINOSUM Davis

[Fig. 291]

1917 Myxidium glutinosum Davis 1917: 235

Habitat: Gall-bladder of Cynoscion regalis; Beaufort.

Vegetative form: Elongated or irregular. Slowly ameboid, moving by means of a broad, lobose pseudopodium of hyaline ectoplasm. Body colorless. Ectoplasm only distinct in pseudopodium. Endoplasm finely granular. The mature spores while still within the mother trophozoites, are surrounded by a clear, refractive gelatinous envelope. Diameter of rounded sporulating trophozoites 20μ . Disporous.

Spore: Cylindrical, ends of valves rounded except at one side, where the polar capsules open at the apex of a small, conical elevation. Spore characterized by the presence of a transparent, homogeneous, gelatinous envelope. Polar capsules pyriform, opening on each side nearly at right angles to the longitudinal axis. Dimensions: length 10 to 11μ , breadth 6μ , length of polar capsules 3μ .

MYXIDIUM PHYLLIUM Davis

[Figs. 292 and 293]

1917 Myxidium phyllium Davis 1917: 235

Habitat: Gall-bladder of Gambusia affinis; Beaufort.

Vegetative form: Exceptionally large; flattened, leaflike, usually folded on itself; motionless. Pseudopodia were not observed. Ectoplasm forming a distinct transparent layer around entire body. After being on slide for some time ectoplasm usually becomes covered with very numerous, short, hairlike processes. Endoplasm finely granular, contains numerous fat globules. Diameter up to 1.35mm. Polysporous.

Spore: Fusiform, slightly truncated at each end where polar capsules open. Shell with numerous longitudinal striations. Sporoplasm finely granular, with several small fat globules. Dimensions: length 11μ , breadth 8μ , diameter of polar capsules 3μ .

MYXIDIUM STRIATUM Cunha et Fonseca

1917 Myxidium striatus Cunha et Fonseca 1917: 321

Habitat: Gall-bladder of Menticirrhus americanus L., Bairdiella ronchus Cuv. et Val.; Brazil.

Vegetative form: More or less spherical. Body small and colorless. Endoplasm granular. Ectoplasm visible when pseudopodia are formed. Pseudopodia filiform, being projected radially. Size variable, 16μ in diameter in average.

Spore: Elliptical. Shell with fine longitudinal striations which run parallel to sutural line. Sutural plane oblique to the longitudinal axis of the spore which is thickened at the extremities. Two ovoidal polar capsules, one at each end. Dimensions: length 10 to 14μ , breadth 6 to 8μ , length of polar capsules 4μ , length of polar filament 30μ .

MYXIDIUM KAGAYAMAI nov. spec. [Figs. 294 and 295]

1916 Myxidium sp.

Kudo

1916:6

Habitat: Gall-bladder of Misgurnus anguillicaudatus Cant.; Tokio (September), 2% of the fish examined infected.

Vegetative form: Not observed.

Spore: Fusiform; one valve being more convex than the other. Suture line straight. Shell with fine longitudinal striations. Dimensions in fixed preparations: length 15 to 18μ , breadth 6 to 7μ , length of polar capsules 7 to 8μ , length of polar filament 60 to 70μ .

Remarks: Tho the vegetative form is still unobserved, the author is compelled to consider the present form as a new species by careful reexamination of the material and proposes the name in honor of Dr. T. Kagayama, Tokio, Nippon.

MYXIDIUM AMERICANUM nov. spec.

[Figs. 622 to 627]

Habitat: In the lumen of urinary tubules of the kidney of *Trionyx spinifera*; Crystal Lake, Urbana, Ill. (July). A single host specimen showed a light infection in the above mentioned organ. No intracellular stage was detected.

Vegetative form: The young trophozoite in the lumen of the tubule of the kidney is multinucleate, and more or less irregular in shape which suggests the ameboid movements of the animal (Figs. 622, 623). The older form with mature spores is rather spherical in form with a distinct outline. The protoplasm is fairly well differentiated into ectoplasm and endoplasm (Fig. 624). The size of the trophozoites varies from 12 to 25μ in diameter. A pansporoblast produces two spores. Polysporous.

Spore: Spindle-form; with the two pointed extremities stretched in opposite directions. Circular in cross-section. The shell is rather thin; sutural line is straight. Fine longitudinal striations on the shell, eight to ten in number on each valve. The polar capsules are nearly spherical, coiled polar filament being visible in fresh material (three turns). The polar filament is easily extruded from the fresh spores under the influence of potassium hydrate solution. The direction of the extruded polar filament forms an angle of about 45° with the main axis of the spore and the two filaments are parallel to each other. Preserved spores do not show any filament extrusion under the influence of the said chemical. The sporoplasm is finely granular, and shows, upon staining, two small nuclei of ring-shape, as their peripheral layer takes stain more deeply than the central portion. Average dimensions of fresh spores: length 15 to 16μ , breadth 5.5 to 6μ , polar capsule 4μ by 3.5μ , length of polar filament 25 to 32μ .



Remarks: Two species of the genus Myxidium were reported to occur in chelonian hosts; i.e., M. danilewskyi (page 109) and M. mackiei (page 112). The former differs from the present form in having an elongated vegetative form which is greenish in color, and in having spores of different shape, dimensions and structure, not to speak of the difference of the host. The latter resembles closely to the species under consideration in dimensions of the spores, but differences in the trophozoite and in the structure of the spore do not allow one to consider two forms as identical. The species is therefore treated as new.

Genus SPHAEROMYXA Thélohan

1892 Sphaeromyxa Thélohan 1892a : 1091-1093

The characters of the genus are described on page 58.

Type species: Sphaeromyxa balbianii Thélohan.

SPHAEROMYXA BALBIANII Thélohan

[Figs. 296 to 307]

1892	Sphaeromyxa balbianii	Thélohan	1892a: 1091-1093
1895	Sphaeromyxa balbianii	Thélohan	1895:342
1912	Sphaeromyxa balbianii	Parisi	1912:288
1916	Sphaeromyxa balbianii	Georgévitch	1916 : 92 -9 3
1917	Sphaeromyxa balbianii	Davis	1917 : 235-236

Habitat: Gall-bladder of Motella tricirrata Bl., M. maculata Risso., Cepola rubescens L., Clupea pilchardus, Siphostoma floridae, S. louisianae; Roscoff (September), Concarneau, Marseille, Banyuls, Napoli (September), Beaufort (June to August).

Vegetative form: Flattened leaf-like or disc-form, reaching 3 to 4mm. in diameter. Often forms spherical with opaque appearance. The protoplasm is distinctly differentiated into endoplasm and ectoplasm. Ectoplasm forms rounded lobes which exhibit slow movements and show a clear radially striated structure in sections. Endoplasm reticular, contains nuclei, young and mature spores and fat globules. Each pansporoblast develops two spores. Polysporous.

Davis mentions that the largest form he observed was 900μ in diameter. Georgévitch recognized a large number of small trophozoites which were formed by repeated plasmotomous multiplication.

Spore: Fusiform, with truncate ends. Shell longitudinally striated. One polar capsule at each end. Polar filament is wound around an imaginary axis perpendicular to the longitudinal axis of the spore. When extruded, the polar filament is seen as a short, conical and hollow thread-like structure. Sporoplasm finely granular with two nuclei. Dimensions: length 15μ , width 5μ , length of polar filament 15μ .

Parisi gave the following dimensions: length 15 to 20μ , width 5 to 6μ , polar capsule 7μ by 4.7μ , length of polar filament 25 to 30μ .

Davis' measurements are as follows: length 17 to 20μ , breadth 5 to 6μ , length of polar filament 20μ .

Georgévitch observed young spores with both ends tapering into a point. Later they assume the typical form with truncated ends. He did not recognize the striations on the shell. He also mentions the occurrence of abnormal spores, such as elliptical, spherical forms, etc.; or with only one polar capsule.

SPHAEROMYXA IMMERSA (Lutz) Thélohan [Figs. 308 to 311]

1889	Cystodiscus immersus	Lutz	1889 : 84-88
1895	Sphaeromyxa immersa	Thélohan	1895:343
1899	Cystodiscus immersus	Lühe	1899 : 291-293

Habitat: Gall-bladder of Bufo marinus L. and Leptodactylus occilatus L.; Brazil.

Vegetative form: Leaf-like or disc form, visible thru the bladder wall. Upper and lower sides slightly convex. Size up to 1.5 or 2mm. in diameter; thickness being 1/20 to 1/10 of the diameter. Protoplasm is well differentiated. Ectoplasm transparent and membranous, often contains a large number of micrococcus-like bodies. No ameboid movements nor change of form. Endoplasm highly vacuolar, contains fat globules. Plasmotomic multiplication probably occurs. Spores always arranged in pairs. Polysporous.

Spore: Oval with rounded extremities. Shell more or less thick, with fine transverse striations. Spherical polar capsule at each end. Sutural plane is oblique to the longitudinal axis of the spore. Sporoplasm transparent. Dimensions: length 12 to 14μ , breadth 9 to 10μ , length of polar filament 50 to 70μ (4 to 5 times that of the spore) (KOH).

SPHAEROMYXA INCURVATA Doflein [Figs. 312 to 314]

1898 Sphaeromyxa incurvata Doflein 1898 : 286-287

Habitat: Gall-bladder of Blennius ocellatus; Napoli.

Vegetative form: Trophozoites are found in large masses (Plasmodia?), in which they form a thin, hollow ball, 5-7mm. in diameter.

As the surface is greater than the inner surface of the bladder, some parts of the body are folded up. Body bluish white and transparent. Protoplasm highly vacuolar, contains numerous fat globules, nuclei and spores. Polysporous.

Spore: Curved to one side along sutural plane and also in a plane at right angles to it. Polar capsule at each end. Sporoplasm with two nuclei.

Polar filament is wound along the longer diameter of the capsule, and relatively thick, but thinner than that of S. balbianii Thél. Dimensions: length (along the inner side of the arch) 30 to 35μ , breadth 8μ , distance between two polar capsules 12 to 15μ , polar capsules 12 to 15μ by 4 to 5μ .

SPHAEROMYXA SABRAZESI Laveran et Mesnil [Figs. 315 and 322]

1900	Sphaeromyxa sabrasesi	Laveran et	
	•	Mesnil	1900 : 380-382
1906	Sphaeromyxa sabrazesi	Schröder	1906:455-466
1907	Sphaeromyxa labrazesi*	Schröder	1907 : 359-381
1910	Sphaeromyxa sabrazesi	Schröder	1910 : 1-5
1912	Sphaeromyxa sabrazesi	Parisi	1912:288
1913	Sphaeromyxa sabrazesi	Jameson	1913:2
1916	Sphaeromyxa sabrazesi	Georgévitch	1916 : 91 -9 2
1916	Sphaeromyxa sabra s esi	Georgévitch	1916a:3

Habitat: Gall-bladder of Hippocampus brevirostris Cuv., H. guttulatus Cuv.; Syngnathus acus, Motella tricirrata, Nerophis annulatus, Siphonostoma rondeletii; Arcachon, Rovigno, Napoli, Roscoff (September), Monaco, Villefrance, (March to June).

Vegetative form: Disc form. Diameter up to 2mm. Thickness variable. Body whitish in color. Ectoplasm thin, transparent and homogeneous. Young trophozoites may probably have lobose pseudopodia. Endoplasm highly vacuolated, contains nuclei of various sizes, pansporoblasts, spores and more or less refringent granules. Polysporous.

Schröder observed larger forms up to 5mm. Ectoplasm also was found to project numerous fine short (1μ) hair-like processes from the surface. Each pansporoblast develops into two spores.

Spore: Cylindrical, bent in arch form; with truncated ends. Large cylindrical polar capsule at each end. Sporoplasm granular, contains one nucleus. Polar filament short and conical, is extruded under the action of nitric acid. Dimensions: length 28μ , width 4.3μ , polar capsule 9 to 10μ by 3μ , distance between the polar capsules 8μ , length of polar filament 8μ .

Schröder noticed the stained sporoplasm contained one or two nuclei. He observed indistinctly marked longitudinal striations on the shell. Dimensions: length 22 to 25μ , breadth 3 to 4μ , polar capsule 8μ by 2 to 3μ , length of polar filament about 12μ .

Georgévitch described the presence of a hyaline substance, containing pale granules, in the spore cavity. Young spores were found to take the form of Myxidium type. In mature spores, he always found two nuclei by staining.

^{*}Misprinted in Schröder's paper.

SPHAEROMYXA HELLANDI Auerbach

[Figs. 323 and 324]

1909	Sphaeromyxa hellandi	Auerbach	1909 : 78–79
1910	Sphaeromyxa hellandi	Auerbach	1910ь : 772-774
1910	Sphaeromyxa hellandi	Auerbach	1910c : 174-175
1912	Sphaeromyxa hellandi	Auerbach	1912 : 4, 4 0

Habitat: Gall-bladder of Molva vulgaris Flem., Centronotus gunellus, Brosmius brosme Asc.; Bergen, Torghatten.

Vegetative form: Large and rounded disc form. Protoplasm is distinctly differentiated into ectoplasm and endoplasm. Thickness up to 160μ , folded in the bladder. Ectoplasm finely granular; in unstained specimens, it is recognizable as 10 to 12μ thick layer, in which about 2μ thick radially striated outer layer and 8 to 10μ thick inner finely granular region can be distinguished. In stained sections, the outer layer remains unstained. Endoplasm highly alveolar, contains refractive granules of different size which are not stained with Sudan III. Each pansporoblast develops into two spores. Polysporous.

Spore: Arch form in front view. The degree of curvature varies greatly. Sutural line curved in S-shape and well marked. Both ends more or less truncated. Polar capsule at each end. Polar filament being wound along the longest axis of the polar capsule and is extruded with KOH. Sporoplasm rounded with one or two nuclei. Dimensions: length 20.8 to 26μ , breadth and thickness 5.4μ , length of polar capsule 10 to 10.8μ .

SPHAEROMYXA EXNERI Awerinzew [Figs. 325 and 326]

1913 Sphaeromyza exneri Awerinzew 1913a: 155

Habitat: Gall-bladder of Thysanophris japonicus; Lorenço Marques (Africa).

Vegetative form: Not observed.

Spore: Somewhat resembles that of *S. hellandi* Auer. in being bent to one side with sutural line of S-form but differs in dimensions. Both ends slightly tapering. Polar capsules two, one at each blunt end, in which the polar filament is wound parallel to its longer axis. Sporoplasm comparatively small and sharp-contoured, contains only one nucleus. Dimensions: length 75 to 80μ , breadth 18 to 20μ , length of polar capsule 30 to 35μ .

SPHAEROMYXA GASTEROSTEI Georgévitch [Fig. 327]

1916 Sphaeromyxa gasterostei Georgévitch 1916:88

Habitat: Gall-bladder of Gasterosteus spinachia; Roscoff (September). Vegetative form: Trophozoites form large plasmodia.

Spore: Large, elongated fusiform; ends less truncated than those of the spore of *Sphaeromyxa balbianii*. As the spore becomes mature, the ends assume more pointed shapes. Polar capsules two, one at each end. Sporoplasm with two nuclei, fills the extracapsular cavity. Dimensions: twice or three times larger than those of *Sphaeromyxa balbianii* Thélohan.

Genus ZSCHOKKELLA Auerbach

1910 Zschokkella

Auerbach

1910:62-63 1910a:240-256

1910c:175

The characters of the genus are described on page 58.

Type species: Zschokkella hildae Auerbach.

ZSCHOKKELLA HILDAE Auerbach

[Figs. 328 to 331]

 1910
 Zschokkella hildae
 Auerbach
 1910 : 62-63

 1910
 Zschokkella hildae
 Auerbach
 1910a : 240-254

 1912
 Zschokkella hildae
 Auerbach
 1912 : 40-41

Habitat: Urinary bladder of Phycis blennioides Br., Gadus aeglefinis, G. callarias L., G. virens L.; Norway.

Vegetative form: Trophozoites float in the bile or attach themselves to the epithelial layer of the bladder. Youngest ameboid form about 4.5 to 6μ . In floating form, pseudopodia more or less long, lobose, are formed; while in the attached form those similar to the pseudopodia of *Myxidium bergense* Auer. are developed. Plasmogamy occurs. Size varies greatly according to the number of spores which are formed in each individual. Monosporous (with or without the remnant of protoplasm), disporous and polysporous (up to 4 spores).

Spore: Semicircular in front view, with slightly and equally attenuated ends. At each end, large spherical polar capsule is situated which opens not at the extremity, but on the flat surface. Shell bivalve and thick. Sutural line S-form. Sporoplasm with two nuclei. Dimensions: length 16 to 28.8μ , breadth 13 to 18μ , polar capsules 5.6 to 7.2μ in diameter, length of polar filament 72μ (KOH).

ZSCHOKKELLA NOVA Klokačewa [Figs. 332 and 333]

1914 Zschokkella nova

Klokacewa

1914:184-186

Habitat: Gall-bladder of Carassius vulgaris; Russia?

Vegetative form: Not observed.

Spore: Outline irregular. Observations on fixed materials alone. Two large round polar capsules open at the side near ends. Sporoplasm with two nuclei. On some spores, striations that run parallel to the sutural line were observed. Dimensions: length 9.5 to 11.5μ , breadth 6.5 to 7μ , diameter of polar capsule 3 to 3.5μ .

ZSCHOKKELLA ACHEILOGNATHI Kudo [Figs. 334 to 338]

1916 Zschokkella acheilognathi

Kudo

1916:3-5

Habitat: Gall-bladder and gall-duct of Acheilognathus lanceolatum Temm. et Schl.; Tokio (May). Over 80% of the fish examined were found to be infected.

Vegetative form: Disc-shape. In bile duct, large trophozoites are folded up. Body colorless and transparent. Protoplasm is well defined into two regions. Ectoplasm finely granular in vivo. In stained sections, it shows two layers; thin outer layer (2μ thick) presents very fine striations, while inner layer (6 to 8μ thick) is finely vacuolated without any enclosure. Endoplasm is highly vacuolated. Lobose pseudopodia formed only in younger individuals (15 to 30μ in diameter), in which ameboid movements are not slow. Size: up to 720μ by 550μ , thickness 5 to 30μ . Polysporous.

Spore: Form resembles Zschokkella hildae, but slightly elongated. Form varies to some extent. Some spores are of Myxidium type. Sutural line curved. Longitudinally striated. Spherical polar capsule at each end opening near the extremity. Dimensions: length 10 to 14μ , breadth 6 to 7μ , diameter of polar capsule 3 to 5μ , length of polar filament 65 to 70μ (KOH).

ZSCHOKKELLA GLOBULOSA Davis

[Figs. 339 and 340]

1917 Zschokkella globulosa

Davis

1917:236

Habitat: Urinary bladder of *Spheroides maculatus*; Beaufort (August). Vegetative form: Rounded; slowly ameboid, forming short, lobose pseudopodia. Body colorless and transparent. Ectoplasm not distinct. Protoplasm granular, characterized by the presence of several large fat globules. Sporulating trophozoites about 15 to 16μ in diameter. Monosporous and disporous.

Spore: Semicircular. Sutural line twisted on its axis and oblique to longitudinal axis; sutural ridge distinct. Polar capsules opening on flat surface. Sporoplasm finely granular and very transparent. Dimensions: length 11μ , breadth 7μ , diameter of polar capsules 3μ .

Family MYXOSOMATIDAE Poche

1913 Myxosomatidae

Poche

1913 : 230

The characters of the family are described on page 58.

Genus MYXOSOMA Thélohan

1892 Myxosoma

Thélohan

1892:175

The characters of the genus are described on page 58.

Type species: Myxosoma dujardini Thélohan.

MYXOSOMA DUJARDINI Thélohan

[Figs. 341 to 343]

1841		M üller	1841 :486-487
1845		Dujardin	1845 : 644
1892	Myxosoma dujardini	Thélohan	1892:175
1895	Myxosoma dujardini	Thélohan	1895 : 343-344
1905	Myxosoma dujardini	Nufer	1905 : 77, 79, 186
1910	Myxosoma dujardini	Wegener	1910 : 72-7 3
1916	?Myxosoma dujardini	Kudo	1916 : 3

Habitat: Branchial lamellae of Scardinius erythrophthalmus L., Perca luviatilis, Leuciscus rutilus L. and Cyprinus carpio L.; France, Frisches Haff, Kurisches Haff (February, April, May), Tokio (May), Switzerland.

Vegetative form: White cysts being branched, rounded, spherical or irregular; 1 to 1.5mm. in diameter.

Wegener's form 1 to 1.7mm. long.

Spore: Ovoidal, flattened, with attenuated anterior end which is slightly bent laterally. Two pyriform polar capsules at the anterior end. Sporoplasm without any iodinophilous vacuole. Dimensions: length 12 to 13μ , breadth 7 to 8μ .

Wegener's form: polar capsules 6μ by 3μ .

Kudo's form: polar capsules 6 to 7μ by 2μ , length of polar filament 70μ .

MYXOSOMA (?) LOBATUM Nemeczek

[Fig. 348]

1911 Myxosoma (?) lobatum Nemeczek 1911 : 160-162

Habitat: Branchiae of Leuciscus leuciscus L. and Aspius rapax Ag.; Austria.

Vegetative form: Cysts spherical, oval or elongated; of white color. Size from 0.5 to 3mm. by 0.5 to 1mm. Those in Aspius rapax, oval to spindle-shape, 1 to 3mm. long and 1 to 1.5mm. wide.

Spore: Ovoidal; anterior end narrowly pointed and straight; posterior end rounded, with lobose appendix (about 6μ long). A transverse fold on the shell behind the polar capsules in fresh as well as preserved spores. No iodinophilous vacuole. Dimensions: length 12.6μ , breadth 8.2μ , length of polar capsule 4.2μ , length of polar filament 80 to 90μ . Spores found in Aspius rapax, had slight difference in dimensions, the structure, however, being similar to the above.

Remarks: Nemeczek doubts if this form is really Myxosoma because of the following: 1) different shape of the cysts compared with that of the type species as described by Thélohan; 2) spores observed might develop later into other forms like Henneguya.

MYXOSOMA FUNDULI Kudo [Figs. 344 to 347]

1918 Myzosoma funduli Kudo 1918: 12-14

Habitat: Branchiae of Fundulus majalis Wal. and F. heteroclitus L; Woods Hole (August, September).

Vegetative form: Cysts. Spherical and small; 150μ in average diameter. Largest form observed 360μ by 264μ . Spores, young and mature, were found in the cysts. Polysporous.

Spore: Pyriform. Shell uniformly thick with 7 to 10 folds on sutural edge at the posterior portion. Sutural ridge, fairly well marked. Two polar capsules pyriform and of equal size at the anterior end. Sporoplasm finely granular with two nuclei but without any iodinophilous vacuole. Dimensions: length 14μ , breadth 8μ , thickness 6μ , polar capsule 8μ by 2μ , length of polar filament 38 to 42μ (perhydrol, KOH).

Remarks: The writer could not find any evidence of an iodinophilous vacuole by treatment with various iodine mixtures, which is the most important characteristic of the genus. Hahn's form (Myxobolus funduli, p. 151) should be distinguished from the present form.

Genus LENTOSPORA Plehn

1905 Lentospora Plehn 1905: 150

The characters of the genus are described on page 58. Type species: Lentos pora cerebralis (Hofer) Plehn.

LENTOSPORA CEREBRALIS (Hofer) Plehn [Figs. 349 to 354]

1903	Myxobolus cerebralis	Hofer	1903:8
1904	Myxobolus chondrophagus	Hofer	1904:53
1905	Lentos por a cerebralis	Plehn	1905 : 145-166
1909	Lentospora cerebralis	Plehn	1909:38
1910	Lentospora cerebralis	Auerbach	1910c : 176

Habitat: Cartilage and perichondrium of Trutta iridea Gibb., Salmo fontinalis Mitch., Trutta salar L.; Germany (Karlsruhe and other localities).

Vegetative form: Ameboid form. Size varies greatly. Small ameboid form probably grows up into large individual which has often fifty or more ringform nuclei and breaks up into numerous small forms by division. No sporous character is observed except a figure of a disporous form.

Spore: Circular in front view; lenticular in side view, with more or less extensive variation in length and breadth. Shell smooth. Sutural ridge distinctly thickened. Two polar capsules pyriform and convergent, are usually of same size. Extruded polar filaments cross each other. Sporoplasm with two ring-form nuclei but without any iodinophilous vacuole.

Dimensions: diameter 6 to 10μ , length of polar capsule 2/5 that of the spore, length of polar filament 40 to 50μ (limewater, 1% KOH).

Remarks: Plehn noticed that the present form causes the chronic form of "Drehkrankheit" among young fish in German waters. She was unable to extrude the polar filament with mineral (?) acids. Auerbach, however, could extrude the filament by means of acids.

LENTOSPORA MULTIPLICATA Reuss [Fig. 355]

1906 Lentos por a multiplicata Reuss 1906 : 203

Habitat: Muscle of Idus melanotus Heck.; Volga?, Russia.

Vegetative form: Not described.

Spore: Oval. Sutural edge broad with many folds. No iodinophilous vacuole. Dimensions: length 12μ , breadth 9.5μ , thickness 6μ , polar capsules 4μ by 2.25μ .

LENTOSPORA ENCEPHALINA Mulsow

1911 Lentospora encephalina Mulsow 1911: 483-485

Habitat: In the blood vessel of the brain, especially of the mid-brain of Cyprinus carpio L.; Munich (spring). Blood vessels are the only seat of infection. In most cases many individuals lie parallel to one another. The infection occurs frequently and heavily. The effect, however, is undetermined.

Vegetative form: Trophozoite elongated, worm-like and circular in cross section. The body is covered with a pellicula. The protoplasm is distinctly differentiated into homogeneous ectoplasm layer and inner endoplasm. In the latter are found numerous granules, small nuclei and spores.

Spore: Almost circular in front view; profile? No iodinophilous vacuole is found. The polar filament is easily extruded by means of a highly diluted KOH solution. Diameter: 5 to 5.5μ .

LENTOSPORA ASYMMETRICA Parisi [Figs. 356 to 359]

1912 Lentos pora asymmetrica Parisi 1912: 292-293

Habitat: Connective tissue of kidney of Crenilabrus pavo C. et V.; Napoli (September).

Vegetative form: One trophozoite found; a small, rounded form with thin and hyaline ectoplasm which could be distinguished from the endoplasm with coarse yellowish globules, containing two spores. Disporous?

Spore: Oval from the front; flattened and fusiform in profile. Sutural edge with many triangular folds, which are more clearly seen in material

preserved in formalin than in fresh condition. Two polar capsules of same size, are situated asymmetrically, opening at the side near the anterior end. Sporoplasm granular and with two nuclei, but without any iodinophilous vacuole. The polar filament not being extruded by ordinary reagents, probably because the spores were not full-grown. Dimensions: length 10 to 11μ , breadth 6.5 to 7μ , length of polar capsules 5μ .

LENTOSPORA ACUTA (Fujita) Kudo [Figs. 360 to 362]

1912 Sphaerospora acuta

Fujita

1912:260-261

Habitat: Epithelium of branchial lamellae of Carassius auratus L.; Sapporo, Nippon.

Vegetative form: Fujita's description is simply as follows: Sporoblast contains about two spores.

Spore: Spherical in front view, with slightly pointed anterior end; spindle shaped in side-views. Shell thin and smooth. Two convergent polar capsules are of different sizes, occupying about 5/8 in space of the spore. No vacuole could be made out in sporoplasm. Dimensions: length 8 to 10μ , breadth 7 to 8μ , thickness 5 to 6μ , polar capsules 5μ by 4μ .

Remarks: This species, recorded incompletely by Fujita as Sphaerospora, shows characters of the genus *Lentospora* in spore form so that it is provisionally placed here.

LENTOSPORA DERMATOBIA Ishii [Figs. 594 to 596]

1916 Lentospora dermatobia

Ishii

1916:472-474

Habitat: In the integument of Anguilla japonica Temm. et Schl.; Shizuoka, Nippon. From the same specimen which harboured Myxidium anguillae, see page 114. The number of cysts reaches probably "several hundreds."

Vegetative form: Cysts, beneath the epidermis, usually subcircular, more or less irregularly triangular or quadrilateral under the magnifier, with the largest diameter of from 142 to 267μ . The epidermis is slightly lifted up by the cyst. No chromatophore on the surface of the cyst. The cysts separated from each other, are found mostly in the central region of the body, head and fins being free from cysts. In cross-section, cysts exhibit oval or lenticular shape with the longest diameter, which is twice as long as the depth, placed parallel to the surface of the skin. No particular pathological change was noticed.

Spore: Circular in front view; broad fusiform or lenticular in side view. Sutural ridge fairly distinct. Sutural edge comparatively broad, especially

at the posterior margin, where a few folds (three are figured) are seen. Two oval polar capsules convergent and of equal size. Sporoplasm is sharply contoured, no iodinophilous vacuole being recognized. Dimensions in preserved material (?): diameter 6.3 to 7μ , thickness 4.2 to 4.9μ , length of polar capsule 2.8 to 3.5μ .

Family MYXOBOLIDAE Thélohan

1892	M yxobolidées	Thélohan	1892 : 173, 176
1893	M yxobolidae	Gurley	1893 : 412, 413
1895	M yxobolidées	Thélohan	1895 : 347

The characters of the family are described on page 58.

Genus MYXOBOLUS Bütschli

1882	M vxobolus	Bütschli	1882 : Pl. 38 : 6-10
1004	MI YAUUUNNA	Duiscun	1002 . 11. 30 . 0-10

The characters of the genus are described on page 58.

Type species: Myxobolus mülleri Bütschli.

MYXOBOLUS MÜLLERI Bütschli

[Figs. 397 to 403]

1881		Bütschli	1881 : 630
1882	Myxobolus mülleri	Bütschli	1882:595
1895	Myxobolus mülleri	Thélohan	1895 : 349
1905	Myxobolus mülleri	Nufer	1905': 77, 79, 186
1906	Myxobolus mülleri	Cépède	1906 : 64-65
1906	Myxobolus mülleri	Schröder	1906:195
1908	Myxobolus mülleri	Auerbach	1908:456
1909	Myxobolus mülleri	Auerbach	1909a: 54, 71
1910	Myxobolus mülleri	Wegener	1910 : 81
1911	Myxobolus mülleri	Nemeczek	1911:160
1912	Myxobolus mülleri	Parisi	1912:293

Habitat: Air bladder and branchiae of Leuciscus cephalus L.; kidney and ovary of L. phoxinus L.; eye of Crenilabrus melops L. and Alburnus lucidus; branchiae of Aspro asper L., Barbus vulgaris Flem., Leuciscus rutilus L., Squalius cephalus L., S. agassizii Heckel, Lota vulgaris L., Phoxinus loevis Ag.; pseudobranchiae of Cottus gobio L.; intestine of Mugil auratus Risso.; France, Germany [Karlsruhe, Alle (October), Pregel, Frisches Haff], Switzerland (Neuchatel Lake), Italy (Napoli, September).

Vegetative form: White cysts in the connective tissue. Form elongated oval, 2 to 3mm. in diameter. No clear differentiation of protoplasm is observed even in young forms. In sections, some cysts show radiate striations in the thick granule-free ectoplasm. Endoplasm filled with nuclei.

Cépède writes as follows: Cysts in branchiae, subspherical or elliptical, 1.5mm. by 0.5mm.

Wegener's form: Cysts small and rounded, 0.2 to 0.3mm. in diameter. Spore: Ordinarily spherical or subspherical. Two polar capsules

with a small triangular intercapsular appendix. Polar capsules pyriform and of same size. Sutural edge exhibits folds (7 to 9).

Thélohan's dimensions: length 10 to 12μ , breadth 9 to 11μ , length of polar capsule 5μ .

Cépède gave the following dimensions in vivo: length 10μ , breadth 9μ , thickness 6μ , length of polar capsule 5μ .

Wegener's form. Usually oval, often almost spherical. Length 10 to 11μ , breadth 8 to 9μ , diameter of spherical form 9μ , polar capsule 4 to 5μ by 2 to 3μ .

MYXOBOLUS PIRIFORMIS Thélohan [Figs. 363 to 364]

1852		Remak	1852 : 144
1883		Balbiani	1883 : 197-198
1884		Balbiani	1884:125
1891		Pfeiffer	1891:132
1892	Myxobolus piriformis	Thélohan	1892:177
1893	Myxobolus piriformis	Gurley	1893:414
1894	Myxobolus piriformis	Gurley	1894:211
1895	Myxobolus piriformis	Thélohan	1895 : 348
1905	Myxobolus piriformis	Nufer	1905 : 77, 186
1910	Myxobolus piriformis	Plehn	1910 : 22-27
1910	Myxobolus piriformis	Wegener	1910 : 73

Habitat: Branchiae, spleen, kidney of *Tinca tinca* L., *Cobitis fossilis* L. and subcutaneous connective tissue, spleen, liver, connective tissue of the intestine of *Leuciscus* sp.; France, Germany (Pregel), Switzerland.

Vegetative form: Small, long thread-like cysts. Color white. Polysporous.

Wegener's form: average size, length 1mm., breadth 0.09 to 0.1mm Spore: Elongated oval; flattened. Anterior end highly attenuated and slightly bent to one side. One pyriform polar capsule at this end. Dimensions: length 16 to 18μ , breadth 7 to 8μ , length of polar filament 30μ .

Wegener gives the following dimensions: length 18μ , breadth 7.5μ , polar capsule 7.5μ by 3.5μ .

MYXOBOLUS UNICAPSULATUS Gurley

[Figs. 365 to 366]

1841		Müller	1841:487
1893	Myxobolus unicapsulatus	Gurley	1893:414
1894	Myxobolus unicapsulatus	Gurley	1894 : 210-211

Habitat: In the skin of Labeo niloticus For.; Nile.

Vegetative form: Cysts very small pustules in the skin of the head.

Spore: Form similar to Myxosoma dujardini. A single polar capsule at the anterior end, obliquely directed. Dimensions: length 0.0051", breadth 0.0034".

MYXOBOLUS FUHRMANNI Auerbach [Fig. 367]

1909Myzobolus fuhrmanniAuerbach1909:65-681910Myzobolus fuhrmanniAuerbach1910c:178-179

Habitat: Connective tissue under the mucous membrane of the mouth of Leuciscus rutilus L.; Neuchatel Lake.

Vegetative form: Cysts, of pea-size, surrounded by several membranous layers of connective tissue with a few nuclei. Finely granular ectoplasm forms outer layer. Endoplasm is dense and contains faintly stained nuclei. Pansporoblasts and spores are found in the central portion of the cyst. Polysporous.

Spore: Elongated pyriform, with attenuated anterior and rounded posterior ends. Majority with a single polar capsule; spores with two polar capsules were also observed. Shell thick, at the posterior end. 4 to 6 notch-like markings on the posterior part of the shell. Sutural ridge thickened and fairly well marked. Coiled polar filament visible in preserved material. The opening of the polar capsule is either at the anterior end or near it. Sporoplasm with two nuclei of unequal size and a comparatively large iodinophilous vacuole, stained brown with iodine alcohol. Dimensions: length 18 to 20μ , breadth about 8μ , thickness 6μ , length of polar capsule 9 to 10μ .

MYXOBOLUS OCULI-LEUCISCI Trojan [Fig. 368]

1909 Myzobolus oculi-leucisci Trojan 1909: 679-682

Habitat: Vitreous body of the eye of Leuciscus rutilus L.; Prague (May?).

Vegetative form: Two cysts, spherical and subspherical, 100 to 180μ in diameter. Ectoplasm finely granular. Outer portion of endoplasm with small nuclei, then larger nuclei each surrounded by protoplasm, while the central portion contains spores. Polysporous.

Spore: Elongated oval, flattened dorso-ventrally. Posterior margin rounded. At the anterior end, a single polar capsule with distinctly visible coiled polar filament. Shell smooth without any markings. Sporoplasm with one nucleus, usually elongated oval $(2.8\mu$ in diameter) and one vacuole, occupies more than half of the space of the spore. Dimensions: length 9 to 10μ , breadth 4.5 to 5.5μ , thickness 3μ , polar capsule 5μ by 2μ .

MYXOBOLUS TOYAMAI Kudo [Figs. 369 to 370]

 1915
 Myzobolus toyamai
 Kudo
 1915 : 517-523

 1917
 Myzobolus toyamai
 Kudo
 1917 : 163-170

Habitat:—Connective tissue of branchial lamellae of Cyprinus carpio L.; Tokio (July).

Vegetative form: Cysts, ovoidal or in shape of calabash. Small form 67 by 50μ , shows clear differentiation of protoplasm. Ectoplasm radially striated, often, differentiates fine processes (2 to 3μ long). Endoplasm coarsely granular, contains nuclei from 1 to 4μ in diameter. Size up to 190μ in greatest diameter in sections. Two spores are formed in each pansporoblast. Polysporous.

Spore: Pyriform, with attenuated anterior and rounded posterior ends. No bilateral symmetry. Lateral sides are curved. Calabash shaped spores often occur. Shell without any marking, thickened at the anterior end. Sutural ridge shows sometimes a short $(1.5\mu \log)$ tail-like process at the posterior tip. A single pyriform polar capsule at the anterior end; in stained preparations, a small, oblong mass of protoplasm is seen between the polar capsule and the shell. Coiled polar filament distinct. Sporoplasm with two nuclei of usually same size and a relatively large iodinophilous vacuole, 3μ in diameter. Dimensions: length 15μ , breadth 7 to 8μ , thickness 5 to 6μ , polar capsule 7 to 8μ by 3 to 4μ , length of polar filament 40 to 45μ (pressure, perhydrol, KOH).

MYXOBOLUS NOTATUS Mavor [Figs. 371 to 372]

1916 *Myxobolus notatus* Mavor 1916a : 70-71

Habitat: Connective tissue of the voluntary muscles on the sides or tail of *Pimephales notatus* Raf.; Georgian Bay, Canada (Summer).

Vegetative form: Cysts as large as 3mm. in diameter, are surrounded by a layer of columnar epithelial cells (origin and significance?) and a dense layer of connective tissue. Protoplasm is not clearly differentiated, tho the cyst is surrounded by an area devoid of nuclei. In the outer region of endoplasm, numerous nuclei each with a caryosome, are recognized. In the course of spore formation, two nuclei for polar capsules appear at first, one of which degenerates later. Polysporous.

Spore: Pyriform, with a posterior extension forming a process, 5μ in length and as broad as the spore. A single polar capsule at the anterior end. An iodinophilous vacuole in the sporoplasm. Dimensions: length 17 to 18μ , breadth 7.5 to 8μ , polar capsule 7μ by 4μ , length of polar filament 95μ .

MYXOBOLUS sp. Kudo

1918 Myxobolus sp.

Kudo

1918:15

Habitat: Spleen of *Perca flavescens*; West Falmouth, Mass. (August). Isolated spores were noticed in one fish, in smears and section preparations. Vegetative form: Not observed.

Spore: Ovoidal, attenuated at the anterior end. Shell uniformly thick. A single polar capsule opens at the anterior tip. Sporoplasm contains an iodinophilous vacuole and two nuclei of equal size (2μ) . Dimensions: length 18 to 20μ , breadth 8μ , polar capsule 7 to 9μ by 3 to 6μ .

MYXOBOLUS ROHITAE Southwell et Prashad [Figs. 373 to 374]

1918 Myzobolus rohitae Southwell et Prashad 1918: 344-347

Habitat: Branchiae of *Labeo robita*; Turag river, Mirpur, Dacca district, Bengal (June). Type specimens of Indian Museum P48/1. Infection was heavy. In one case 53 cysts were found on one surface of a single gill.

Vegetative form: Cysts in the gill-filaments. The cysts preserved in alcohol are of a creamy-yellow color, oval to cylindrical in form, lying with the long axis parallel to the gill-filaments. Cysts attached to the gill-filaments with the flattened surface. Size: length 3.1 to 3.8mm.; breadth 0.8 to 1.2mm. Cyst-wall striated vertically, covered with an epithelium, two to three layers thick. In the central portion and at the periphery, mature spores and pansporoblasts as well as immature spores were found respectively. Polysporous.

Spore: Elongated pyriform, rounded at the posterior end and acutely pointed anteriorly. Sutural ridge slightly raised. One polar capsule present, being of conspicuous size. Coiled polar filament is distinctly observed in the polar capsule. An iodinophilous vacuole, 3.6μ in diameter, in the sporoplasm. "Lying just posterior to it is the nucleus of the spore. A few granules of chromatin were also seen lying scattered in the protoplasm." Dimensions: length 30 to 32μ , breadth 7 to 8μ , length of the polar capsule 22 to 23μ , that of polar filament 92 to 97μ .

MYXOBOLUS SENI Southwell et Prashad [Figs. 375 to 376]

1918 Myzobolus seni Southwell et Prashad 1918: 347

Habitat: On the median and caudal fins of Labeo robita; Mirpur, Dacca (January). Type specimens in Indian Museum numbered P 53/1. Vegetative form: Trophozoites form cysts which are elongated ellipsoidal. Size from 4.7mm. to 5.4mm. in length, 2.9mm. to 3.7mm. in

breadth. Color of the cyst whitish with black scattered granules on the surface.

Spore: Oval, much wider behind than in front and pointed at the anterior end. Sutural ridge is slightly thickened. A single polar capsule, showing much coiled polar filament. Iodinophilous vacuole 2.3μ in diameter. Dimensions: length 13.2 to 13.6μ , breadth 10.1 to 10.3μ , length of polar capsule 4μ , length of polar filament 43μ (in one case).

MYXOBOLUS MISGURNI nov. spec.

[Figs. 377 to 378]

1916 Myxobolus fukrmanni Kudo 1916 : 5

Habitat: Gall-bladder of *Misgurnus anguillicaudatus*; Tokio (September). About 50% of the fish examined showed a few isolated spores floating in the bile.

Vegetative form: Unobserved.

Spore: Form elongated pyriform, with attenuated anterior and rounded posterior ends. Shell uniformly thick. Over sutural edge, shell exhibits many (up to 12) triangular markings. Sutural ridge distinct. A single pyriform polar capsule at the anterior end. Sporoplasm contains an iodinophilous vacuole and two nuclei. Coiled polar filament distinct in vivo. Dimensions of fresh spores: length 14 to 15.5 μ , breadth 6 to 7.3 μ , thickness 5 to 6 μ , polar capsule 6.3 μ by 2 to 3 μ , length of polar filament up to 100 μ .

Remarks: The writer reported this species as identical with *Myxobolus fuhrmanni* Auerbach. By repeated reexamination and comparison with Auerbach's description, however, he came to the conclusion that the present form should be treated as a new species, on account of the difference of the host and the characters of the spore.

MYXOBOLUS PFEIFFERI Thélohan

[Figs. 379 to 385]

1890	M yxos poridian	Pfeiffer	1890 : 30-37
1891	M yxosporidian	Pfeiffer	1891 : 100, 105-110, 130
1893	M yxos poridian	Pfeiffer	1893 : 118-130
1895	Myxobolus pfeifferi	Thélohan	1895 : 350
1898	Myxobolus pfeifferi	Doflein	1898 : 306, 320, etc.
1906	Myxobolus pfeifferi	Cépède	1906 : 59
1906	Myxobolus pfeifferi	Stazzi	1906 : 14–19
1906	Myxobolus pfeifferi	Mercier	1906 : 427–428 ;
			1906a : 763-764
1908	Myxobolus pfeifferi	Keysselitz	1908 : 253–273,
		-	286-306
1909	Myxobolus pfeifferi	Mercier	1909 : 5-30

Habitat: Muscle and connective tissue of kidney, spleen, intestine, ovary, etc., of Barbus barbus L., and branchiae of B. fluriatilis Ag. and B. plebejus Val.; Drac (June), Neckar, Prag, Milano. The cause of well known "Boil disease" (Beulenkrankheit) or Myxoboliasis tuberosa (Hofer) of the barbels in European waters. Among many observers Keysselitz made a thoro study of the parasite. His observations are as follows: The disease occurs among the fish at any stage of growth. About 8% of the fish, 7 to 15cm. long, caught in May and June between Conz and Trier were infected with the parasites. The heaviest infection, however, occurs among fish up to 40cm. in length; fish 50cm. long or larger show the tumors caused by the parasites, rather rarely. Most of the fish die as the result of the infection between the early part of April and the end of October. The highest mortality is reached in the hottest months, i.e., July and August. The temperature greatly affects the growth of the parasites. Fish kept in the aquarium at a temperature of 25° C. or higher demonstrate the growth of the boil in size daily. The boils are not noticed during the winter and spring, they are formed from the early part of April to the middle of October.

Vegetative form: The parasites develop tumors of conspicuous size.

Keysseltiz's observations are as follows: The tumor varies in size from millet-grains to hen's eggs. Form spherical, oval or elongated. The number of cysts on a single fish, is usually 3 to 4; often one, in some fish, however, 23 were recognized on one fish. Usually tumors separated from each other, rarely many forming one tumor. In one fish, 27cm. in length, a tumor of 7cm. long, 4cm. broad and 3cm. thick, was observed in July. The seat of infection is: the muscle of the body, muscle of pectoral and anal fins, often in peritoneum and rarely in intestine. As the result of breaking up of the cyst membrane, spores are also found in the testis, liver and kidney.

The tumor is composed of many vegetative forms, rounded, oval, elongated, variously branched or flattened. Size reaches to 1.5mm. in diameter. Protoplasm is usually differentiated into ectoplasm and endoplasm. The surface is not often smooth, but shows irregular outline. Ectoplasm is seen often as a very thin, uniformly hyaline, indistinctly granular or radially striated layer, giving the network-like appearance to the surface of the body. Endoplasm, stained more deeply around the peripheral part than other portion, shows a coarsely alveolar structure in the central region. It contains vegetative nuclei, developmental stages of propagative nuclei, granules, fat-like, often leucocytes and red blood corpuscles. The leucocytes, uninuclear or multinuclear, were seen at the periphery, apparently in the course of degeneration. Red blood corpuscles were found, in section, inside of the apparently intact parasite. Each pansporoblast develops into two spores. Polysporous.



Cépède observed one cyst, about 2mm. in diameter, in the connective tissue of the third gill arch.

Spore: Thélohan described as follows: Ovoidal. Sutural edge shows folds. A small triangular intercapsular appendix. Dimensions: length 12μ , breadth 10μ . Cépède's form showed exactly the same dimensions.

Keysselitz gave the characters of the spore as follows:

Flattened oval. Shell smooth. A small intercapsular appendix. Sutural edge having a number of small flat enlargement, size and number being variable. Two convergent narrow canals (foramina) penetrate the shell at the anterior end. Two polar capsules, pyriform and of equal or nearly equal size, are located at the anterior half. Coiled polar filament distinct, coiled 7 to 8 times. No distinct connection between polar capsule and the filament. Sporoplasm fills the posterior half of the spore, extending into intercapsular cavity. It is finely reticular, exhibits one or two rounded or oval vesicular nuclei and an iodinophilous vacuole. Fatlike substance is often seen around the polar capsules. Spores kept in water for four months remain intact in large numbers. Dimensions: length 12 to 12.5μ , breadth 10 to 10.5μ , length of polar capsule, 5.5 to 6μ , length of polar filament 28 to 34μ .

MYXOBOLUS INAEQUALIS Gurley

[Fig. 411]

1841		M üller	1841 : 487-488
1893	Myxobolus inaequalis	Gurley	1893 : 414
1894	Myxobolus inaequalis	Gurley	1894:212

Habitat: In the skin of the head of *Piramutana blochi* Cuv. et Vil. and Synodontis schall Bl. Schn.; Guiana, Surinam.

Vegetative form: Very small pustules in the skin of the head.

Spore: Ovoidal. Two polar capsules of unequal size at the anterior end. Dimensions: length 0.0052", breadth 0.0033".

MYXOBOLUS DISPAR Thélohan [Fig. 386]

1895	Myxobolus dispar	Thélohan	1895 : 348
1904	Myxobolus dispar	Hofer	1904:50
1910	Myxobolus dispar	Wegener	1910 : 73-74
1911	Myxobolus dispar	Nemeczek	1911:145

Habitat: Branchiae of Carassius carassius L., branchiae and epithelium of intestine of Cyprinus carpio L., also muscle and spleen of Scardinius erythrophthalmus L. and in the skin and the connective tissue of Alburnus lucidus Heck.; France, Austria, Königsburg (March, July, September).

Vegetative form: Not described by Thélohan.

Wegener's description is as follows: Cysts: white in color; spindle shape with pointed ends. Cysts in Carassius carassius L. smaller and oval.

Size 3.5mm. by 0.8mm. Cysts are surrounded by thick layers (7 to 8μ) of the connective tissue of the host. Ectoplasm seems to be undifferentiated. Endoplasm granular, contains a larger number of spores. Polysporous.

Spore: Thélohan's diagnosis is as follows:

Ellipsoidal or slightly oval. Shell with 3 to 5 folds along sutural edge. Polar capsules of unequal size, with a small intercapsular body. The vacuole is difficult to stain with iodine. Dimensions: length 10 to 12μ , breadth 8μ , polar capsule 7μ by 5μ .

Wegener's form is as follows: length 11 to 12μ , breadth 7.5 to 8μ , larger polar capsule 6 to 7μ by 3.5μ , smaller one 4μ by 2.5 to 3μ . The sporoplasm is shifted toward the smaller polar capsule.

MYXOBOLUS ELLIPSOIDES Thélohan

[Figs. 387 to 389]

1852		Remak	1852 : 144-146
1892	Myxobolus ellipsoides	Thélohan	1892:177
1895	M yxobolus ellipsoides	Thélohan	1895 : 350-351
1898	Myxobolus ellipsoides	Doflein	1898: 324, etc.
1905	Myxobolus ellipsoides	Nufer	1905 : 77, 79, 186.
1910	Myxobolus ellipsoides	Wegener	1910 : 74-75
1912	Myxobolus ellipsoides	Lo Giudice	1912 : 1-79

Habitat: Connective tissue of air bladder, branchiae, kidney, spleen, liver and cornea of Tinca tinca L., branchiae of Abramis brama L., Alburnus lucidus Heck., Leuciscus rutilus L., Squalius cephalus L., Abramis vimpa Cuv., Blicca björkna L., Idus melanotus; France, Vierwaldstätter See, Prague, Masurische See, Italy.

Vegetative form: Thélohan does not describe.

According to Wegener, white cysts, elongated oval; 2mm. by 0.5mm. in size. Polysporous.

Spore: Thélohan described as follows: Flattened elliptical, rather elongated. Sutural edge broad without any folds. Shell with no marking. Form of the spore somewhat variable. Two polar capsules of equal size, capsulogeneous nuclei present even when fully grown. Abnormal spores are of frequent occurrence. Dimensions: length 12 to 14μ , breadth 9 to 11μ , length of polar capsule 4μ .

Wegener's form: length 14 to 15μ , breadth 10 to 11μ , polar capsule 4 to 5μ by 3μ . Shell comparatively thick. One spore had a tail 5μ long.

MYXOBOLUS EXIGUUS Thélohan

[Figs. 390 to 395]

	· · · · · · · · · · · · · · · ·			
1891	Myxosporidium mugilis?	Perugia	1891 : 23	
1895	Myzobolus exiguus	Thélohan	1895 : 349-350	
1906	Myxobolus exiguus	Schröder	1906:195	
1910	Myxobolus exiguus	Wegener	1910 : 75	
1912 [°]	Myxobolus exiguus	Parisi	1912 : 294-295	

Habitat: Branchiae of Abramis brama L. and Chondrostoma nasus L., wall of stomach, pyloric coecum and intestine, branchiae, spleen, kidney of Mugil chelo Cuv., M. capito Cuv. and M. auratus Riss.; Le Vivier-sur-mer, Banyuls, Marseille, Heidelberg, Pregel, Frisches Haff, Kurisches Haff, Geneva, Napoli.

Vegetative form: No description by Thélohan.

Wegener writes as follows:

Cysts of variable size. Color white. Usually small and narrow, 0.5 to 0.7mm. long and 0.2mm. wide. Frequently large round cysts of 1.2 to 1.5mm. in diameter, filling the lamella. Cysts are surrounded by 10 to 11μ thick membrane composed of the connective tissue of the host. Ectoplasm, 5μ thick, is faintly stained by hematoxylin. Outer region of endoplasm, alveolar and densely loaded with nuclei, while in the central portion with mature spores in granular ground-mass.

Parisi's observations are as follows:

Cysts in the intestinal wall of Mugil auratus, large; reaching a length of 3mm.

Spore: Thélohan's description is as follows:

Flattened ovoidal, with more or less attenuated anterior end. Sutural edge shows fairly noticeable folds. A small triangular intercapsular appendix. Vacuole in the sporoplasm is usually hard to stain with iodine. Dimensions: length 8 to 9μ , breadth 6 to 7μ , length of polar filament 15μ (KOH).

Wegener observed as follows: Rounded with slightly pointed anterior end. Length 8 to 9.5μ , breadth 6 to 7.5μ , polar capsule 4.5μ by 2 to 3μ . Shell exhibits small folds around the sporoplasm. An intercapsular triangular body indistinctly visible.

Parisi gave the following dimensions: length 8 to 8.5μ , breadth 6 to 7μ , thickness 5.5μ , polar capsule 3 to 4μ by 1.5 to 2μ , length of polar filament 30μ (alkaline). Folds usually 6 in number. Coiled polar filament visible in vivo.

MYXOBOLUS OVIFORMIS Thélohan [Fig. 396]

1854 Lieberkühn 1854:21-22 1892 Myxobolus oviformis Thélohan 1892:177 1895 Myxobolus oviformis Thélohan 1895:351 1905:77, 186 1905 Myxobolus oviformis Nufer 1906 Myxobolus oviformis Cépède 1906:60 1910 Myxobolus oviformis Wegener 1910:76-78

Habitat: Fin (subcutaneous tissue), spleen, kidney and liver of Gobio gobio L.; branchiae of Alburnus lucidus Heck., Cyprinus carpio L., Blicca björkna L., Abramis brama L. and A. vimba L.; France (Isère), Frisches Haff (especially spring months), Switzerland.

Vegetative form: Thélohan gave no description.

Wegener's observations are as follows:

Cysts, white, 0.75 to 1.7mm. by 0.4 to 0.7mm. In sections, cysts are shown to be surrounded by a thick (10 to 20μ , average 16μ) layer of connective tissue. Ectoplasm a thin (6 to 8μ thick) layer, exhibits a transverse striation. The striation is often absent at places in ripe cysts. Endoplasm finely granular. In young cysts, it is, however, reticulated, with nuclei of 1.5μ in diameter.

Spore: Thélohan described as follows: Flattened ovoidal with pointed anterior end. Shell smooth. No folds. Polar capsule comparatively large. Dimensions: length 10 to 12μ , breadth 9μ , polar capsule 6μ .

Cépède observed numerous spores in the liver and kidney of Gobio gobio. Dimensions in vivo: length 10 to 12μ , breadth 9μ , length of polar capsule 6μ . Polar capsules of equal size. Coiled polar filament distinct.

Wegener's form: length 10.5 to 11μ , breadth 7.5 to 8μ , polar capsule 5 to 6μ by 3μ .

Remarks: Wegener recognized another form, which seems to be of very rare occurrence and which can not be distinguished distinctly from the above described form. Cysts at the end of the branchial lamellae. Size 1.7 to 2mm. in largest length. Spore resembles more closely the figure given by Thélohan for Myxobolus oviformis than the above mentioned form which he observed. A small intercapsular appendix (rounded) indistinct. Sporoplasm comparatively small. Length 12.5 to 13.5 μ , breadth 9μ , polar capsule 7.5μ by 3μ .

MYXOBOLUS LINTONI Gurley

[Figs. 404 to 408]

1891		Linton	1891 : 99-102
1893	Myxobolus lintoni	Gurley	1893 : 414
1894	Myxobolus lintoni	Gurley	1894 : 238

Habitat: Superficial musculature and subcutaneous tissue of Cyprino-don variegatus; Woods Hole (August).

Vegetative form: Cysts, not closed, but fungoid masses of an irregular shape, varying in size from 4mm. by 2.5mm. to 10mm. by 4mm., projecting as much as 3mm. above general surface of skin. The skin of the host overlying these tumors, is more or less cracked and broken, the scales being scattered.

Spore: Elliptical in the front view; lenticular in side view. Shell thick. Sutural ridge marked. Two polar capsules, convergent, at the anterior end. Spores kept in alcohol, extruded polar filaments under the action of iodine water and sulphuric acid. Sporoplasm with a large iodinophilous vacuole. Dimensions: length 13.9μ , breadth 11μ , thickness 8μ .

MYXOBOLUS GLOBOSUS Gurley [Figs. 409 and 410]

1893	Myxobolus globosus	Gurley	1893:415
1894	Myxobolus globosus	Gurley	1894 : 241

Habitat: Branchial lamellae of Erimyzon sucetta oblongus Lac. (Catostomus tuberculatus Le Sueur); Kinston (N.C.), Columbia, (S.C., March), tributaries of Fox River.

Vegetative form: Cysts, whitish, elongated elliptical or rod-shaped, surrounded by very thin membrane? Size up to 0.5mm. in max. length. Polysporous.

Spore: Globose, subcircular in outline. Shell thin and very transparent. Sutural ridge very wide, being one third of the thickness of the spore. Polar capsules two, of equal size, divergent. Vacuole present, but not clearly contoured. Dimensions: length 7 to 8μ , breadth 6 to 7μ , thickness 5μ .

MYXOBOLUS OBLONGUS Gurley

[Figs. 412 to 416]

1841		Müller	1841 : 487-490
1893	Myxobolus oblongus	Gurley	1893:414
1894	Myxobolus oblongus	Gurley	1894 : 234-238

Habitat: Beneath the skin, chiefly of the head of Erimyson sucetta oblongus Lac. (Catostomus tuberculatus Le Sueur); Kinston, tributaries of Fox River.

Vegetative form: Cysts, round or elliptic, not over 1mm. in diameter, covered by resistant membrane. Color whitish. Polysporous.

Spore: Spatular, approaching roundish-oblong. Shell thin and transparent. Sutural ridge wide. Two polar capsules, pyriform, of equal size. Sporoplasm extending forward along the upper surface. Vacuole could not be detected. Dimensions: length 14 to 17μ , breadth 8.5μ , thickness 5 to 6μ .

MYXOBOLUS TRANSOVALIS Gurley [Figs. 417 and 418]

1893	Myxobolus transovalis	Gurley	1893:415
1894	M vrobolus transonalis	Gurley	1894 : 242

Habitat: Under scales on external surface of *Phoxinus* (Clinostomus) funduloides Girard; 4 Mile Run, Carlius, Va., tributary of Potomac River (June). No fish of the same species caught from the same locality on August 29 of the same year was found infected.

Vegetative form: It is not certain whether cysts exist or not. Spores in mass, appear to be held together by a small gelatinous or mucoid mass

which has no attachment to the subjacent connective tissue. It forms a thin discoidal mass situated in the center of the concave surface of the scale. The color of the mass slightly more yellowish than the surrounding tissue, when coagulated. It is exceedingly difficult to detect its presence in the fresh state.

Spore: Elliptical, with the largest diameter passing thru two polar capsules. Shell thin. Sutural edge narrow. Two polar capsules of equal size convergent. Polar filament is extruded under the action of glycerine and sulphuric acid. The vacuole in the sporoplasm is difficult to detect. Sporoplasm also contains two nuclei, rarely one, 1 to 1.5μ in diameter. Dimensions: length 6 to 7μ , breadth 8μ .

MYXOBOLUS OBESUS Gurley [Figs. 419 and 420]

1883		Balbiani	1883 : 203
1893	Myxobolus obesus	Gurley	1893:415
1894	Myxobolus? obesus	Gurley	1894 : 239
1899	Myxobolus obesus	Labbé	1899:100
1906	Myxobolus obesus	Cépède	1906 : 60-61

Habitat: On Alburnus alburnus L.; branchiae and kidney of A. lucidus Heck. (A. mirandella Bl.); Lac du Bourget.

Vegetative form: Balbiani gave no observation.

Cépéde observed as follows: Cysts, ovoidal, more or less elongated or variable in form, not exceeding 800μ in length. In kidney, numerous cysts were of subspherical, ovoidal or rarely irregularly elongated form. Subspherical cysts 500 to 600μ in average diameter. Polysporous.

Spore: Cépède describes as follows: Subcircular or ovoidal in front view; lenticular in side view. Sutural edge exhibits variable numbers (4 to 5) of fold-like markings on the shell. Polar capsules pyriform and of equal size. Coiled polar filament distinct. A small triangular intercapsular appendix. Sporoplasm with a subspherical and clearly outlined vacuole and two nuclei. Dimensions in vivo: length 11.5 to 12μ , breadth 7.5 to 8μ , thickness 5μ . Those of fixed and stained spores: length 11.25 to 11.50μ , breadth 7.25 to 7.50μ , length of polar capsule 5μ .

Remarks: Cépède mentions that Alburnus alburnus L. mentioned by Gurley is "without doubt" identical with A. lucidus Heckel.

MYXOBOLUS CYCLOIDES Gurley

[Fig. 421]
1841 Müller

1893 Myxobolus cycloides Gurley 1893:415 1894 Myxobolus cycloides Gurley 1894:239 1906 Myxobolus cycloides Cépède 1906:61-63 1910 Myxobolux cycloides Wegener 1910:79-80

1841:481,486

Habitat: Opercle, pseudobranchiae and kidney of Leuciscus rutilus; branchiae of Scardinius erythrophthalmus, Blicca björkna L., Gobio gobio L., Abramis vimba L., A. brama L., Rhodeus amarus Bl., Alburnus alburnus L., Lota lota L.; France (Isère), Germany (Pregel, Frisches and Kurisches Haff, Masurische See, January, May).

Vegetative form: Wegener observed cysts as follows. A type: 1 to 2mm. by 0.4 to 0.7mm. Form exactly like that of *Myxobolus oviformis*. B type: small and round, present in groups. C type: small 0.5mm. by 0.2mm.

Spore: Gurley gave the following short diagnosis from the observations of J. Müller: subcircular-ovate or broadly rounded elliptic, length 12μ .

Cépède distinguishes three different types of spores as follows: Lenticular in side view; subcircular $(13.5\mu$ by 13μ), oval $(14.7\mu$ by 11.4μ) and ovoidal $(16\mu$ by 11μ) in front view. Two polar capsules of equal size $(6\mu$ by 4μ), closely set or separated $(3\mu$ apart) from each other. Coiled polar filament distinct. A small triangular intercapsular appendix. Sporoplasm refractive and finely granular. Sutural edge exhibits folds of variable number at the posterior portion. Dimensions of fixed and mounted spores: length 10.5 to 12μ , breadth 7.5 to 8μ .

Wegener, without noticing Cépède's paper, also mentions three different types chiefly distinguished by the spore as follows:

A type (common form), in the branchiae of Lota lota, Abramis brama, A. vimba, Blicca björkna, Leuciscus rutilus, Alburnus alburnus and scardinius erythrophthalmus. Cysts mentioned above.

Spore. Rounded or oval; flattened. A tail, 15μ long, was noticed twice. A triangular intercapsular appendix. Sutural edge usually having folds. Polar capsules often differ in form and size in different cysts, tho they are constant in one and the same cyst, causing the variability in size of sporoplasm. Dimensions: length 11 to 12.5μ , breadth 8 to 9μ , polar capsule 4.5 to 6μ by 3 to 3.7μ , in many cysts 7.5μ by 4μ .

B type. In the fifth gillarch of Gobio gobio L. Cysts mentioned above. Spore. Elongated oval. A triangular intercapsular appendix. Indistinct folds on sutural edge. Dimensions: length 12.5 to 13.5 μ , breadth 8 to 10μ , polar capsule 5 to 6μ by 3 to 4μ .

C type. In the branchiae of Rhodeus amarus Bl. and Alburnus alburnus L. (April and May). Cysts mentioned above.

Spore. Rounded. Distinct intercapsular appendix. Folds distinct on sutural edge. Dimensions: length 12 to 15μ , breadth 9 to 10μ , polar capsule 5 to 7μ by 3 to 4μ .

MYXOBOLUS SPHAERALIS Gurley

1874		Clapar è de	1874 : 113-114
1893	Myxobolus sphaeralis	Gurley	1893:415
1894	Myxobolus sphaeralis	Gurley	1894 : 240

Habitat: Mucosa of branchiae of Coregonus lavaretus L. (C. fera); Lake Geneva.

Vegetative form: Cysts, 0.25 to 0.33mm. in diameter. Polysporous.

Spore: Spherical, 9μ in diameter.

MYXOBOLUS ANURUS Cohn [Figs. 422 and 423]

1895	Myxobolus anurus	Cohn	1895 : 42-43
1896	Myxobolus anurus	Cohn	1896:266
1899	Henneguya psorospermica		
	anura	Labbé	1899:102
1910	Myxobolus amurus	Wegener	1910 : 76
1911	Henneguya psorospermica	-	
	anura	Nemeczek	1911 : 146

Habitat: Branchiae of Esox lucius L.; Königsberg (March, December), Frisches Haff, Pregel, Masurische See, Lotzen, (September, October).

Vegetative form: Cysts small rounded and of white color. Cohn measures length 0.6mm., breadth 0.34mm. Wegener's form: length 0.3 to 0.5mm, and breadth 0.2 to 0.3mm.

Spore: Cohn's descriptions are as follows: More or less oval. Dimensions: length 12 to 15μ , breadth 4 to 6.8μ , polar capsule 5.5 to 7μ by 2.1 to 2.5μ , length of polar filament 32 to 38μ .

Wegener's form: Elongated and narrow, often with a tail. Dimensions: length 15μ (maximum up to 18μ), breadth 6 to 7μ , polar capsule 8μ by 3μ .

Remarks: The Labbé classified this as a subspecies of *Henneguya pso-*rospermica Thélohan, Wegener's observation gives stronger basis for placing this form in the genus Myxobolus.

MYXOBOLUS sp. Gurley [Fig. 424]

1882		Bütschli	1882 : 590
1894	Myzobolus sp. incert.	Gurley	1894 : 214
1899	Myzobolus sp.	Labbé	1899:100

Habitat: Nais lacustris L. (N. proboscidea); Locality? Vegetative form: Cysts, 8mm. by 4.25mm. Polysporous.

Spore: Oval or circular; tailed or untailed. These spores of different form occur, often, without order in the same cyst.

MYXOBOLUS sp. Gurley [Fig. 425]

1894 Myzobolus sp. incert. Gurley 1894: 239

Habitat: Body cavity of Carassius carassius L.; Leipsic.

Vegetative form: Not observed.

Spore: Broadly elliptic; shell bivalve; valves equally convex. Sutural ridge. Two equal polar capsules. Sporoplasm with a vacuole. Dimensions: length 14μ , breadth 10μ , thickness 5μ .

Remarks: This species seems to be very similar to M. carassii Klokačewa (page 150).

MYXOBOLUS sp. Gurley [Figs. 426 to 429]

1841		•	M üller	1841 : 4 80
1894	Myxobolus sp.		Gurley	1894 : 240-241
1899	Myxobolus sp.		Labbé	1899:100

Habitat: Skin of opercle, in the branchiae, on the head or on the fin of Lucioperca lucioperca L.; Germany, Don.

Vegetative form: Cysts 1.09 to 2.18mm. in diameter. Color whitish. Polysporous.

Spore: Rounded. Thickness equal to half the breadth. Sutural ridge. Two polar capsules, of equal size, converging.

MYXOBOLUS CYPRINI Doflein [Figs. 430 to 432]

1896		Hofer	1896 : 2, 38-39
1898	Myxobolus cyprini	Doflein	1898 : 288, 320, 325
1904	Myxobolus cyprini	Hofer '	1904 : 66-67
1909	Myxobolus cyprini	Doflein	1909:780-783
1916	Myzobolus cyprini	Doflein	1916: 1026-1027

Habitat: Suppurative connective tissue and epithelium of kidney, liver and spleen of *Cyprinus carpio* L., rarely *Tinca vulgaris* Cuv. and *Abrams brama* L.; Germany, Austria. According to Hofer the parasites cause so-called "small pox of carp" among carp in German waters.

Vegetative form: Small ameboid. Form irregular. The youngest form with a single or many nuclei, is found in the epithelium of the kidney. Multiplication by multiple division, the nuclei undergoing amitotic division. Endoplasm contains homogeneous, yellow and refractive bodies. Also found in the state of diffuse infiltration. Spores are found in the parenchym of the kidney.

Spore: Oval. Shell thickened $(1.5\mu$ wide) along the sutural edge. Two converging polar capsules cross each other, in front view, at the anterior tip. Sporoplasm with an iodinophilous vacuole. Dimensions: length 21μ , breadth 15μ , length of polar capsule 6μ . Doflein (1916:1027) gives the following dimensions: length 10 to 16μ , breadth 8 to 9μ .

Hofer gives the following dimensions: length 10 to 12μ (up to 16μ), breadth 8 to 11μ , polar capsule 5 to 6μ by 3μ , sutural edge 1.5μ .

MYXOBOLUS NEUROBIUS Schuberg et Schröder [Figs. 433 to 436]

1905 Myzobolus neurobius Schuberg and Schröder 1905: 49-56

Habitat: Nervous tissue of Trutta fario L.; Gutach (May?).

Vegetative form: Cysts, usually elongated, often spherical. Elongated form 0.9mm. by 0.02mm. The seat of the cysts is between the medullary sheath and sheath of Schwann. Neither medullary sheath nor axis-cylinder was infected. Cyst-membrane could not be made out. Cysts contained only full-grown spores without any younger stage. Polysporous.

Spore: Broad oval in front view; spindle shaped in side view. Anterior end attenuated, posterior end rounded. Shell somewhat thick. Sutural ridge is not particularly marked. Edge without any fold. No intercapsular appendix. Sporoplasm, with a large and spherical iodinophilous vacuole and a single nucleus, occupies less than one half of the inner space of the spore. Two polar capsules, pyriform, fuse into one at the anterior end. Coiled (8 to 10 times) polar filament distinct. Dimensions: length 10 to 12μ , breadth 8μ , thickness 6μ , polar capsule 6 to 7μ by 2μ .

MYXOBOLUS AEGLEFINI Auerbach

[Figs.	437	to	4411
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1906	Myxobolus aeglefini	Auerbach	1906 : 568-570
1906	Myxobolus aeglefini	Auerbach	1906a:115-119
1907	Myxobolus esmarkii	Johnstone and Woodcock	1907 : 204–208
1909	Myxobolus aeglefini	Auerbach	1909:76-78
1910	Myxobolus aeglefini	Auerbach	1910c : 181-182
1911	Mvxobolus aeglefini	Nemeczek	1911:162

Habitat: Cartilage and bone of cranium and eye of Gadus aeglefinis G. callarias, G. merlangus L., G. morrhua L., G. esmarkii and Molva vulgaris Flem.; Norway, Morecambe (March).

Vegetative form: Cysts in cartilage and bone of cranium and in cartilagineous layer of the sclerotic of the eye. Protoplasm is distinctly differentiated. Ectoplasm somewhat vacuolated; endoplasm granular with numerous small nuclei. Polysporous.

Johnstone's observations are as follows: Round the peripheral part of the cornea, and covered loosely by conjunctiva are a number of milk-white rounded or oval bodies, from about 1 to 3mm. in diameter. Several of these fused to form elongated mass which lie along the curvature of the periphery of the eye. These cysts also invade the lateral and posterior parts of the bulbus oculi. In sections, the cysts lie within the thickness of cartilaginous layer of the sclerotic. This latter is enlarged into thick layer (2mm.) by the presence of the cysts.

Nemeczek mentions irregular cysts of 1.5mm. in diameter.

Spore: Elliptical in front view. Two polar capsules convergent. No intercapsular appendix. Sutural edge rather thick with a number of folds on the posterior margin. Sporoplasm with two nuclei and an iodinophilous vacuole. Dimensions: length 10.8 to 11.7μ , breadth 9.9 to 10.4μ , thickness 7.2 to 9μ , length of polar capsule 4.5 to 5μ .

Woodcock's form has a spore with the following characters:

Slightly ovoid. Sporoplasm always contains a large and well defined vacuole and two nuclei. Dimensions: length 10μ , breadth 8μ , length of polar capsule 3.25 to 3.5μ .

MYXOBOLUS GIGAS Auerbach [Figs. 442 to 445]

1906	Myxobolus gigas	Auerbach	1906 : 386-391
1910	Myxobolus gigas	Auerbach	1910c : 182
1912	Myxobolus gigas	Parisi	1912 : 293-294

Habitat: Subcutaneous connective tissue of the operculum of *Abramis brama* L.; Karlsruhe, Pavia. Parisi observed cysts on the side, on the caudal fin (5 cysts on rays), on other fins, branchiae and in the internal organs of the fish.

Vegetative form: Cysts, spherical or ovoidal. No cyst membrane composed of the connective tissue of the host. Protoplasm is indistinctly differentiated. Ectoplasm thin and radially striated, which gradually turns into endoplasm. Endoplasm finely granular, contains numerous nuclei (2.5 to 2.7μ in diameter). Size of greatest form 360μ by 290 to 300μ . According to Parisi size up to 1.5 mm.

Spore: Elliptical when viewed from the front. Sutural edge somewhat narrow, having a number of folds at the posterior portion. Sporoplasm with an iodinophilous vacuole and two nuclei. Dimensions: length 16.9 to 21.6μ , breadth 13 to 16.2μ thickness 9μ , length of polar capsules 7.8μ , length of polar filament 90μ (sulphuric acid).

Parisi gives 150µ for the length of polar filament.

MYXOBOLUS VOLGENSIS Reuss [Figs. 446 to 448]

1906 Myxobolus volgensis Reuss 1906: 200-201

Habitat: Branchiae, cornea and dorsal fin of Lucioperca volgensis Pall; Volga.

Vegetative form: Cysts, spherical, 0.3 to 1mm. in diameter. Polysporous.

Spore: Broad elliptic or rounded. Sutural edge has at least 3 folds. Sporoplasm with an iodinophilous vacuole. Dimensions: length 8.25 to 9.5μ , breadth 7.25 to 8.25μ , thickness 4.5 to 5.5μ , polar capsule 3μ by 2μ .

MYXOBOLUS SCARDINII Reuss [Fig. 449]

1906 Myzobolus scardinii Reuss 1906: 201

Habitat: Branchiae of Scardinius erythrophthalmus L.; Volga.

Vegetative form: Cysts, elongated oval. Smaller cysts rounded oval, 0.8mm. by 0.5mm., the larger forms elongated, 1.2mm. by 0.5mm. Polysporous.

Spore: Broad elliptical. Sutural edge narrow, having folds. A larger triangular intercapsular process. An iodinophilous vacuole in sporoplasm. Dimensions: length 11 to 12μ , breadth 9 to 9.5μ , thickness 4.5 to 5μ , polar capsules 5μ by 2.5μ .

MYXOBOLUS PHYSOPHILUS Reuss [Figs. 450 and 451]

906 Myzobolus physophilus Reuss 1906 : 201-202

Habitat: Surface of air bladder of Scardinius erythrophthalmus L.; Volga.

Vegetative form: Cysts, rounded, 1.5mm. in diameter. Polysporous. Spore: Oval, with attenuated anterior end. Sutural edge narrow and smooth. Polar capsules rather large. An iodinophilous vacuole in sporoplasm. Dimensions: length 12 to 13μ , breadth 8.25 to 9μ , thickness 6.5 to 7μ , polar capsules 6μ by 2.5μ .

MYXOBOLUS MACROCAPSULARIS Reuss [Fig. 452]

1906 Myxobolus macrocapsularis Reuss 1906: 202

Habitat: Branchiae of Blicca björkna L.; Volga.

Vegetative form: Cysts, Elongated oval. Size: 1mm. by 0.5mm. Polysporous.

Spore: Oval with greatly attenuated anterior portion. Sutural edge broad and without any fold. Polar capsules rather large. An iodinophilous vacuole in sporoplasm. Dimensions: length 11 to 13μ , breadth 8.25 to 9.25μ , thickness 5.5μ , polar capsules 6μ by 2.5 to 3μ .

MYXOBOLUS SANDRAE Reuss [Fig. 453]

1906 Myxobolus sandrae Reuss 1906: 202-203

Habitat: Muscle of Lucioperca sandra Cuv.; Volga.

Vegetative form: Cysts. Rounded, 0.5mm. in diameter. Polysporous.

Spore: Oval. Sutural edge broad with many distinct folds. An iodinophilous vacuole in sporoplasm. Dimensions: length 9.25 to 10μ , breadth 7.25 to 8.25μ , thickness 4 to 5μ , polar capsules 3.5μ by 2μ .

MYXOBOLUS BRAMAE Reuss

[Fig. 454]

1906 Myxobolus bramae

Reuss

1906:203-204

Habitat: Branchiae of Abramis brama L.; Volga.

Vegetative form: Cysts. Oval, 0.5mm. long, 0.25mm. broad. Polysporous.

Spore: Oval to nearly spherical. Sutural edge narrow and with indistinct folds. Two polar capsules, with a small triangular intercapsular process. An iodinophilous vacuole. Dimensions: length 11 to 12μ , breadth 9.25 to 10μ , thickness 4.5 to 5.5μ , polar capsules 4 to 5μ by 2.25μ .

MYXOBOLUS CYPRINICOLA Reuss

[Fig. 456]

1906 Myxobolus cyprinicola

Reuss

1906:204

Habitat: Branchiae of Cyprinus carpio L.; Volga.

Vegetative form: Cysts, oval, 0.5mm. by 0.3mm. Polysporous.

Spore: Elongated oval. Sutural edge narrow with many indistinct folds. An iodinophilous vacuole. Dimensions: length 9.25 to 10μ , breadth 7 to 7.25μ , thickness 5 to 5.5μ , polar capsules 4.5μ by 2.5 to 3μ .

MYXOBOLUS BALLERI Reuss

[Fig. 455]

1906 Myzobolus balleri

Reuss

1906: 204-205

Habitat: Branchiae of Abramis ballerus L.; Volga.

Vegetative form: Cysts. Elongated, 1.5mm. by 0.5mm. Polysporous. Spore: Oval, slightly pointed at the anterior end. A triangular intercapsular appendix. Sutural edge smooth. An iodinophilous vacuole. Dimensions: length 11 to 12μ , breadth 9.25 to 10μ , thickness 5.5 to 6.5μ , polar capsules 5.5μ by 2.75μ .

MYXOBOLUS SQUAMAE Keysselitz [Figs. 457 to 459]

908 Myxobolus squamae

Keysselitz

1908 : 273-274

Habitat: Inner surface of the scales of Barbus fluviatilis Agass.; Mosel and Neckar.

Vegetative form: Form variable; rounded, oval, elongated or rarely branched. The outline of the body is not smooth but irregular with numerous small tooth-like projections with which the body comes in contact with the surrounding substance. The parasites seem to be able to dissolve the substance composing the scale. Length 50 to 800μ . In one scale, one or many, up to 8, individuals were found. All showed only advanced stages of spore formation. The parasites are surrounded by a variously developed envelope of connective tissue. Polysporous.

Spore: Elongated oval. Two polar capsules, with 7 to 8 times coiled polar filament. A triangular intercapsular projection. Sporoplasm with an iodinophilous vacuole. Dimensions: length 10 to 10.5μ , breadth 8 to 8.5μ , length of polar capsule 4.5μ .

MYXOBOLUS CORDIS Keysselitz [Figs. 460 and 461]

1908 Myzobolus cordis Keysselitz 1908: 279-282

Habitat: Muscle of ventricle, rarely that of bulbus arteriosus of *Barbus fluviatilis* Ag., spores found in kidney, liver and spleen in the condition of somewhat scattered infiltration; Germany (Mosel and Neckar).

Vegetative form: Elongated, oval, sausage or club form. The body whitish, later yellowish. Size from 0.25 up to 4mm., usually 1 to 1.5mm. in length. Propagative stage and cysts observed. One end of the body is held more or less deeply in the muscle and is covered by cellular envelope as in *Myxobolus musculi*, while remaining larger portion of the body is suspended freely inside of the ventricle, covered with a thin layer probably of endocardiac cells. Fish 30 to 45cm. long harboured 40 to 60 parasites. No movements. Ultimately the cysts are formed with differentiated protoplasm. Polysporous.

Spore: Oval. Shell very thin at the anterior end. At the posterior end, cell-like appendage, 2 to 3μ wide which is probably formed by both valves, is present. Two pyriform polar capsules at the anterior end, which show the polar filament coiled 7 to 8 times. Sporoplasm with a comparatively large and oval iodinophilous vacuole and two nuclei, rarely one (syncaryon). Dimensions: length 12μ , breadth 10μ , length of polar capsule 4.5μ .

MYXOBOLUS MUSCULI Keysselitz [Figs. 462 to 464]

1908 Myzobolus musculi Kevsselitz 1908: 282-286

Habitat: Muscle of the main body, rarely that of fins and operculum, and kidney of *Barbus fluviatilis* Agass. of various size (youngest fish found infected, 2 months old), spores in liver, spleen, kidney and ovary (not the ovum) in diffuse infiltration; Mosel and Neckar.

Vegetative form: Elongated. Body whitish opaque, with differentiated protoplasm. Smallest individual observed, 24μ . Large form 2mm. in length. Many trophozoites are found closely situated, forming a large mass of parasites that reached dimensions of 4mm. by 2mm. The surrounding envelope, varying in thickness, composed of cells with elongated nuclei as those of perimysium. Young cysts surrounded by thin layer of ectoplasm. Polysporous.

Spore: Oval. Two polar capsules usually unequal. Shell as in M. cordis with a small peg closer to the anterior end, polar filament coiled 4 to 5 times, visible in the capsule. Sporoplasm with rarely one (syncaryon), but usually two nuclei and an iodinophilous vacuole. A posterior process as is seen in the spores of M. cordis, but much smaller, was occasionally observed. Dimensions: length 11μ , breadth 8μ , polar capsules 6μ and 4μ long.

MYXOBOLUS sp. Miyairi

1909 Myxobolus sp.

Miyairi

1909:126

Habitat: Branchiae of loach (Misgurnus anguillicaudatus Cant.?); Fukuoka? (Nippon).

Vegetative form: Cysts were not observed.

Spore: No description.

MYXOBOLUS sp. Wegener

[Fig. 465]

1910 Myxobolus sp.

Wegener

1910:78

Habitat: Branchiae (gill-arch) of *Perca fluviatilis* L.; Germany (Frisches Haff, March). Only one case.

Vegetative form: Cysts on a gill-arch, white and round, with a diameter of 1.1mm. Polysporous.

Spore: Form and size very variable. Rounded or elliptical, pointed at the anterior end. Sutural edge showing folds at the posterior portion. Dimensions: length 8 to 10μ (in round form) and 11μ (in elliptical form), breadth 8 to 9μ , polar capsules 4 to 5μ by 2 to 3μ , length of polar filament 40μ .

MYXOBOLUS PERMAGNUS Wegener

[Fig. 466]

1910 Myxobolus permagnus

Wegener

1910:78-79

Habitat: Branchiae and operculum of *Perca fluviatilis* L., air bladder of *Scardinius erythrophthalmus* L.; Königsberg (May), Pregel (March).

Vegetative form: Cysts rounded in form and white in color, resemble to those of M. gigas. No clear ectoplasm layer, nor typical protoplasmic structure. Polysporous.

Spore: Oval, sharply pointed at the anterior end. Sutural edge with 5 to 6 distinct folds at the posterior portion. Polar filament visible in the polar capsules. Dimensions: length 17 to 18μ , breadth 10 to 13μ , polar capsules 7 to 8μ by 3.5 to 4μ .

MYXOBOLUS ROTUNDUS Nemeczek

[Fig. 467]

1911 Myxobolus rotundus

Nemeczek

1911:156-157

Habitat: Branchiae of Abramis brama L.; Austria.

Vegetative form: Cysts, ovoidal or spindle form, 1 to 3mm. long and 1 to 1.5mm. wide. Body white. An extraordinary large number of spores were found in the cysts. Polysporous.

Spore: Round or slightly oval, when viewed from the front. Greatly flattened in side view. Polar capsules convergent, with no intercapsular body. Shell smooth. Sutural edge narrow, without folds. Dimensions: length 10μ , breadth 9.8μ , thickness 3μ , polar capsules 3.8 to 5μ long, length of polar filament 40μ .

MYXOBOLUS MINUTUS Nemeczek [Fig. 468]

1911 Myxobolus minutus Nemeczek 1911: 160

Habitat: Branchiae of Leuciscus sp.; Austria.

Vegetative form: Cysts spherical, oval or elongated with white color. Size: 0.5 to 3mm. by 0.5 to 1mm. Polysporous.

Spore: Rounded oval, similar to that of Myxobolus rotundus. Shell smooth. Sutural edge narrow without folds. Sporoplasm with an iodinophilous vacuole. No intercapsular appendix. Dimensions: length 6μ , breadth 4.2 to 5μ , polar capsule 3μ by 2μ , length of polar filament 50 to 60, often 70μ .

MYXOBOLUS sp. Lebzelter

1912 Myxobolus sp. Lebzelter 1912: 296-297

Habitat: Gall-bladder of Thymallus thymallus L.

Vegetative form: Not observed.

Spore: Sutural ridge distinct. Dimensions, length 5μ , breadth 3μ .

MYXOBOLUS MAGNUS Awerinzew [Figs. 469 and 470]

1913 Myxobolus magnus Awerinzew 1913: 75-76

Habitat: Eye of Acerina cernua L.; Petrograd.

Vegetative form: Trophozoites form white spots in the tissue of iris, with many spores (300 to 400). Each pansporoblast forms in most cases two, sometimes 3 or 5 spores! Polysporous.

Spores: Large, elongated roundish, slightly flattened. Sutural edge somewhat thick, forming a wide ridge, with 4 to 5 folds at the posterior portion. Polar capsules do not cross each other. Sporoplasm with an iodinophilous vacuole and two nuclei. Dimensions: length 38 to 45μ , breadth 32 to 38μ , thickness 28 to 35μ , length of polar capsules 15 to 17μ , diameter of the vacuole 12 to 16μ .

MYXOBOLUS CARASSII Klokačewa [Figs. 471 to 473]

1914 Myzobolus carassii Klokacewa 1914: 182-184

Habitat: Body cavity, liver and intestine of Carassius vulgaris L.; Petrograd?

Vegetative form: Cysts spherical. Those in liver and intestine yellowish, surrounded by an envelope composed of fibrous connective tissue. Secondary cysts are formed. Polysporous.

Spore: Oval, in front view. Two ovoidal polar capsules convergent at the slightly attenuated anterior end. Coiled polar filament visible. Sporoplasm with an iodinophilous vacuole and two nuclei. Sutural edge shows folds in some cases. Dimensions: length 13 to 17μ , breadth 8 to 10μ , thickness 5 to 7μ , polar capsules 6 to 7μ long.

Remarks: Compare with Myxobolus sp. Gurley on page 142.

MYXOBOLUS sp. Southwell

1915 Myxobolus SD.

Southwell

1915:312-313

Habitat: Subcutaneous intermuscular tissue of Rasbora (Cyprinus) daniconius Day; from a stream near Katwan, Mirzapore (U.P.), India.

Vegetative form: 6 cysts found on four fish. The seat is immediately below the scales, in the epidermis. Color milky white. Soft, flattened and roughly oval in shape. Greatest length found, 1.1mm. No pigment was present on the cyst.

Spore: Two equal capsules, with a very short tail-like process. Sporoplasm with vacuole; iodine treatment could not be carried out. Dimensions: length 13μ , breadth 13μ , polar capsule 4μ by 4μ (?).

Remarks: Dimensions, especially that of polar capsule seem to be misprinted. Southwell gave one figure of a fish with a cyst near the dorsal fin. He thinks that "it is quite possible that our parasites may belong to Myxobolus cyprini." The incomplete observation without any figure, leads the writer to leave the form also as Myxobolus sp. Southwell.

MYXOBOLUS FUNDULI Kudo

[Figs. 474 to 476]

 1915
 Myzobolus musculi
 Hahn
 1915 : 201-205

 1917
 Myzobolus musculi
 Hahn
 1917 : 91-104

Habitat: Branchiae and muscle of Fundulus heteroclitus, F. majalis; Woods Hole. Hahn claims that he succeeded in causing experimental infection in F. diaphanus and Cyprinodon variegatus by inoculation.

Vegetative form: Hahn uses quite a number of different terms from those that are ordinarily used in describing Myxosporidia, without giving any definitions. Naturally it is hard to put what he wrote in several pages in the following lines. Granular vegetative forms produce a great many pansporoblasts, each with a single spore. "Trophoplasm" is difficult to stain. Size: 74 by 33μ , 24 by 19μ . Cysts within and between the muscle fibers, containing several hundred spores.

Spore: Hahn's descriptions may be summarized as follows: Dimensions: length 14.3μ , breadth 6.7μ , thickness 6.7μ to 2/3 of width, polar

capsule, 6.5μ by 2μ , polar filament 3 to 4 times the length of the spore (42.9 to 57.2μ). Polar filament coiled 10 to 14 times. Shell thin, almost invisible. The spores found in the gill: length 12 to 13.4μ , breadth 6μ to 10.4μ . A vacuole is present in the sporoplasm.

Remarks: Examination of Hahn's first paper suggested that he was dealing with the present form as a new species tho he did not mention at all Keysselitz who gave the name Myxobolus musculi Keysselitz to the parasite of Barbus fluviatilis from German rivers. I was informed by Hahn that he gave the name, Myxobolus musculi, without knowing the fact that it was preoccupied by Keysselitz (1908) (see page 148) and that tho he became aware of it later, he can not determine differences by which the two forms can be distinguished. A comparison of the descriptions of Keysselitz and Hahn, however, shows that these two forms differ in several respects. Hence the latter form is recorded here as a distinct species under the new name.

It is interesting to note that very similar forms, one without an iodinophilous vacuole at any stage of spore-formation (*Myxosoma funduli* Kudo, see page 125) and the other with a vacuole, occur in the same hosts in the same locality. As mentioned above, the reader is requested to refer to Hahn's original paper for further data.

MYXOBOLUS PLEURONECTIDAE Hahn [Fig. 477]

1917 Myzobolus pleuronectidas Hahn 1917: 160-161

Habitat: Subcutaneous muscular tissue of Pseudopleuronectes americanus; Woods Hole.

Vegetative form: Similar to that of Myxobolus funduli.

Spore: Hahn writes as follows: Shape and appearance resembles Myxobolus pfeifferi. Dimensions: length 14.5μ , breadth 11.9μ , polar capsules 6μ by 3.7μ .

MYXOBOLUS CAPSULATUS Davis [Fig. 478]

1917 Myxobolus capsulatus Davis 1917: 237

Habitat: Visceral connective tissues of Cyprinodon variegatus; Beaufort. Vegetative form: Irregular form. In the state of diffuse infiltration. Polysporous.

Spore: Pyriform, flattened. Polar capsules large and pyriform, filling almost entire cavity of the spore. Sporoplasm relatively small. Iodinophilous vacuole visible in living spore. Dimensions: length 16μ , breadth 10 to 11μ , polar capsules 11μ by 4μ , length of polar filament 84μ .

MYXOBOLUS NODULARIS Southwell et Prashad [Figs. 479 and 480]

1918 Myzobolus nodularis Southwell and Prashad 1918: 347

Habitat: In the muscles of Rasbora daniconius occurring in two fish on the sides, and in another as a globular cyst near the anus; Mirpur, Dacca (June). Type specimens, numbered P 52/1.

Vegetative form: Cysts rounded or slightly elongated, varying in length 3.5 to 3.8mm. and 2.3 to 2.8mm. in width. Creamy yellow in color, in one case appearing blackish owing to the large number of black granules scattered in its surface.

Spore: Ovoidal. Sutural ridge very wide (about 1/5 thickness of the spore). Two polar capsules of equal size, which show coiled polar filaments clearly. Dimensions: length 9μ , breadth 7.2μ , length of polar capsule 3.4μ , that of polar filament 18.3μ .

MYXOBOLUS HYLAE Johnston et Bancroft

[Figs. 591 to 593]

1888		Fletcher	1888:337
1890		Haswell	1890 : 661
1909	Myxobolus sp.	Johnston	1909:29
1910	Myxobolus sp.	Cleland and	
	-	Johnston	1910 : 25
1918	Myxobolus hylae	Johnston and	
		Bancroft	1918 : 171-175

Habitat: In the testes, vasa efferentia and oviducts of Hyla aurea; Sidney, Australia (April, other months not mentioned). Fletcher observed the parasites also in the urinary bladder of both sexes, which fact was not confirmed by Johnston and Bancroft on account of the scarcity of the material. The latter authors could not infect Hyla caerulea by feeding infected testes or the cysts, giving the conclusion that the parasite is specific to H. aurea. The male is more often attacked by the parasite than the female. The infected animal appeared sickly and emaciated. As to the infection in kidneys, they write as follows: In one male specimen both testes and both kidneys were affected, and the upper parts of the ureters adjacent to the kidneys were swollen and milky in appearance. In another, in addition to the testes, the adjacent kidney and mesentery were attacked. No spores have yet been detected by them in sections of the kidney tubules.

Vegetative form: Johnston and Bancroft describe as follows:

Cysts: in male, either imbedded in the tissue or may project freely into the coelom of the testes; in female, lying between the layers of the wall, being projected into the lumen of the oviduct. Size from those of microscopic dimensions up to 2 to 3mm. in diameter. In sections, the protoplasm



is differentiated into two regions. The outer layer (ectoplasm) surrounds the body as a thin, light-staining region, while the endoplasm being denser and of more or less granular structure filled with spores especially in the central portion.

Spore: Johnston and Bancroft describe as follows:

Form somewhat variable, caused by the reduction in length. Oval, egg-shaped or nearly circular in front view. Sutural ridge slightly thickened. Two pyriform polar capsules are located at the anterior end. Sporoplasm with an iodinophilous vacuole (2μ in diameter), shows usually two distinct nuclei, rarely but one. Dimensions: length variable, diameter of circular form 7 to 8μ , breadth 8 to 10μ , thickness about 6μ , thickness of shell 1μ , polar capsules 4 to 5μ by 2μ , length of polar filament 90 to 98μ (acids or alkalies).

MYXOBOLUS AUREATUS Ward [Figs. 643 to 649]

1919 Myxobolus aureatus

Ward

1919:49

Habitat: Between the ectodermal layers of the fin membrane of *Notropis anogenus*; Put-in-Bay, Lake Erie (August). Out of thirty fish, two to three cm. in length, seven were found to be infected. The infected fish were not inferior in size or vigor to others of the same species. The most heavily infected one was the most vigorous of all. The number of cysts in the individual fish, varied from one to forty, being confined in the fin. The cysts are always separated from each other, tho in a few instances they were apparently connected.

Vegetative form: The parasite forms cysts between the ectodermal layers of the fin membrane. The cyst is a smooth margined ellipsoid, measuring from 1 to 1.6 mm. in layer diameter and from 0.8 to 1.2 mm. along its transverse axis. The opaque cyst is of a clear orange yellowish color. This gilt color is contained in the cyst wall, fading away in alcohol and formol. The chromatophores of the skin of the host are distinctly more abundant on the cyst than in other parts of the skin, and the older the cyst the more abundant the chromatophores. The wall of the cyst is noticeably tough and thick. In section, the protoplasm shows a poor differentiation into ectoplasm and endoplasm. The former granular and reticular, covers the entire surface as a thin layer, while the latter is highly vacuolated, containing only mature spores. Polysporous.

Spore: Ovoid; slightly pointed anterior and rounded posterior ends in front view; slightly compressed in lateral view. Sutural ridge distinct. The shell is of moderate thickness, and bears a flange at the posterior half in some spores. Two pyriform polar capsules, frequently of slightly different dimensions, are at the anterior part of the spore. No intercapsular appendix is present. When the spore is allowed to stand for 24 hours

or more in water, the polar filaments are extruded. The binucleated finely granular sporoplasm shows an iodinophilous vacuole. Dimensions: length 12.4 to 13.5 μ , breadth 6.5 to 7.5 μ , thickness 5 μ , length of polar capsule 6 to 7 μ (rarely 7.5 μ), length of polar filament about 20 to 26 μ , diameter of iodinophilous vacuole about 2 μ .

MYXOBOLUS MIYAIRII nov. spec.

[Fig. 481]

1909 Myxobolus sp.

Miyairi

1909:130,131-132

Habitat: Intestinal wall of *Parasilurus asotus* L.; Fukuoka? (Nippon) Vegetative form: Cysts. Size rather small up to 0.5mm. Full-grown spores as well as those in developmental stages fill the central portion of cysts, while numerous nuclei are chiefly found along the periphery of endplasm.

Spore: Elongated elliptic. Two polar capsules of nearly same size. Sporoplasm with a comparatively large iodinophilous vacuole. Dimensions: length 13 to 14.5μ , breadth 6 to 7μ , length of polar capsules 4.5μ , length of polar filament 30 to 35μ .

Remarks. As the descriptions show the form and structure are distinguishable from other species, the writer establishes the present species.

MYXOBOLUS KOI nov. spec. [Figs. 482 to 485]

Habitat: In the connective tissue of the gill filament of Cyprinus carpio L.; Tokio (April). One fish was found infected in a slight degree.

Vegetative form: Cysts small and spherical; white in color. Size up to 230μ in largest diameter. The seat similar to *Myxobolus toyamai*. The structure of the cysts, observed in section preparations, is also similar to the above mentioned unicapsular Myxobolus.

Spore: Oval with attenuated anterior and rounded posterior ends in front view; elongated pyriform in side view. Shell comparatively thin. No marking on shell. Sutural ridge fairly well marked. No intercapsular appendix. Two polar capsules are pyriform, large, and of usually equal form and size. Coiled polar filament distinct in vivo. Sporoplasm rather small, finely granular, shows two nuclei in almost all spores. An iodinophilous vacuole is deeply stained by Lugol's solution. Dimensions: 14 to 16μ , breadth 8 to 9μ , thickness 5 to 6μ , polar capsule 8 to 9μ by 2.5 to 3μ , length of polar filament 72μ in average (KOH).

MYXOBOLUS ORBICULATUS nov. spec. [Figs. 566 to 576]

Habitat: Muscle of myotomes of *Notropis gilberti* J. et M.; Stony Creek, Ill. (November). The fish was kept alive in an aquarium from November

11, 1918, until March 10, 1919, when it was killed, being then nearly dead. The material was examined on March 15. A few isolated spores occurred in the muscle of *Notropis blennius* (Homer Park, Ill., November).

Vegetative form: In and between the muscle bundles of the myotomes. Size variable. Color opaque white under the dissecting microscope. Smallest rounded ameboid forms with a single or numerous nuclei, in the muscle bundle, have the size of from 10μ to 30μ in greatest diameter (Figs. 573 to 575). The largest form observed was 400μ by 120μ . Young forms without any differentiated protoplasm, shows indistinct granular and reticular structure with deeply staining spherical or ring-form chromatinic granules. The number of the nuclei increases with the growth of the body. Larger form (Fig. 576), spindle shape, circular in cross-section, lies with its long axis parallel to the muscle fibres. The protoplasm vacuolated, contained mostly mature spores. Spores were also found in the state of diffuse infiltration. Polysporous.

Spore: Form somewhat variable. Typical form almost circular, slightly pointed at the anterior end (Fig. 566) in front view; spindle shaped in profile (Figs. 569 and 570). Sutural ridge marked. Shell uniformly thick, usually exhibiting four triangular folds on the surface along the posterior margin (Figs. 566, 568 and 571). No intercapsular appendix. Two pyriform polar capsules are, as a rule, of the same size and form. Frequent occurrence of the inequality of the polar capsules together with abnormalities in the form of the spore, were noticed especially among comparatively young spores. The granular sporoplasm, shows two spherical nuclei when stained. The iodinophilous vacuole, spherical and 2μ in average diameter, is deeply stained with Lugol's solution. Dimensions of unstained preserved spores: length and breadth 9 to 10μ , thickness 6.5 to 7μ , polar capsule 6 to 7.5μ by 2.5 to 3μ .

MYXOBOLUS DISCREPANS nov. spec.

[Figs. 597 to 601]

Habitat: Branchial lamellae of Carpiodes difformis; Salt Fork, Urbana, U.S.A. (May). One fish caught, died (soon after the capture) two hours before being fixed. Length 8.5cm.

Vegetative form: The parasites formed numerous cysts on the branchial lamellae. Cysts slightly yellowish white and mostly rounded or elongated along the lamella, occur in groups, often occupying the entire lamella. Infection was fairly heavy. Every gill arch harbored ten to twenty cysts mostly on the outer surface. Size of the cyst varies, small rounded one 500μ in diameter up to elongated forms 2mm. by $0.5 \, \text{mm}$., the majority being from 0.5 to 1mm. in diameter. The cyst is surrounded by a thin connective tissue layer of the host. The protoplasm shows little differen-



tiation. The ectoplasm is a rather narrow zone around the entire body and the endoplasm is filled with various nuclei, several stages of developing pansporoblasts, and mature spores. Each pansporoblast produces two spores. Polysporous.

Spore: Approximately circular with broad anterior and more or less narrower posterior end in front view; broadly fusiform in profile. Shell uniformly thin with 5 to 6 markings on the posterior margin. Two polar capsules broadly oval and convergent, fill the anterior half of the spore. A small triangular intercapsular appendix presents. Coiled polar filament is fairly visible in vivo. The spores from the cysts which were fixed with alcohol-acetic and preserved in 95 per cent alcohol, showed the extrusion of the polar filament under the influence of potassium hydrate solution (35 per cent) even after a considerable length of time as is shown in the following:

Material fixed on May 29.

June 2; Extrusion took place in almost all spores.

June 10; Extrusion took place in almost all spores.

June 26; Extrusion took place in almost all spores.

July 28; Extrusion took place in almost all spores.

August 29; Extrusion took place in numerous spores.

September 29; Extrusion took place in about 70 per cent of the spores, some filaments being rather short, and not fully extended.

October 20; Extrusion took place in about 50 per cent of the spores, most filaments being short, and not fully extended.

Sporoplasm coarsely granular shows clearly two ring-form nuclei in fresh preparations. Dimensions of preserved spores: length 11.4 to 13.5 μ , breadth 9.5 to 11 μ , thickness 8.5 to 9.5 μ , polar capsule 5.5 to 6 μ by 3.5 to 4 μ , length of polar filament 50 to 55 μ .

Remarks: The present species differs from the hitherto known species. Myxobolus lintoni (page 138) and Myxobolus orbiculatus (page 155) which are the nearest to the present form, differ from Myxobolus discrepans in the host, organ of infection, vegetative form and form and structure of the spore.

MYXOBOLUS MESENTERICUS nov. spec.

[Figs. 628 to 631]

Habitat: In the mesentery, liver, spleen and wall of stomach, pyloric coecum, intestine, and gall-bladder of *Lepomis cyanellus*; Crystal Lake, Urbana, Ill. (June and July). Out of thirty-six host fish, 10 cm. in average length, seven were found to be infected. In every case, except one, the mesentery was the main seat of infection, harboring conspicuous cysts. The number of cysts found in the host body varied from three to seven. The infected fish did not exhibit any recognizable pathological changes. Other species of fish caught at the same time, were free from the infection.

Vegetative form: The cysts are mostly spherical in form, and are covered by a tough resistant envelope composed of the connective tissue of the host. They are uniformly white in color, and have the variable dimensions of from 0.5 to 1.5mm. in diameter. In section, the protoplasm shows a coarsely reticulated structure without distinct differentiation. In all cysts of various sizes fully mature spores were only observed. The spore formation could not be worked out. Polysporous.

Spore: Broadly oval with a slightly truncated anterior end in front view (Fig. 628), lenticular in side or end view (Fig. 629). No intercapsular appendix is seen. The shell is rather thick, and shows about eight folds on the sutural edge, two of which located laterally being more conspicuous than others. The sutural ridge is rather fine. Two convergent polar capsules equal in size occupy the anterior half of the spore. The coiled polar filament becomes more distinctly visible with the addition of Lugol's solution, altho it is faintly observable in fresh state. Fresh spores extruded their polar filaments under the action of potassium hydrate solution. In some spores, the extruded filaments cross each other near the The preserved spores showed no extrusion of the filament as in the last species. The sporoplasm is extremely finely granulated. The iodinophilous vacuole is comparatively large. When stained, the spore shows two nuclei in the sporoplasm. Dimensions of fresh material: length 10 to 11.5 μ , breadth 8.5 to 9.5 μ , thickness 6.5 μ , polar capsule 4.75 μ by 1.5 to 2μ , length of polar filament 32 to 40μ . Average dimensions of unstained preserved spores: length 9.5\mu, breadth 8\mu, polar capsule 4.75 μ by 2μ .

Remarks: The habitat and the structure of the spores, lead the writer to record the species as a new species.

Genus HENNEGUYA Thélohan

1892	Henneguya	Thélohan	1892 : 167, 176
1895	Henneguya	Thélohan	1895:352

The characters of the genus are described on page 59. Type species: Henneguya psorospermica Thélohan.

HENNEGUYA PSOROSPERMICA Thélohan [Figs. 486, 487 and 496]

		•	
1895	Henneguya psorospermica	Thélohan	1895 : 353
1896	Myxobolus psorospermica s. s	tr. Cohn	1896: 261
1899	Henneguya psoros permica		
	typica	Labbé	1899:101
1905	Henneguya psorospermica	Nufer	1905 : 77, 185
1910	Henneguya pseres permica	Wegener	1910 : 81-82
1911	Henneguya psorospermica		•
	lubica	Auerhach	1011 · 5 etc

Habitat: Branchiae of Esox lucius L. and Perca fluviatilis; France,

Frisches and Kurisches Haff, Pregel, Masurische Seen (all the year round, but rarer in Winter) Switzerland.

Vegetative form: Thélohan's observations on the structure of the cyst, are as follows: The surface of the cyst is covered by a layer, homogeneous, refringent and deeply stained, with which the cyst comes in direct contact with the surrounding epithelial cells of the host. Inside of this layer, there is a "pseudoectoplasmic" zone, in which the protoplasm is dense at places, forming radiate irregular striations, enclosing numerous irregular masses which are composed of apparently the same substance that forms the external layer. Toward the central portion of the cyst, there are masses of spores (Fig. 496).

Cohn's descriptions are as follows: The purely white cyst is elliptical; length 1.15mm. and breadth 0.85mm. The seat is under the epidermis. It is surrounded by the host tissue with small, elongated and scattered nuclei. The outer layer of the cyst is a thin membraneous protoplasm.

Wegener writes as follows: The white cysts are round or elliptical, usually on the upper end of the branchial lamella. Size of larger cysts, 1.5 to 2mm. long and 1.1 to 1.5mm. wide.

Spore: Elongated; anterior part fusiform and anterior end blunt. Polar capsules elongated and parallel to each other. Coiled polar filament visible in fresh conditions. Shell unstriated. Dimensions: total length 40μ in average, largest breadth 7μ , length of polar capsule 7 to 8μ .

Cohn's form is described by him as follows: Spore narrow with blunt anterior end. Sporoplasm with 6 horns (no figure to explain this expression!). When kept in water, sporoplasm takes round form and becomes highly refractive. Dimensions: length 29 to 38μ , length between the tip and the posterior margin of the cavity (15 to 20μ) 18μ , breadth 9 to 10μ , polar capsule (8 to 11μ) 9μ by 2μ , length of "starren Fäden" 14μ , length of tail 14 to 18μ .

Wegener's form is as follows: total length 35 to 38μ , breadth 7 to 8μ , length of the spore cavity 15μ , length of tail 15 to 20μ , polar capsule 8μ by 2 to 3μ .

HENNEGUYA TEXTA (Cohn) Labbé

1895	Myxobolus textus	Cohn	1895 : 38-39
1899	Henneguya psorospermica texta	Labbé	1899 : 101
1910	Henneguya texta	Wegener	1910 : 82-83

Habitat: Branchiae of *Perca fluviatilis* L.; Pregel, Frisches and Kurisches Haff (all the year round).

Vegetative form: Cohn observed as follows: Cyst distinctly elliptical. Length 0.75mm., breadth 0.375mm. The cysts surrounded by a thick layer of the host tissue. In the peripheral portion of the cyst, the protoplasm exhibits a network-like structure which forms a fibrous structure further inside, crossing the cyst at right angles to the long axis of the cyst.

Wegener writes as follows: The white cysts are elongated, 1.2 to 1.8mm. long and 0.5 to 0.7mm. wide.

Spore: Cohn mentions dimensions exactly the same as those of *Henneguya psoros permica* and can not distinguish the two species by the spore.

Wegener gives the following dimensions: length 30 to 40μ , breadth 7 to 8μ , length of the cavity of spore 15 to 18μ , length of tail 15 to 25μ , polar capsule 8μ by 2 to 3μ .

HENNEGUYA MINUTA (Cohn) Labbé [Figs. 488 and 489]

1895	Myxobolus minutus	Cohn	1895 : 39-40
1899	Henneguya psorospermica		
	minuta	Labbé	1899:102

Habitat: Branchiae of Perca fluviatilis L.; Frisches Haff, Lesina.

Vegetative form: Cohn's description is as follows: Cysts oval and small, difficult to distinguish them from those of *Henneguya psoros permica*. Size, 130μ by 115μ . The parasite was met only once. But the number of the cysts was far greater than that of *Henneguya psoros permica*, often 5 to 6 on one lamella, reaching up to 200 cysts on a single gillarch.

Spore: Cohn gives the following dimensions: total length (28 to 45μ) about 36μ , length from the tip to the end of cavity (20 to 28μ) about 26μ , breadth 10 to 11μ , thickness 8μ , polar capsule 11 to 14μ by 2 to 3μ , length of polar filament 42 to 45μ , length of tail (8 to 17μ) 12μ . Cohn gives a figure (Fig. 489) of a spore with two vacuoles(?).

HENNEGUYA OVIPERDA (Cohn) Labbé [Figs. 490 and 491]

1892		Weltner	1892 : 28-36
1895	Myxobolus oviperdus	Cohn	1895 : 40-41
1899	Henneguya psorospermica ovi-		
	perda	Labbé	1899:102
1904	Henneguya psoros permica ovi-		
	perda	Fuhrmann	1904 : 469-471
1911	Henneguya psorospermica ovi-		
	perda .	Auerbach	1911 : 5-22
1911	Henneguya psorospermica ovi-		
	per da	Nemeczek	1911 : 146

Habitat: Ovary of Esox lucius L.; Switzerland, Berlin, Frisches Haff (all the year round), Upsala (May), Austria (December).

Vegetative form: Cohn writes as follows: No real cyst exists. The parasite occupies the ovum.

Auerbach, however, mentions the presence of cysts in the connective tissue and follicle epithelium of the ovary. Dimensions, 1mm. up to 5 or 6mm. in diameter.

Spore: Cohn states the form and dimensions are very much similar to those of *H. psorospermica*.

HENNEGUYA LOBOSA (Cohn) Labbé [Figs. 492 and 493]

1895	Myxobolus lobosus	Cohn	1895:42
1899	Henneguya psorospermica lo-		
	bosa	Labbé	1899:102
1910	Henneguya(?) lobosa	Wegener	1910 : 83
1911	Henneguva(?) lobosa	Auerbach	1911 : 22-25

Habitat: Branchiae of *Esox lucius* L.; Frisches Haff, Pregel, Karlsruhe. Vegetative form: Cysts irregular in shape, size up to 2.5mm.

Wegener noticed that the cyst resembles that of Myxosoma dujardini with the dimensions of 2.2 to 2.8mm. by 1 to 1.1mm.

Spore: Cohn gives the dimensions as follows: total length 30 to 40μ , length from the tip to the posterior margin of cavity 11.5 to 15μ , breadth 5 to 6.5μ , polar capsules 6.5 to 8μ by 2 to 2.5μ , length of tail 22 to 28μ .

Wegener's form: oval; length 35 to 40μ , breadth 5μ , polar capsule 6 to 7μ by 2.5 to 3μ , length of the cavity of spore 13 to 15μ , length of tail 20 to 25μ , the iodinophilous vacuole could not be detected.

Auerbach gave the following dimensions: total length 30μ , breadth 4 to 6μ , length of polar capsule 6μ , length of polar filament 48 to 54μ .

Remarks: Wegener and Auerbach did not observe the iodinophilous vacuole.

HENNEGUYA PERI-INTESTINALIS Cépède

1906	Henneguya psorospermica peri-			
	intestinalis	Cépède	1906 : 67	
1907	Henneguya psorospermica peri-	•		
	intestinalis	Cépède	1907 : 137	
1912	Henneguya psorospermica peri-	•		
	intestinalis	Parisi	1912:295	

Habitat: Intestine of Esox lucius L.; Lac du Bourget, Pavia. (June). Vegetative form: Cysts.

Spore: Cépède mentions that it resembles that of Henneguya psoro-spermica.

HENNEGUYA MEDIA Thélohan [Figs. 494 and 495]

1890		Thélohan	1890 : 198-200
1892	Henneguya media	Thélohan	1892:177
1894	Myxobolus medius	Gurley	1894 : 248
1895	Henneguya media	Thélohan	1895 : 353
1898	Henneguya media	Doflein	1898 : 342

Habitat: Renal tubules of kidney and ovary of Gasterosteus aculeatus and G. pungitius L.; France. Mixed infection with Sphaerospora elegans.

Vegetative form: Rounded or elongated. In larger individuals, clear differentiation of protoplasm. Monosporous (?) and polysporous.

Spore: Fusiform. Shell striated. A vacuole in sporoplasm. Dimensions: length 20 to 24μ , breadth 5 to 6μ , polar capsules 4 to 5μ . Tail short.

HENNEGUYA BREVIS Thélohan

1854		Lieberk ühn	1854 : 357
1892	Henneguya brevis	Thélohan	1892:177
1895	Henneguya brevis	Thélohan	1895 : 354

Habitat: Similar to H. media Thélohan.

Vegetative form: Undescribed.

Spore: Fusiform with short tail. Dimensions: length 14 to 15μ , breadth 5 to 6μ , polar capsules 1.4 to 5μ , tail 4 to 5μ long.

HENNEGUYA SCHIZURA (Gurley) Labbé [Figs. 497 to 499]

1841		M üller	1841 : 477-478
1893	Myxobolus schizurus	Gurley	1893:417
1894	Myxobolus schizurus	Gurley	1894:255
1899	Henneguya schizura	Labbé	1899:102-103

Habitat: In cellular tissue of the eye muscles, in that of the sclerotic, and in that between the sclerotic and choroid of Esox lucius L.; Germany, U. S. A.

Vegetative form: Cysts white; membrane delicate; 0.44 to 1.09mm. in diameter.

Spore: Oval. Dimensions: length 12μ , breadth 6μ , thickness one-half the breadth, tail 3 to 4 times length of the body.

HENNEGUYA CREPLINI (Gurley) Labbé [Figs. 500 to 503]

1842		Creplin	1842 : 61-63
1894	Myxobolus creplini	Gurley	1894 : 248-249
1899	Henneguya creplini	Labbé	1899:103
1910	Henneguya creplini	Wegener	1910 : 84

Habitat: Branchiae of Acerina cernua L.; Pregel (March), Frisches and Kurisches Haff.

Vegetative form: Wegener describes as follows: Cysts, usually elongated oval and are located at the end of branchial lamella. Color white. Size 1 to 1.1mm. by 0.5mm. During winter, the cyst has only pansporoblasts, but no fully grown spores.

Spore: Creplin writes as follows: Elongated elliptical. Length 1/120", breadth 1/360", tail about as long as or a little longer than the body.

Wegener's form: elongated spindle shape; length 20μ , breadth 8 to 9μ , polar capsule 8μ by 2 to 3μ (parallel to each other).

Remarks: Wegener thinks that the present species and *Henneguya* acerinae Schröder, are one and the same species, and that the differences between the dimensions are due to the miscalculation of measurement in lines given by Creplin on the part of Gurley and Labbé.

HENNEGUYA LINEARIS (Gurley) Labbé [Fig. 504]

1841		Müller	1841 : 489
1893	Myxobolus linearis (part)	Gurley	1893:417
1894	Myxobolus linearis	Gurley	1894:255
1899	Henneguya linearis	Labbé	1899:103

Habitat: Membrane lining branchial cavity of *Pimelodus sebae* Cuv. et Val., branchiae of *Platystoma fasciatum* L.; South American rivers.

Vegetative form: Not described.

Spore: Very narrow. Length 3 to 4 times breadth.

HENNEGUYA GURLEYI Kudo

[Fig. 505]

1893	Myxobolus linearis (part)	Gurley	1893:417
1894	Myxobolus cf. linearis	Gurley	1894 : 253-254
1899	Henneguya linearis var.	Labbé	1899:103

Habitat: Base of spines of the second dorsal fin of Ameiurus melas Raf.; Iowa (Storm Lake) (August).

Vegetative form: Spherical cysts, 1mm. in diameter.

Spore: Lanceolate. Dimensions: length of the body 19μ , width 5 to 6μ , thickness about 3μ .

Remarks: The species is most probably different from *Henneguya linearis* judging from the difference in the form and structure of spores, the seat of infection, and host species. Hence, it is recorded here as an independent species.

HENNEGUYA STRONGYLURA (Gurley) Labbé

[Fig. 506]

1841		M üller	1841 : 480
1894	Myzobolus strongylurus	Gurley	1894 : 249
1899	Henneguya strongylura	Labbé	1899 : 103

Habitat: Skin of cephalic region of Synodontis schall Bl. Schn.; Nile.

Vegetative form: Cysts over 2.18mm. in diameter.

Spore: Dimensions: length of the body 9μ , breadth 5.4μ . Tail always undivided. Two polar capsules of equal size.

HENNEGUYA MONURA (Gurley) Labbé [Fig. 507]

1880		Ryder	1880 : 211-212
1893	Myxobolus monurus	Gurley	1893:416
1894	Myzobolus monurus	Gurley	1894 : 249-250
1899	Неппедиуа топига	Labbé	1899:103

Habitat: Subcutaneous intermuscular tissue of Aphredoderus sayanus Gill.; New Jersey (Woodbury).

Vegetative form: Cysts, lenticular, large, white, opaque and numerous (20). Membrane thin,

Spore: Lenticular or slightly obovate. Tail 2 to 3 times longer than the body.

HENNEGUYA KOLESNIKOVI (Gurley) Labbé [Fig. 508]

1886		Kolesnikov	1886 : 242-248
1894	Myxobolus kolesnikovi	Gurley	1894:256-257
1898	Myxobolus bicaudatus (part)	Zschokke	1898 : 602-604, 646-
			655, 699-703
1899	Henneguya kolesnikovi	Labbé	1899:103-104

Habitat: Interstitial connective tissue of the thoracic and intercostal muscles of *Coregonus lavaretus* L.; Russia.

Vegetative form: Cysts numerous (80), spherical or oval; length 10 to 30mm.. breadth 7 to 20mm.

Spore: Oval with a pointed anterior end. Tail three times longer than the body.

Remarks: Zschokke thinks the present species is identical with *Henneguya zschokkei*. But the evidence is not clear enough to bring one to agree with him due to the incomplete description of the present species.

HENNEGUYA MACRURA (Gurley) Thélohan

[Figs. 509 to 512]

1893		Evermann	1893:76
1894	Myxobolus macrurus	Gurley	1894:250-253
1895	Henneguya macrura	Thélohan	1895 : 35 4

Habitat: Subcutaneous connective tissue of head of Hybognathus nuchalis Ag.; Neches River, Texas (November, temperature of water 9°.4C.) Of frequent occurrence.

Vegetative form: Cysts, elongated 6mm. by 2mm. or less.

Spore: Rounded oblong. Dimensions: length 10 to 11μ , breadth 6 to 8μ , thickness 4μ . Shell-valves unequally convex. Tail 30 to 40μ .

HENNEGUYA ZSCHOKKEI (Gurley) Doflein

[Fig. 513]

1884		Zschokke	1884 : 234-235
1894	Myxobolus (?) zschokkei	Gurley	1894 : 244
1898	Myxobolus bicaudaius (part)	Zschokke	1898 : 602-607, 646- 655, 699-703
1898	Myxobolus bicaudatus	Zschokke	1898a : 213-214
1901	Henneguya zschokkei	Doflein	1901:202
1904	Henneguya zschokkei	Hofer	1904:56
1905	Henneguya zschokkei	Nufer	1905 : 77, 185

Habitat: Subcutaneous and superficial intermuscular tissue of *Coregonus fera*, *C. schinzii* Fatio, *C. hiemalis* Jur. and muscular tissue and branchia of *C. wartmanni nobilis* and *C. exiguus albellus*; Neuchâteler See, Zurich See, Genfer-see, Thuner-see, Vierwaldstätter-see.

Vegetative form: Zschokke writes as follows: Cysts rounded or oval surrounded by a compact membrane with many nuclei. The largest 32mm. by 16mm. Protoplasm granular. Polysporous.

Spore: Rounded oval in front view; broad elliptical in side view. Anterior end rounded; posterior end tapering, forming tail. Sutural ridge distinct. Tail is either bifurcated along the entire length or a single form, no intermediate form being observed. Dimensions: total length 55μ , length of the body 10μ , breadth 7μ , length of tail 4 to 5 times the length of the spore-body, length of polar filament 6 to 10 times that of the body of the spore.

Remarks: Zschokke thinks that *Henneguya kolesnikovi*, *H. zschokkei* and *H.* sp. Gurley are one and the same species, for which he proposed the name *Myxobolus bicaudatus*.

HENNEGUYA sp. (Gurley) Labbé [Fig. 514]

1886		Benecke	1886 : 211
1894	Myxobolus sp. inc.	Gurley	1894 : 24 4
1899	Henneguya sp.	Labbé	1899:104
1904	Henneguya sp.	Hofer	1904 : 51

Habitat: Integument (?) of Leuciscus rutilus L. The parasites formed boil-like enlargement in the skin.

Vegetative form: Not described.

Spore: Not described.

HENNEGUYA sp. (Gurley) Labbé

1874		Claparède	1874:114
1894	Myxobolus sp. inc.	Gurley	1894 : 253
189 8	Myxobolus bicaudatus (part)	Zschokke	1898 : 602-607, 646-
	•		655, 699-703
1899	Henneguva SD.	Labbé	1899 : 104

Habitat: Branchial-arches of Coregonus fera; Genfer-see.

Vegetative form: One cyst, 1mm. in diameter.

Spore: Tail short. Zschokke quotes: length 8 to 10μ .

Remarks: According to Zschokke, this species is identical with H. zschokkei.

HENNEGUYA TENUIS Vaney et Conte [Fig. 515]

1901 Henneguya tenuis Vaney and Conte 1901: 103-106

Habitat: Connective tissue of alimentary tract of Acerina cernua L.; Lyon (February).

Vegetative form: Numerous cysts particularly in the pyloric coecum. Usually spherical. Size: 30 to 150 μ in diameter.

Spore: Oval and small. Tail short. Two polar capsules at the anterior end. Sporoplasm with a nucleus, rod-shaped, with somewhat enlarged ends which is located at right angles to the longitudinal axis. Iodinophilous vacuole could not be traced. Dimensions: length 4μ , breadth 2μ .

HENNEGUYA NÜSSLINI Schuberg et Schröder [Figs. 516 and 517]

1905 Henneguya nüsslini Schuberg and Schröder 1905: 56-59

Habitat: Subcutaneous connective tissue at the base of dorsal fin of Trutta fario L.; Gutach.

Vegetative form: Trophozoites form cysts (2 cysts found). Cysts lenticular, 1.5 to 2mm., surrounded by many concentric layers of fibrous connective tissue. Cysts containing only mature spores.

Spore: Broad oval form, flattened. Anterior end rounded. Tail at the posterior end. Shell somewhat thick, often shows sutural ridge. Tail filaments two. A "dark part" which in side-view is of triangular form, runs into the tail. Sporoplasm, occupying the posterior half of the spore, projects a narrow portion between the polar capsules beyond the middle of the capsules. Sporoplasm, uniformly granular, contains an iodinophilous vacuole and one, sometimes two nuclei connected by nuclear bridge. Polar capsules, pyriform, opening independently. Coiled polar filament observable, coiled 6 to 7 times. Dimensions: length excluding tail 12μ , length with tail 32μ , breadth 8 to 9μ , polar capsules 5μ by 3μ , length of polar filament 4 to 5 times longer than that of spore excluding tail (48 to 60μ).

HENNEGUYA LÉGERI Cépède [Figs. 518 to 523]

1905	Henneguya légeri	Cépède	1905 : 905 -9 13
1906	Henneguya légeri	Cépède	1906 : 66
1913	Henneguya légeri	Cépède	1913 : 302-305

Habitat: Urinary bladder of Cobitis barbatula L.; Isère (January).

Vegetative form: Young trophozoites subcircular, irregularly elliptical or elongated with distinct differentiation of protoplasm into ectoplasm and endoplasm. Plasmotomic multiplication takes place during winter months, when no spore is formed.

Spore: Oval with short tail, mostly bifurcated at the free end. The anterior end is more rounded, occasionally acuminated. Two polar capsules of equal size. Coiled polar filament distinct in vivo. Sporoplasm granular, contains two nuclei and a vacuole. The spore often shrinks in fresh conditions, probably owing to the poorly developed thin valves. Dimensions of spores mounted in balsam: length variable. Examples: Total length 22.5μ , tail 8.5μ ; total length 19.5μ , tail 8μ ; length of main part 8.5μ , breadth (comparatively constant) 6μ .

HENNEGUYA ACERINAE Schröder [Figs. 525 and 526]

1906	Henneguya acerinae	Schröder	1906 : 186-196
1910	Henneguya creplini	Wegener	1910 : 8 4
1911	Henneguya acerinas	Nemeczek	1911:155

Habitat: Branchiae of Acerina cernua L., Aspro zingel Cuv., Lucioperca lucioperca L. and L. sandra Cuv. (?); Heidelberg (Necker), Apatin, Komitat Baco-Bodrog, Hungary (May).

Vegetative form: Schröder describes as follows: Rounded or spherical cysts in the connective tissue of branchial lamella. Full-grown cysts up to 300μ in diameter. Protoplasm is differentiated into ectoplasm and endoplasm. Ectoplasm shows fine radial striations. Endoplasm granular, contains many nuclei, especially lying in the middle portion. Well developed cyst, containing only spores, is surrounded by a membrane. On the surface of the ectoplasm, numerous edge-like elevations, branched and joining together, were recognized. Polysporous.

Nemeczek observed the largest cyst, spherical and 600µ in diameter.

Spore: Pyriform in front view; flattened. The anterior end is more or less blunt. Shell uniformly thin. Sutural edge slightly enlarged. Sporoplasm finely granular, contains an iodinophilous vacuole and two nuclei. Polar capsules approximated closely, each having an independent opening. Dimensions: length 20 to 22μ , breadth 8 to 9μ , thickness 6 to 7μ , length of tail 50 to 60μ , polar capsules 10μ by 2 to 3μ , length of polar filament 80 to 90μ (water and nitric acid).

Nemeczek's form is as follows:

The tail is bifurcated along its entire length. In one case (May, 1909), however, all the spores had no bifurcated tail, while the polar capsules were of unequal size. Dimensions in fresh state: total length 37.6 to 41.8 μ , length, excluding tail 12.6 to 16.8 μ , breadth 4.5 μ , length of polar capsule 6.3 to 8.4 μ , length of polar filament 67 μ , length of tail 25 μ .

Nemeczek observed two more different (?) forms. One form found in *Lucioperca sandra*, tho the size differs from the dimensions given by Schröder, is thought to be identical with the present species. Another form in the branchiae of *Aspro singel*, which is also to be one and the same species with the present species has the following dimensions: total length 35μ , length of spore excluding tail 15μ , breadth 5μ , length of polar capsule 6μ , length of tail 20μ .

HENNEGUYA GIGANTEA Nemeczek

[Figs. 527 to 535]

1911 Honneguya gigantea Nemeczek 1911 : 146-154 1914 Honneguya gigantea Georgévitch 1914 : 387-409

Habitat: Branchiae of Lucioperca sandra Cuv.; Apatin, Komitat Bacs-Bodrog, Hungary, Petrograd. Nemeczek mentions that the infection takes place only among young fish.

Vegetative form: Cysts numerous and of conspicuous size in the free end of branchial lamella. In average, each gill-arch has about 100 cysts which are of creamy color. Young cysts 400 to 450 µ in diameter. They gradually begin to increase the size, from autumn until toward the end of spring, during which period, the contents remaining in the stages of pansporoblasts formation. Older cysts rounded spindle shape with the length of 4 to 7mm, and the breadth of 2 to 3mm. The connective tissue and epithelial cell layers form the cyst membrane. The connective tissue either simply surrounds the parasite or branches in the surface of the parasite, increasing in thickness and forming more or less enclosed chambers of the parasite. The membrane of the cvst which contains mature spores is usually very thin. Throughout the growth of the cyst, "chromatoid body" is seen in the endoplasm, which appears first as a filiform structure, stained deeply with nuclear stain. Later they gather together and form a compact body, situated excentrically. Fine branches from it become directed toward the surface of the body, anastomosing each other so that a network is formed on the surface of the cyst. The latter develops small ovoidal or columnal bodies (1.2 μ long and about 1 μ wide), which are arranged radially and densely. The number and quantity of these bodies increase in proportion to the number of propagative nuclei and they begin to disappear, first in the central portion, then in the periphery, so that in fully grown cysts (in summer months) these chromatoidal bodies are more rudimentary. Differentiated protoplasm is only recognized in young individuals, in which case ectoplasm is homogeneous and endoplasm reticular. Polysporous.

Spore: Nemeczek gives the following accounts.

Spindle shape, with truncate anterior end and very long thread like tail at the posterior end. The tail seems split into two at about the middle part of its length. Gentian violet stains the tail so intensively that its entire length could easily be made out. Dimensions: total length 87.5 to 110.5μ , length of the body 10.5μ , breadth 5μ , length of tail 77 to 100μ , length of polar capsule 5μ , length of polar filament 70μ (pressure or dessication followed by immersion in water).

Georgévitch's form: length excluding tail 15μ , breadth 6μ , length of tail 75μ , length of polar capsule 6μ , length of polar filament 75μ , diameter of the iodinophilous vacuole 4μ .

Remarks: Nemeczek mentions that from October on, cysts had no spores, only containing propagative cells. The velocity of the development of spores depends upon the temerature of water.

Georgévitch worked out the spore formation of the species and observed that the binucleated sporeplasm emerged from the posterior end of the spore.

HENNEGUYA (?) sp. Nemeczek [Figs. 536 to 539]

1911 Henneguya sp.

Nemeczek

1911:157-159

Habitat: Branchiae of Abramis brama; Komorn, Komitat Komorn, Hungary (March).

Vegetative form: Cysts in the branchiae.

Spore: Besides normal spores of Myxobolus rotundus (page 149), spores of Henneguya type in small number were found. The anterior part of these spores resembles that of the species mentioned above, while the breadth is much smaller (8μ) than the latter. Majority of spores have a thread like tail, 10 to 15μ long, which was often bifurcated. An iodinophilous vacuole was fairly marked.

Remarks: It is placed here as a species of Henneguya by reason of the bifurcate tail.

HENNEGUYA GASTEROSTEI Parisi

[Figs. 540 to 543]

1912 Henneguya gasterostei

Parisi

1912:296-297

Habitat: Kidney of Gasterosteus aculeatus L.; Lago di Garda (February).

Vegetative form: Rounded or oval, usually with two, but rarely with four spores. Ectoplasm thin and hyaline. Endoplasm contains numerous granules, most probably of fatty nature and decreasing in number as spores grow. Free full-grown spores were seen abundantly in the connective tissue of renal tubules, glomeruli, etc. Disporous and polysporous.

Spore: Oval with slightly attenuated anterior end; posterior end tapering into tails, which end in one point or bifurcated; asymmetrical in shape, one valve is more curved than the other. This asymmetry of the shell-valves in profile enables the present species to be distinguished from other species. Shell striated longitudinally. Two polar capsules pyriform and well developed, reaching to the middle of the spore. Sporoplasm with a round iodinophilous vacuole. Dimensions: total length 38 to 48μ , length of the cavity of the spore 15μ , breadth 6 to 7.5μ , polar capsules 7.5 to 9μ by 3 to 3.5μ , length of polar filament 50μ .

HENNEGUYA NEAPOLITANA Parisi [Figs. 544 and 545]

1912 Henneguya neapolitana Parisi 1912: 297-298

Habitat: Connective tissue of the renal tubule of kidney of Box salpa C. et V.; Napoli (August).

Vegetative form: Small cyst (40 to 50μ in diameter) surrounded by thin membrane, containing a number of spores, numerous pigment granules and coarse yellowish globules.

Spore: Oval, slightly flattened. Anterior end rounded when seen from the front, but attenuated in profile. Shell tapering into a long fine tail posteriorly. The fine distal portion of the tail wraps around the thicker part. Two polar capsules, pyriform, occupying the anterior half of the cavity of the spore, cross each other when seen from the front. Sporoplasm finely granular with two nuclei, the iodinophilous vacuole being hardly visible. Dimensions: total length 50 to 60μ , length of the cavity of spore 8.5 to 9.5μ , breadth 8.5 to 9.5μ , internal breadth 6.3 to 7μ , thickness 8μ , polar capsules 4.7 to 5.5μ by 3μ .

HENNEGUYA WISCONSINENSIS Mavor et Strasser [Figs. 558 and 559]

1916 Henneguya wisconsinensis Mavor et Strasser 1916: 676-682

Habitat: Urinary bladder of Perca flavescens; Lake Mendota, Wisconsin (April).

Vegetative form: Trophozoites are usually elongated and have the general form and shape of a limax ameba. It may reach a size of 300μ by 70μ . Clear differentiation of ectoplasm and endoplasm. Pseudopodia lobose. Two spores are formed in each pansporoblast. Polysporous.

Spore: Ovoid, bilaterally symmetrical, and have a bifurcated caudal process. Two polar capsules at anterior end. Coiled polar filament visible in vivo (5 windings). Dimensions: length excluding tail 11.5μ , breadth 7μ , tail 9.6μ , polar capsules 3.5μ by 2.5μ , length of filament 33μ .

HENNEGUYA BRACHYURA Ward

[Figs. 650 to 653]

1919 Henneguya brachyura

Ward

1919:57

Habitat: In the cartilageous fin ray of the caudal fin of Notropis anogenus; Put-in-Bay, Lake Erie (August). The species was found encysted in the same fish which was heavily infected by Myxobolus aureatus.

Vegetative form: Cysts rounded with slightly irregular contour imbedded in the fin ray. The size varies from 160μ in diameter up to 360μ by 240μ . No particular cyst membrane could be recognized. The differentiation of the protoplasm into ectoplasm and endoplasm is distinct. The ectoplasm covering the entire surface of the parasite as a layer 4 to 6μ thick, shows structure of a very finely granular nature. The endoplasm coarsely alveolar, is filled with mature spores in the central portion, while numerous nuclei and young spores in various developmental stages are present at the peripheral portion. Polysporous.

Spore: Rounded oval in front view; spindle shape with symmetrically built valves in profile. Shell rather thick. Sutural ridge fairly well marked; sutural edge exhibiting a variable number of folds (8 to 10). Two pyriform polar capsules are usually of the same size and form. The tail is a single process, usually more or less bent or irregularly curved, very rarely being straight. In general, it is sinuous with two or three shallow curves and is rather short, tapering gradually to a point. In young spores which are less deeply stained by any stain, various developmental stages of the tail are reasily recognized. Giemsa solution stains the shell proper in clear blue, while the tail takes on a beautiful pink color, a distinct difference in affinity for dyes between the material in the tail and the shell. It seems probable that the tail of this type is entirely different in its development from that of the ordinary bifurcated type. Dimensions in section: length 10 to 11.5μ , breadth 8 to 8.75μ , thickness 4 to 5μ , polar capsules 3 to 4μ by 2μ , length of the tail up to 17μ .

HENNEGUYA SALMINICOLA Ward

[Figs. 654 to 656]

1914 ?Henneguya sschokkei Zschokke and Heitz 1914: 200-201
 1919 Henneguya salminicola Ward 1919: 59

Habitat: In the sub-dermal tissue of Onchorhynchus keta and O. kisutch (Zschokke and Heitz, Kamtschatka) and in the connective tissue in body muscles of Oncorhynchus keta, Stickeen River, Alaska (Ward, September). The last named author undertook a careful examination of a part of the infected tissue preserved in formol. The species forms conspicuous cysts in the muscle from the sub-peritoneal to the sub-dermal connective tissue, tho all are sub-peritoneal in position.

Vegetative form: Ward describes as follows: The whitish opaque cysts are pyriform, and fairly uniform in size (3 to 6mm. in diameter). The cyst is covered by numerous layers of connective tissue which form a tough membrane around the parasite. The cyst contains young spores in various stages of development, which showed that two spores are formed in one pansporoblast, and mature spores thickly massed together in the central area. Polysporous.

Spore: Oval with rounded anterior and more or less attenuated posterior ends; elliptical in profile with attenuated anterior end. Shell smooth. Sutural edge exhibits folds variable in number (usually 6 to 7). Tail double, composed of two fine and equal halves which are the prolongation of the shell valves. The processes usually run roughly parallel to each other. Two pyriform polar capsules are of slightly different dimensions. Coiled polar filament is indistinct in preserved unstained specimens. Sporoplasm finely granular, shows a large iodinophilous vacuole. Dimensions of stained and mounted spores: total length 47μ (42.75 to 52.44 μ), length of the main part 12μ (11.97 to 14.25μ), breadth 8μ (7.12 to 8.43μ), thickness 4.78μ , length of tail 35μ (30.78 to 38.19μ), polar capsule 3.70 to 4.55μ by 1.59 to 2.85μ .

Remarks: Zschokke and Heitz (1914) observed a species from Kamtschatka, which they thought to be identical with *Henneguya zschokkei* (page 165). The writer is inclined to think that the species is identical with the species just described from Alaska.

HENNEGUYA MIYAIRII nov. spec.

[Fig. 524]

1909 Henneguya sp.

Mivairi

1909:127-129

Habitat: Subcutaneous tissue of head of Carassius auratus L.; Fukuoka (Nippon).

Vegetative form: Trophozoites form cysts and are also found in the condition of diffuse infiltration around the cysts. Cyst-membrane fibrous and thin. Ectoplasm and endoplasm fairly well differentiated, though the border line is not sharply marked. At the periphery of endoplasm, pansporoblasts with 7 to 12 nuclei are present (Two spores are formed in each pansporoblast?). Polysporous.

Spore: Oval, with broadly rounded anterior and slightly elongated posterior ends, the latter ending in long and fine tails. Two polar capsules at the anterior portion, are pyriform, small and convergent. Sporoplasm with an iodinophilous vacuole. Dimensions: length 12μ , breadth 8μ , length of the tails 10 to 30μ , length of polar filaments 23 to 40μ .

Remarks: As the description gives the details by which the species can be distinguished from other species, the writer establishes it on an independent basis.

HENNEGUYA MICTOSPORA nov. spec. [Figs. 546 to 557]

Habitat: Urinary bladder of Lepomis cyanellus Raf., L. humilis Gir. and Micropterus salmoides Lac.; Stony Creek, Ill. (November).

In one out of three (6.5 to 8cm. long) of the first, in one out of two (7 and 9.5cm. long) of the second and in one of the third species, examined in the middle of November, was found the present form. None showed a heavy infection, a number of scattered trophozoites and spores being observed. The host did not show any pathological change.

Vegetative form: Polymorphous. Generally rounded or elongated oval. In small monosporous and disporous forms, the tail of the spores developed inside, is extruded from the body, so that these trophozoites show long processes (Figs. 546, 553, 555). Pseudopodia lobose, and extruded from the entire surface of the body (Fig. 547), tho sometimes they are well formed at one end of the body. Protoplasm is differentiated distinctly into ectoplasm and endoplasm. Ectoplasm is homogeneous and hyaline, forming the outer layer. Endoplasm is of reticular structure. The body is colorless, often yellowish, when the endoplasm is loaded with numerous yellowish coarse granules. The size varies from 6 or 7μ up to 60μ . In a rounded form of 38μ in longest diameter, five pansporoblasts, each developing two spores and many nuclei were observed. In another oval form of 45μ by 60μ in size, numerous nuclei were stained, showing that no development of pansporoblast has yet taken place. Disporous, polysporous and monosporous, tho of rare occurrence.

Spore: Broad spindle shape with attenuated anterior end. Shell rather thin. Each valve has 6 to 8 longitudinal striations on the surface. A long tail composed of two halves, is developed at the posterior end. Two pyriform polar capsules with distinctly visible coiled polar filament opens at the anterior tip. Sporoplasm, finely granular, contains an iodinophilous vacuole which is made distinctly visible by treating with Lugol's solution. When stained two typical nuclei are recognized in the sporoplasm. Dimensions of the fresh spores: length excluding tail 13.5 to 15μ , breadth 8 to 9μ , thickness 6 to 7.5μ , length of tail 30 to 35μ , often up to 40μ , polar capsule 5 to 6μ by 3μ , length of polar filament 40μ .

Genus HOFERELLUS Berg

 1898
 Hoferellus
 Berg
 1898 : 41

 1898
 Hoferia
 Doffein
 1898 : 288-289

The characters of the genus are described on page 59. Type and only species: *Hoferellus cyprini* Doflein.

HOFERELLUS CYPRINI Doflein [Figs. 577 to 581]

1898	Hoferia cyprini	Doflein	1898:289-290
1908	Hoferellus cyprini	Mercier	1908 : LIII-LIV
1910	Hoferellus cyprini	Plehn	1910 : 20-22

Habitat: In lumen and epithelial cells of renal tubules of kidney of Cyprinus carpio L.; France and Germany.

Vegetative form: Young trophozoites live in epithelium. Adults free in the urinary tubules. Form rounded or oval. No clear differentiation of protoplasm. Pseudopodium unobserved. Endoplasm contains numerous granules and many nuclei. Each pansporoblast forms two spores. Smaller individuals 20 to 30μ in diameter. Polysporous.

Spore: Pyramidal with two short tail-like processes at the posterior end, which are formed from the shell-valves like those of Henneguya. Between these two processes, rarely small protoplasmic pointed processes occur. Each shell-valve has 9 to 10 longitudinal striations on it. Two polar capsules at the anterior part, show clearly the coiled polar filaments. Sporoplasm has two neclei and an iodinophilous vacuole. Dimensions: total length 10 to 12μ , breadth 8μ , tail-process 2μ long, polar capsule 3μ long.

MYXOSPORIDIA GENERA ET SPECIES INCERTAE

Gen. et spec. incert. Leydig

1851		Leydig	1851 : 222
1894	Gen. et spec. incert.	Gurley	1894 : 186

Habitat: Cysts in the root of tongue of Chondrostoma nasus L.; Germany.

Gen. et spec. incert. Leydig

1851		Leydig	1851 : 222
189 4	Gen. et spec. incert.	Gurley	1894 : 186

Habitat: Heart (auriculo-ventricular valve) of Leuciscus rutilus L.

Gen. et spec. incert. Leydig

1851		Leydig	1851 : 223
1894	Gen. et spec. incert.	Gurley	1894 : 186

Gen. et spec. incert. Heckel et Kner

	•		
1851		Heckel and Kner	1851 : 12
1894	Gen. et spec. incert.	Gurley	1894 : 186-187

Habitat: Branchiae of Lucioperca lucioperca L.; Austria.

Gen. et spec. incert. Borne

1886		Borne	1886 : 211
1894	Gen. et spec. incert.	Gurley	1894:187

Habitat: Scomber scombrus L.

Genus incert. MERLUCII Perugia [Figs. 582 and 583]

1891	Myxosporidium merlucii	Perugia	1891 : 22-24
1894	Myxobolus? merlucii	Gurley	1894 : 242-243
1899	Myxobolus merlucii	Labbé	1899:100

Habitat: Gall-bladder of Merlucius merlucius L.; Italy.

Vegetative form: Various form. No differentiation of protoplasm. Disporous (?).

Spore: Oval, with two polar capsules.

Remarks: The species was placed in the genus Myxobolus by previous authors. The figures given by Perugia show that the spores are at least dimorphous. From the habitat and the disporous characters, one should place it rather in one of the genera of the Family Ceratomyxidae.

Genus incert. CONGRI Perugia [Figs. 584 and 585]

1891	Myxosporidium congri	Perugia	1891 : 24-25
1894	Genus incert. congri	Gurley	1894 : 182
1912	Myxobolus congri	Parisi	1912 : 28 4

Habitat: Gall-bladder of Conger conger L.; Genova.

Vegetative form: Floating in the bladder. Form variable. Movements incessant, slow and ameboid.

Spore: Not described.

Gen. et spec. incert. Linton [Fig. 590]

1891		Linton	1891 : 359-361
1894	Gen. et spec. incert.	Gurley	1894 : 182-183

Habitat: Subcutaneous tissue of *Notropis megalops* Raf.; Ohio (Black River; September, October).

Vegetative form: Cysts. Globular, discrete or aggregated into clusters, white, with minute patches of black pigment from host; size varying from 2.5mm. (single cyst) to 7mm. by 5mm. (clusters); cyst-membrane composed of connective tissue.

Spore: Top-shaped, somewhat flattened; with pointed anterior and broadly rounded posterior end. Shell thick, with elevated sutural ridge. Polar capsules could not be detected. Protoplasm finely granular. Dimensions: length 17μ , breadth 10μ , thickness 6μ .

Remarks: The cysts and figures of spores given by Linton suggest that it is most probably a unicapsular Myxobolus. As Linton could not detect (?) the polar capsule, tho his figures faintly show the said structure, it is placed in this group.

Gen. et spec. incert. Mingazzini

 1892
 Mingazzini
 1892 : 398

 1899
 Labbé
 1899 : 113

Habitat: Ovarian egg of Lacerta sp.

Vegetative form: Ameboid with hyaline pseudopodia and granular protoplasm.

Spore: Not observed.

Gen. et spec. incert. Nufer

1905 Myzobolus sp. . Nufer 1905 : 71, 77, 79, 85, 186

Habitat: In the connective tissue of branchia of *Chondrostoma masus*; Lake of Lucerne. A single cyst in a single host fish.

Vegetative form: Cyst white, and of 1mm. in diameter.

Spore: With two polar capsules at one pole and the sporoplasm. Dimensions or any other characters are not given.

Remarks: Altho Nufer placed the form in the genus Myxobolus, this must be brought into the present group in view of the fact that the iodin-ophilous vacuole was not detected, and that the observation is too incomplete to place it to any one of the genera.

Gen. et spec. incert. Mavor [Figs. 586 and 587]

1915 Mavor 1915 : 27–28, 32–33 1916 Mavor 1916 : 553–554

Habitat: Gall-bladder of Urophycis chuss; St. Andrews (July to September).

Vegetative form: Mavor writes as follows:

Attached, usually in large numbers, to the epithelium of the bladder, occurs a spherical or ellipsoidal trophozoite which in stained preparations is found to contain numerous nuclei. Very often clusters of *Ceratomyxa acadiensis* are found adhering to the free surface of myxosporidium. In fresh preparations the appearance is that of budding from a parent organism. An examination of sections has shown a sharp division between the myxosporidium and *Ceratomyxa acadiensis*.

Spore: Not found.

Remarks: Mavor supposed that the form under discussion probably was some species of Myxidium or Chloromyxum.

Gen. et spec. incert. Mavor [Figs. 588 and 589]

1916 Mayor 1916a : 68-69

Habitat: Urinary bladder of Stizostedion vitreum Mitch.; Georgian Bay (Canada).

Vegetative form: Free forms vary greatly in shape, being rounded, elongated or branched. The largest individual 200μ . Ectoplasm layer clearly visible, sometimes projecting many bristle-like short processes. Endoplasm contains greenish granules. Trophozoites also attached to the epithelium by means of deeply stainable portion of the body.

Spore: Not observed.

Remarks: Mavor mentions resemblance of the present form to Myxidium lieberkühni Bütschli in many respects.

named.

KEYS TO THE GENERA AND SPECIES OF MYXOSPORIDIA

No key to the genera and species of Myxosporidia has been published up to the present time. This is due of course to the difficulties which accompany such an attempt. These difficulties lie chiefly in the incompleteness of the observations and descriptions of the majority of the species of Myxosporidia.

The writer has attempted in the following pages to carry out this task. The key is by no means complete, as is unavoidable in the present state of knowledge concerning this particular group of the Protozoa.

Altho the spore is the fundamental factor used in constructing this key, it was necessary to refer also to some other secondary characters such as vegetative form and habitat.

Some authors are inclined to think that the difference in host species gives an ample basis on which to record the parasite as a new species. In some cases the parasite is specific in a certain host species while in other cases a number of different host species are infected by one and the same parasite. Therefore one can not lay much emphasis upon a difference of hosts in fixing the identification of a Myxosporidian.

KEY TO THE GENERA OF MYXOSPORIDIA

1(6)	Spore approximately spherical
	Suborder Sphaerosporea Kudo 1919 2
2(3)	Spore with four polar capsules
	Family Chloromyxidae Thélohan 1890
	Genus Chloromyxum Mingazzini 1890(183)
3(2)	Spore with two polar capsules
	Family Sphaerosporidae Davis 1917 4
4(5)	Sutural line of spore straight
	Genus Sphaerospora Thélohan 1892(185)
5(4)	Sutural line of spore sinuous
	Genus Sinuolinea Davis 1917(186)
6(1)	Spore not spherical
7(16)	Largest diameter of spore at right angles to sutural line; two polar capsules, one
	on each side of sutural line
	Suborder Eurysporea Kudo 1919
	Family Ceratomyxidae Doflein 1899 8
8(11)	Shell-valves prolonged laterally
9(10)	Shell-valves hemispherical or rounded
	Genus Leptotheca Thélohan 1895(179)
10(9)	Shell-valves conical; free end tapering to a more or less pointed end
	Genus Ceratomyza Thélohan 1892(180)
11(8)	Shell-valves rather elongated; circular in cross-section
12(13)	Spore rounded oblong; shell longitudinally striated; polar capsules pyriform, with
	or without long and fine posterior filaments
	Genus Mitraspora Fujita 1912 emend. Kudo 1919(183)
Nu	mbers enclosed in parentheses refer to the page of the article on which is found the description of the species

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13(12)	Spore angular, not rounded
14(15)	Spore pyramidal in front view; with its base at anterior end; with or without dis-
	tinct anterior processes; shell smooth
	Genus Myxoproteus Doflein 1898(183)
15(14)	Spore isosceles triangular in front view; anterior end attenuated; polar capsules
	spherical and large; shell with fine network-like ridges; with posterior fringe-
	like processes
	Genus Wardia Kudo 1919(183)
16(7)	Largest diameter of spore coincides with or at an acute angle to sutural plane;
	one or two polar capsules which are in sutural plane
	Suborder Platysporea Kudo 1919
17(22)	Spore fusiform; two polar capsules, one at each end of spore
2. (22)	Family MYXIDIDAE Thélohan 1892
18(21)	Spore more or less regularly fusiform; shell-valves symmetrical
19(20)	Polar filament fine and long
19(20)	Genus Myxidium Bütschli 1882(186)
20/10\	Polar filament thick and short
20(19)	
04 (40)	Genus Sphaeromyza Thélohan 1892(188)
21(18)	Spore semi-circular in front view; polar filament fine
00(45)	Genus Zschokkella Auerbach 1910(188)
22(17)	Spore not fusiform; with one or two polar capsules at anterior extremity 23
23(26)	Sporoplasm without iodinophilous vacuole
	Family MYXOSOMATIDAE Poche 1913
24(25)	Spore elongated ovoid in front view; anterior end mostly pointed
	Genus Myxosoma Thélohan 1892(189)
25(24)	Spore more or less rounded in front view
	Genus Leniospora Plehn 1905(189)
26(23)	Sporoplasm always with iodinophilous vacuole
	Family MYXOBOLIDAE Thélohan 1892
27(30)	Spore with posterior process; shell sometimes striated
28(29)	Process more or less long, projecting posteriad along median line of spore; process
	either single or double; shell sometimes striated
	Genus Henneguya Thélohan 1892(193)
29(28)	Process short projecting posteriad from sides; shell longitudinally striated
	Genus Hoferellus Berg 1898(173)
30(27)	Spore without posterior process; shell unstriated; one or two polar capsules
	Genus Myxobolus Bütschli 1882(189)
	•
	II. KEY TO THE SPECIES
	Genus LEPTOTHECA Thélohan 1895
4/44	
1(14)	Spore: sutural diameter always more than half of greatest breadth
2(7)	Average sutural diameter less than 10μ
3(4)	Posterior margin of spore concave in front view; sutural diameter 8 to 9μ , breadth
	12 to 14μ. Trophozoite usually with a long process
	Leptotheca longipes Auerbach 1910(63)
4(3)	Posterior margin of spore not concave
5(6)	Posterior margin of spore flattened; polar capsules pyriform; sutural diameter 6
	to 7μ , breadth 11 to 12μ . Trophozoite with actively motile long filiform
	pseudopodia at rounded end
	Leptotheca agilis Thélohan 1892(60)
Nur	nbers enclosed in parentheses refer to the page of the article on which is found the description of the species

named.

6(5)	Spore regularly ovoidal; polar capsules short pyriform, opening on opposite sides; sutural diameter 9 μ , breadth 16 μ . Trophozoite pyriform without any recognizable pseudopodium
	Leptotheca fusiformis Davis 1917(63)
7(2)	Average sutural diameter of spore equal to or larger than 10μ
8(13)	Shell-valves symmetrically built; sutural ridge straight
9(10)	Posterior margin of spore concave in front view; sutural diameter 10μ , breadth 18 to 20μ ; each spore formed independently
10(0)	Leptotheca informis Auerbach 1910(63)
10(9) 11(12)	Spore regularly ovoidal
	Leptotheca polymorpha Thélohan 1895(61)
12(11)	Typical form of trophozoite elongated; anterior end depressed surrounded by short often branched pseudopodia. Spore: sutural diameter 12 to 15μ , breadth 18 to 20μ
	Leptotheca elongata Thélohan 1895(60)
13(8)	Shell-valves asymmetrically built; sutural ridge sinuous; sutural diameter 9 to 10μ , breadth 16 to 18μ . Trophozoite rounded; movements slow
	Leptotheca lobosa Davis 1917(64)
14(1)	Sutural diameter equal to or less than half of greatest breadth
15(18)	Average sutural diameter smaller than 10μ
16(17)	Spore arch-snaped in front view; polar capsules pyritorm; sutural diameter 3 to 4μ , breadth 8 to 10μ
	Leptotheca parva Thélohan 1895(61)
17(16)	Spore cylindrical; sutural diameter 4.5 \(\mu\), breadth 9 \(\mu\) Leptotheca glomerosa Davis 1917
18(15)	Average sutural diameter greater than 10μ
19(20)	Posterior margin of spore slightly concave in front view; anterior end attenuated; polar capsules pyriform; sutural diameter 13μ , breadth 26μ . Trophozoite
	rounded; with active amoeboid movements Leptotheca macrospora Auerbach 1909(62)
20(19)	Posterior margin of spore more or less flattened; anterior end smoothly rounded;
20(17)	polar capsules rounded; sutural diameter 11 µ, breadth 22 µ. Trophozoite
	elongated; pseudopodia often anastomose Leptotheca scissura Davis 1917(64)
Income	letely described species
- 	Leptotheca renicola Thélohan 1895(61)
	Leptotheca hepseti Thélohan 1895(62)
	Leptotheca perlata Gurley 1894(62)
	Leptotheca sp. Awerinzew 1908(62)
	Genus CERATOMYXA Thélohan 1892
1(52)	Spore constant in form and size
2(21)	Sutural diameter equal to or less than one-eighth of total breadth
3(10)	Sutural diameter not less than one-tenth of total breadth
4(9)	Pseudopodia of vegetative form located at rounded end 5
5(6)	Pseudopodia long filiform; with slow whiplash-like movements toward pointed
	extremity. Spore: sutural diameter 12 \mu, breadth 118 \mu
	Ceratomyxa flagellifera Davis 1917(77)

Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named.



6(5) 7(8)	Extremities of spore attenuated; spore large; sutural diameter 10 to 12 \mu, breadth
	90 to 100μ
	Ceratomyza sphaerulosa Thélohan 1892(66)
8(7)	
	39μ
	Ceralomyza streptos pora Davis 1917(79)
9(4)	Pseudopodia unlocalized; from main part of sporulating trophozoite are branched out from one to six long prolongations. Spore: sutural diameter 5 to 7μ ,
	breadth 50μ
	Ceratomyxa appendiculata Thélohan 1892(67)
10(3)	Sutural diameter equal to or less than one-tenth of total breadth
11(14)	Shell-valve terminating in a fine thread-like process at distal end
12(13)	Sutural diameter 7 to 8 \mu, breadth 40 to 50 \mu, lateral process 250 to 300 \mu
(-0)	Ceratomyxa acadiensis Mavor 1915(71)
13(12)	Sutural diameter 5μ , breadth 10 to 12μ , length of lateral process 20μ
10(12)	Ceratomyxa linospora Doflein 1898(69)
14/11\	
14(11)	Shell-valves not terminating in thread-like processes
15(20)	Shell-valve drawn out into a delicate process
16(17)	Lateral process ribbon-like; sutural diameter 6µ, breadth 140 to 150µ
47446	Ceratomyza taenia Davis 1917(74)
17(16)	Lateral process not ribbon-like, but circular in cross-section
18(19)	Posterior margin of main part of spore flattened; sutural diameter 12µ, breadth
	115 to 140μ; trophozoite disporous
	Ceratomyza sphairophora Davis 1917(73)
19(18)	Posterior margin of main part of spore rounded; sutural diameter 7μ , breadth
	80μ. Trophozoite monosporous or disporous
	Ceratomyxa spinosa Davis 1917(80)
20(15)	Shell-valve tapering gradually to attenuated point; asymmetrical; sutural diame-
	ter 9μ , breadth 115μ
	Ceratomyxa attenuata Davis 1917(75)
21(2)	Sutural diameter more than one-eighth of total breadth
22(45)	Sutural diameter equal to or more than one-fifth of breadth
23(34)	Shell-valves symmetrically built
24(29)	Shell-valves attenuated at distal end
25(26)	Pseudopodia peculiar network-like form. Spore: sutural diameter 12 to 20µ,
	breadth 50 to 80 μ
	Ceratomyxa ramosa Awerinzew 1907(69)
26(25)	Pseudopodia never unite together
27(28)	Shell-valves curved greatly posteriad; polar capsules rounded; sutural diameter
()	8 to 9μ , breadth $16(?)\mu$. Trophozoite elongated pyriform.
	* Ceratomyxa recurvata Davis 1917(75)
28(27)	
20(21)	sutural line; polar capsules pyriform; sutural diameter 40 to 45 \mu, thickness 25
	to 30μ , breadth 124 to 140 μ
	Construction Advances Appendix 2 (70)
20(24)	Ceratomyxa tylosuri Awerinzew 1913(70)
29(24)	Shell-valve rounded at distal end
30(31)	Spore arch-shaped; sutural diameter and thickness 12 to 15μ , breadth 50 to 60μ .
	Trophozoite large amoeboid.
a4 (aa)	Ceratomyxa spari Awerinzew 1913(70)
31(30)	Spore straight
	abers enclosed in parentheses refer to the page of the article on which is found the description of the species
named.	

32(33)	Shell-valves shorter (sutural diameter: breadth = 1:1.6)
	Ceratomyxa coris Georgévitch 1916(72)
33(32)	Shell-valves longer (sutural diameter: breadth = 1:2.6)
	Ceratomyxa herouardi Georgévitch 1916(72)
34(23)	Shell-valves asymmetrically built
35(40)	Spore arch-shaped in front view
36(37)	Sutural diameter equal to one-fifth of breadth; sutural diameter 10μ , breadth 50μ
	Ceratomyxa globulifera Thélohan 1895(67)
37(36)	Sutural diameter more than one-fifth of total breadth
38(39)	Spore: breadth shorter; sutural diameter 14 \mu, breadth 17 \mu
	Ceratomyxa abbreviata Davis 1917
39(38)	Spore: breadth longer; sutural diameter 12 to 15 \(\mu\), breadth 45 to 50 \(\mu\)
	Ceratomyxa reticularis Thélohan 1895(68)
40(35)	Spore straight 41
41(42)	Sutural diameter 11 \mu, breadth 27 \mu. Trophozoite always rounded
	Ceratomyxa amorpha Davis 1917(78)
42(41)	Sutural diameter 6µ
43(44)	Trophozoite with active pseudopodia. Spore: sutural diameter 6μ, breadth
	22 to 24μ
	Ceratomyxa undulata Davis 1917(79)
44(43)	Trophozoite with inactive pseudopodia. Spore: sutural diameter 6µ, breadth
	31μ
	Ceratomyxa inaequalis Doflein 1898
45(22)	Sutural diameter less than one-fifth of total breadth 40
46(49)	Breadth of spore equal to or greater than 50μ
47(48)	Shell-valve tapering gradually toward distal end; sutural diameter 8 µ, breadth
	50 to 56μ. Trophozoite usually elongated pyriform
	Ceratomyxa mesospora Davis 1917(73)
48(47)	Shell-valve rounded at distal end; sutural diameter 6 to 7μ , breadth 50μ . Tro-
	phozoite usually rounded or irregular form; size small
	Ceratomyxa aggregata Davis 1917(79)
49(46)	Breadth of spore smaller than 30μ
50(51)	Trophozoite ordinarily spherical, diameter not exceeding 16 to 20μ; protoplasm
, ,	extremely pale looking. Spore: sutural diameter 5μ , breadth 25 to 30μ .
	Ceratomyxa pallida Thélohan 1895(67)
51(50)	Trophozoite pyriform with a long posterior process; movements by wavelike
	motion of ectoplasm; also active backward movements of pseudopodia. Spore
	asymmetrically built; sutural diameter 5 \mu, breadth 24 to 28 \mu
	Ceratomyxa agglomerata Davis 1917(77)
52(1)	Spore variable in size and form
53(54)	Variation in number of shell-valves conspicuous; sutural diameter 5μ, breadth
	25μ
	Ceratomyxa truncata Thélohan 1895(67)
54(53)	Variable in size and form of spore, but not in number of shell-valve 55
55(60)	Trophozoite more or less definite in shape
56(59)	Trophozoite usually pyriform
57(58)	Trophozoite disporous. Spore: sutural diameter 7 to 9μ , breadth 15 to 38μ
. ,	Ceratomyxa lunata Davis 1917(76)
58(57)	Trophozoite monosporous or disporous. Spore: sutural diameter 5 to 6µ,
	breadth 18 to 25 μ
	Ceratomyxa monospora Davis 1917(78)

Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named.



59(56) Trophozoite always rounded, never pyriform. Spore: sutural diameter 6μ , breadth 16μ
Ceratomyxa navicularia Davis 1917(80
60(55) Trophozoite polymorphous
61(62) Shell-valves symmetrically built; sutural diameter 5 to 8μ , breadth 20 to 31μ
Ceratomyza arcuata Thélohan 1892
62(61) Shell-valves often asymmetrically built; sutural diameter 8 to 14μ , breadth 50 to 80μ
Ceratomyxa drepanopsettae Awerinzew 1907(70
Incompletely described species
Ceratomyxa sp. (?) Awerinzew 1913
Ceratomyxa sp. (?) Awerinzew 1913(71
Ceratomyxa sp. Georgévitch 1916(72
Genus MYXOPROTEUS Doflein 1898
1(2) Spore with two long (5μ) processes extending anteriad from sides; sutural diameter
9μ , breadth 12μ
Myxoproteus cornutus Davis 1917(82
2(1) Spore without long process
3(4) Spore with two small spinous processes at anterior end; sutural diameter
25μ, breadth 18 to 20μ (81)
Myxoproteus ambiguus Thélohan 1895
Spore without any process; posterior end singlity pointed; sutural diameter 12μ , breadth 10 to 11μ
Myxoproleus cordiformis Davis 1917(81
III yaopiokas cotasjornas Davis 1717
Genus WARDIA Kudo 1919
Spore isosceles triangular form; shell with network-like striations which end in fringe-like processes at posterior margin; sutural diameter 9 to 10μ, breadth 10
to 12μ , diameter of polar capsule 4μ .
Wardia ovinocua Kudo 1919(82
Doubtfully placed species
Wardia ohlmacheri (Gurley 1894)(83
Genus MITRASPORA Fujita 1912 emend. Kudo 1919
·
 1(4) Spore with posterior filaments 2(3) Posterior filaments short (5 to 6μ long); length 10μ, breadth 8 to 9μ, thickness
6 to 8μ , polar capsule 4μ by 1.5 to 2μ
Mitraspora cyprini Fujita 1912
3(2) Posterior filaments of spore long (up to 28μ); length 10 to 11μ , polar capsule 4 to
4.5μ long
Mitraspora caudata Parisi 1910(85
4(1) Spore without posterior filament; anterior end slightly attenuated; posterior end
truncate; length 15 to 17μ , breadth 5 to 6μ , thickness 4.5 to 5.5μ , polar cap-
sule 7.5 μ by 2 μ
Mitraspora elongata Kudo 1919(85
Genus CHLOROMYXUM Mingazzini 1890
1(4) Spore with posterior appendage
2(3) Posterior appendage fine and numerous; length 6 to 9μ , breadth 5 to 6μ , polar
capsule 2 to 3μ by 1 to 2μ
Chloromyxum leydigi Mingazzini 1890(87)
Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named.

3(2)	Posterior appendage single or bifurcated; length 8μ , breadth 6 to 7μ , appendage 10μ long
	Chloromyxum coudatum Thélohan 1895(88)
4(1)	Spore without posterior appendage
5(34)	Spore circular or subcircular in front view; parasitic in body cavity of host 6
6(29)	Shell-valves marked with striations or ridges
7(24)	Main part of striations or ridges parallel to sutural line
8(11)	Shell-valves partially marked9
9(10)	Ridges on each shell-valve variable in number (six found in original drawing)
	running closely to sutural line; diameter 10.8µ
	Chloromyxum dubium Auerbach 1908(91)
10(9)	Each shell-valve with one ridge from which eight to twelve short ones are directed
, ,	toward centre of valve; oval in profile; length and breadth 8 to 10 \mu, thickness
	5 to 7μ
	Chloromyxum trijugum Kudo 1919(96)
11(8)	Entire shell-valve marked
12(19)	Shell-valve marked with fine striations
13(16)	Spore oval in lateral view
14(15)	Trophozoite larger; size up to 50μ by 20μ ; polysporous (up to eight spores) rarely
11(10)	disporous. Spore: length 8 to 9μ , breadth 6 to 7μ , thickness 5 to 6μ
	Chloromyzum misgurni Kudo 1916(93)
15(14)	Trophozoite smaller; size up to 35μ in diameter; polysporous (up to six spores)
15(14)	or disporous. Spore: length 8μ , breadth 7μ , thickness 5 to 6μ
	Chloromyzum catostomi Kudo 1919(98)
16(13)	Spore circular in lateral view
17(18)	Trophozoite rounded; 40 to 45μ by 28 to 40μ . Spore: diameter 10 to 13μ , polar
17(10)	capsule 4 to 6μ long
18(17)	Chloromyxum protei Joseph 1905
10(1/)	Trophozoite irregular form; 33 to 35 \mu in average length. Spore: striations
	thicker and somewhat wavy; diameter 9 to 9.5 μ , polar capsule 3 μ long
10/12\	Chloromyxum thymalli Lebzelter 1912(92)
19(12)	Shell-valves marked with ridges
20(21)	Trophozoite small (average diameter of adults about 20μ); monosporous, rarely
	disporous. Spore: shell-valves with ridges marked antero-posteriad; diameter
	10 to 11µ
04 (00)	Chloromyxum cristatum Léger 1906(91)
21(20)	Trophozoite large, diameter reaching 40 to 50μ
22(23)	Ridges on shell-valves united into a line at each end and unequal in thickness;
	spore small; length 10 to 12\mu, breadth 8 to 10\mu Chloromyxum fujitai Kudo 1916(93)
	Chloromyxum fujilas Kudo 1916(93)
23(22)	Shell-valve with two circular and two small ridges; spore large; length 16 μ ,
	breadth 10μ
	Chloromyxum koi Fujita 1913(92)
24(7)	
25(26)	Striations irregular; posterior margin thickened at sides; diameter 7.5 to 9μ
	Chloromyxum wardi Kudo 1919(99)
26 (25)	Striations parallel to each other
27(28)	Striations forming acute angles with sutural line; diameter 8 to 9μ
	Chloromyxum truttae Léger 1906(90)
28(27)	Four ridges on posterior half of shell-valve converging toward anterior end;
	diameter 7 μ
	Chloromyxum granulosum Davis 1917(96)
Nun	aber senclosed in parentheses refer to the page of the article on which is found the description of the species
named.	

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Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species

Sphaerospora rostrata Thélohan 1895......(101)

Sphaerospora polymorpha Davis 1917.....(102)

spines at posterior margin; length 12 to 14μ , breadth 10 to 12μ

13(12) Spore smooth-contoured; polar capsules parallel to each other; diameter 7 to 10μ

11(11)	polar capsules unequal in size; length 7 to 8μ , breadth 6 to 7μ , thickness 5μ
-	Sphaerospora angulata Fujita 1912(102)
Incomp	letely described species
	Sphaerospora sp. Davis 1917
	Sphaerospora sp. Southwell et Prashad 1918
	Genus SINUOLINEA Davis 1917
1(4)	Spore with two processes
2(3)	Processes lateral and long (20 μ); spore: 9 to 11 μ long, 9 μ broad, process 18 to 22 μ long
	Sinuolinea brachiophora Davis 1917(106)
3(2)	Processes posteriad from sides and short; diameter 12 to 13 µ. Trophozoite
	opaque
	Sinuolinea opacita Davis 1917(106)
4(1)	Spore without process
5(6)	Trophozoite with active amoeboid movements. Spore: sutural ridge S-shaped
	at anterior part; length 15 μ , breadth 12 μ , thickness 8 μ
4/ E\	Sinuolinea arborescens Davis 1917(105)
6(5) 7(8)	Trophozoite with slow amoeboid movements
1(0)	than half of sporal cavity; polar capsules opening on opposite sides; diameter
	12 to 14μ
	Sinuolinea capsularis Davis 1917(105)
8(7)	
` ,	Sinuolinea dimorpha Davis 1916(104)
	Genus MYXIDIUM Bütschli 1882
1(16)	Breadth of spore equal to or more than half of length
2(7)	Shell-valves unstriated
3(6)	Sutural plane curved into an S
4(5)	Spore small; length 8 to 12 μ , breadth 4 to 6μ
	Myxidium incurvatum Thélohan 1892(108)
5(4)	Spore large; much broader; length 20.8 to 23.4 μ , breadth 13 to 15.6 μ
	Myxidium inflatum Auerbach 1909(111)
6 (3)	Sutural plane straight; spore cylindrical; surrounded by a gelatinous envelope;
	length 10 to 11 μ , breadth 6 μ
7/ 0\	Myxidium glutinosum Davis 1917(115)
7(2) 8(9)	Shell-valves striated
0(9)	Sutural line curved into an S; form oval; circular in cross-section; openings of
	polar capsules pointed; length 11 to 13 \(\mu\), breadth 8 to 9 \(\mu\) Myxidium oviforme Parisi 1912(114)
9(8)	Sutural line straight
10(13)	Sutural line coincides with longitudinal axis of spore
11(12)	Sutural ridge distinct; extremities mucronate; length 9 to 10μ , breadth 5 to 5.6μ ,
	thickness 4.75 to 5 μ . Vegetative form produces cysts, 800 to 900 μ in diameter
	Myxidium giardi Cépède 1906(110)
12(11)	Sutural ridge faintly marked; extremities gradually drawn out; length 11 µ,
	breadth 8µ. Trophozoite large and leaf-like; diameter up to 1.35 mm.
	Myxidium phyllium Davis 1917(116)
	abers enclosed in parentheses refer to the page of the article on which is found the description of the species
named.	

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13(10)	Sutural line forming an acute angle with longitudinal axis of spore
14(15)	Shell thickened at extremities; polar capsules ovoidal; length 10 to 14 µ, breadth
	6 to 8μ , length of polar capsule 4μ
45/44	Myxidium striatum Cunha et Fonseca 1917(116)
15(14)	Shell uniformly thick; polar capsules rounded pyriform; length 10 to 12 \mu, breadth
	6μ , length of polar capsule 3 to 4μ
467.43	Myxidium macrocapsulare Auerbach 1910(113)
16(1)	Breadth of spore less than half of length
17(34)	Breadth more than one-third of length
18(25) 19(22)	Shell-valves unstriated
	Extremities of spore pointed
20(21)	spore: extremities snarply pointed; greatly curved; narrow; length 12 to 14 μ , breadth 5.5 to 6μ , thickness 2.5 to 3μ
	Myxidium depressum Parisi 1912(114)
21(20)	Spore: extremities not so sharply pointed; not greatly curved; broader; length
21(20)	16.2 to 19μ , breadth 7 to 9μ
	Myxidium bergense Auerbach 1909(112)
22(19)	Extremities of spore not pointed
23(24)	Spore larger; length 15 to 20μ , breadth 7 to 8μ
23(24)	Myxidium sphaericum Thélohan 1895(109)
24(23)	Spore smaller; length $6(?)$ to 14μ , breadth 4 to 6μ . Trophozoite microsporous
24(20)	Myxidium gadi Georgévitch 1916(115)
25(18)	Shell-valves striated
26(33)	Spore definite in shape
27(28)	Spore constricted in middle part of length; length 15 μ
(,	Myxidium histophilum Thélohan 1895(109)
28(27)	Spore regularly fusiform
29(30)	Vegetative form produces cyst. Spore: length 12 to 15μ, breadth 6μ
	Myxidium barbatulae Cépède 1906(110)
30(29)	Vegetative form does not produce cyst
31(32)	Sutural line slightly curved in S-form; length 15 to 16 µ, breadth and thickness
` '	5.5 to 6μ
	Myxidium americanum Kudo 1919(117)
32(31)	Sutural line not curved in S-shape, but bent to one side; length 15 to 18 \mu, breadth
	and thickness 6 to 7 μ
	Myxidium kagayamai Kudo 1919(117)
33(26)	Spore variable in form; straight and constricted; one side concave, the other con-
	vex; arch-shaped, etc.; length 13 to 18μ , breadth 5.2 to 5.8μ
	Myxidium pfeifferi Auerbach 1908(111)
34(17)	Breadth of spore equal to or less than one-third of length
35(40)	Shell-valves unstriated
36(37)	Spore greatly elongated (breadth: length = 1:6.2); length 21.6 to 25.2 \(\mu\), breadth
	3.6 to 4μ
	Myxidium procerum Auerbach 1910(112)
37(36)	Spore less elongated (breadth: length=1:3 or 1:3.4)
38(39)	Spore large; valves asymmetrical; length 28μ , breadth 8μ
	Myxidium giganteum Doflein 1898(110)
39(38)	Spore small; valves symmetrical; length 12μ , breadth 3 to 4μ
	Myxidium danilewskyi Laveran 1897(109)
40(35)	Shell-valves striated
41(44)	Spore definite in shape

Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named.

42(4	 Length of polar capsule more than one-fourth of that of spore; spore 18 to 20 long, 5 to 6 μ broad
	Myxidium lieberkühni Bütschli 1882(107)
43(4	2) Length of polar capsule less than one-seventh of that of spore; length 16 to 17μ ,
	breadth 5µ
	Myzidium mackiei Bosanquet 1910(112)
44(4	
	2.8µ. Vegetative form produces cyst
_	Myxidium anguillae Ishii 1915(114)
Inco	mpletely described species
	Myxidium sp. Gurley 1894(109)
	Myxidium sp. Awerinzew 1908(113)
	Myxidium sp. Mavor 1915(115)
	Genus SPHAEROMYXA Thélohan 1892
1(6) Spore straight, not arch-shaped
2(.	
3(4) Ends of spore truncate; striations longitudinal; length 15 to 20μ , breadth 5 to 6μ .
	Sphaeromyza balbianii Thélohan 1892(118)
4(3	3) Ends of spore rounded; sutural plane forming some angles with longitudinal axis
	of spore; striations transverse; length 12 to 14 μ , breadth 9 to 10 μ
	Sphaeromyxa immersa Lutz 1889
5(2	·
•	twice or three times larger than those of Sphaeromyza balbianii
	Sphaeromyxa gasterostei Georgévitch(121)
6()	
7(8	
• • •	4.3µ
	Sphaeromyxa sabrazesi Laveran et Mesnil 1900(120)
8(1	• •
9(10	
- (-	Sphaeromyza ezneri Awerinzew 1913(121)
10(9	
11(12	
,	Sphaeromyza incurvata Dollein 1898(119)
12(1	
(-	breadth and thickness 5.4µ
	Sphaeromyxa hellandi Auerbach 1909(121)
	Genus ZSCHOKKELLA Auerbach 1910
1(4	
2(3	•
-('	breadth 13 to 18µ
	Zschokkella hildae Auerbach 1910(122)
3(2	
	Zschokkella globulosa Davís 1917(123)
4(1	•
5(6	
	length 10 to 14 μ , breadth 6 to 7 μ
	Zschokkella acheilognathi Kudo 1916(123)
1	lumbers enclosed in parentheses refer to the page of the article on which is found the description of the species

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6(5)	Openings of polar capsules on side; polar capsules rounded pyriform; spore smaller; length 9.5 to 11.5 \(\mu\), breadth 6.5 to 7 \(\mu\) Zschokkella nova Klokacewa 1914(122)
	Genus MYXOSOMA Thélohan 1892
1(2)	
-(-,	capsule 6 to 7μ by 2 to 3μ . Cysts polymorphous
	Myxosoma dujardini Thélohan 1892(124
2(1)	
	length 14μ , breadth 8μ , thickness 6μ , polar capsule 8μ by 2μ . Cysts spherical
	up to 360µ in largest diameter Myxosoma funduli Kudo 1918(125)
Ambia	Myzosoma junaus Kudo 1910(125) lous form
vimorge	Myxosoma lobatum Nemeczek 1911(124,
	Genus LENTOSPORA Plehn 1905
1(8)	Spore circular in front view
2(3)	Vegetative form produces cysts. Spore: length and breadth 6.3 to 7μ , thick-
-(•,	ness 4.2 to 4.9μ
	Lentos pora dermatobia Ishii 1916(127)
3(2)	Vegetative form does not produce cysts or cysts unobserved
4(5)	Spore small; trophozoites found in the blood vessel of the brain. Spore 5 to 5.5μ
	in diameter.
5(4)	Lentospora encephalina Mulsow 1911
6(7)	Spore slightly pointed at anterior end; length 8 to 10μ , breadth 7 to 8μ , thickness
0(1)	5 to 6μ
	Lentospora acuta Fujita 1912(127)
7(6)	Anterior end of spore rounded; diameter 6 to 10μ
04.4	Lentospora cerebralis Plehn 1905(125
8(1)	Spore oval in front view.
9(10)	Spore symmetrically built; length 12 μ, breadth 9.5 μ, thickness 6 μ Lentos por a multiplicata Reuss 1906(126)
10(9)	Spore asymmetrically built; length 10 to 11μ , breadth 6.5 to 7μ
	Lentospora asymmetrica Parisi 1912(126
	Genus MYXOBOLUS Bütschli 1882
1/10)	
1(18) 2(9)	Spore with one polar capsule
3(6)	Breadth of spore equal to half of length
4(5)	Spore larger; often calabash-shaped; anterior end drawn out into a rounded tip;
` '	shell thickened at tip; length 15μ , breadth 7 to 8μ , thickness 5 to 6μ
	Myzobolus toyamai Kudo 1915(131)
5(4)	Spore smaller; anterior end pointed; shell of uniform thickness; length 9 to 10µ,
	breadth 4.5 to 5.5μ , thickness 3μ
6(3)	Myxobolus oculi-leucisci Trojan 1909(130)
7(8)	Breadth of spore more than two-thirds of length
. ()	Myxobolus unicapsulatus Gurley 1893(129
8(7)	Polar capsule long and median in position; spore broader; length 13.2 to 13.6 µ,
	breadth 10.1 to 10.3μ
	Myxobolus seni Southwell et Prashad 1918(132)
Nun named.	abers enclosed in parentheses refer to the page of the article on which is found the description of the specie
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190	ILLINOIS BIOLOGICAL MONOGRAPHS	[428
9(2)	Breadth of spore less than half of length	10
10(17)	Breadth of spore more than two-fifths of length	11
11(16)	Spore without any process.	12
12(15)	Shell of uniform thickness.	13
13(14)	Spore bent to one side; shell thickened at slightly rounded anterior tip; sutural	13
13(14)		
	edge without marking; length 16 to 18μ , breadth 7 to 8μ , polar capsule 5	
	to 7μ long	
	Myzobolus piriformis Thélohan 1892	129)
14(13)	Spore straight; shell thickened at posterior part; sutural edge with four to six	
	markings; length 18 to 20μ , breadth 8μ , thickness 6μ , polar capsule 9 to 10μ	
	long	
	Myxobolus fuhrmanni Auerbach 1909	130)
15(12)	Shell of uniform thickness; valves symmetrical; sutural edge with markings up to	
	12 in number; spore 14 to 15.5 μ long, 6 to 7.3 μ broad, 5 to 6 μ thick, polar cap-	
	sule 6.3μ by 2 to 3μ	
	Myxobolus misgurni Kudo 1919	133)
16(11)	Spore with a posterior process, 5μ in length and as broad as spore; length 17 to	•
	18μ , breadth 7.5 to 8μ , polar capsule 7μ by 4μ	
	Myxobolus notatus Mavor 1916	131)
17(10)	Breadth of spore about one-fourth of length; sporelarge; polar capsule extremely	,
11(10)	large; length 30 to 32μ , breadth 7 to 8μ , length of polar capsule 22 to 23μ	
	Myxobolus rohitae Southwell et Prashad 1918	1221
18(1)	Spore with two polar capsules.	
		19
19(24)	Form of spore variable	20
20(23)	Spore with an intercapsular appendix at anterior end	21
21(22)	Spore oval; length 10 to 12μ , breadth 8 to 9μ , thickness 6μ , polar capsule 5μ by 2	
	to 3μ	
	Myzobolus mülleri Bütschli 1882(128)
22(21)	Spore pyriform or elongated oval; length 11 to 16μ , breadth 8 to 13μ , polar cap-	
	sule 6μ by 4μ	
	Myxobolus cycloides Gurley 1893(1	140)
23(20)	Spore without intercapsular appendix; circular form 7 to 8μ , breadth 8 to 10μ ,	
	thickness 6μ , polar capsule 4 to 5μ by 2μ	
	Myxobolus hylae Johnston et Bancroft 1918(1	153)
24(19)	Form of spore definite	
25(28)	Polar capsules in each spore regularly of considerably different size	26
26(27)	Spore with an intercapsular appendix; anterior end rounded; sutural edge with	
	folds (3 to 5); length 10 to 12μ , breadth 8μ	
	Myzobolus dispar Thélohan 1895	135)
27(26)	Spore without intercapsular appendix; anterior end pointed; no fold on sutural	,
J. (J.)	edge	
	Myzobolus inaequalis Gurley 1893	1351
28(25)	Polar capsules approximately of equal form and size	
		47
29(30)	Sutural diameter smaller than breadth; length 6 to 7 μ , breadth 8 μ	1201
20/20\	Myzobolus transoralis Gurley 1893	
30(29)	Length equal to or more than breadth of spore	31
	Length longer than breadth	32
32(37)	Breadth of spore less than half of length	33
33(34)	Extremities of spore equally pointed; length 13 to 14.5 μ , breadth 6 to 7μ , polar	
	capsule 4.5µ long	
	Myxobolus miyairii Kudo 1919	155)
34(33)	Anterior end of spore attenuated; posterior end rounded	35
	ibers enclosed in parentheses refer to the page of the article on which is found the description of the ap	ecies
named.		

35(36)	Shell thickened at posterior margin; spore 12 to 15μ long, 4 to 6.8μ broad, polar capsule 5.5 to 7μ by 2.1 to 2.5μ
	Myxobolus anurus Cohn 1895(142)
36(35)	Shell of uniform thickness; spore 14.3 μ long, 6.7 μ broad, polar capsule 6.5 μ by 2 μ
30(33)	Myxobolus funduli Kudo 1919(151)
37(32)	Breadth of spore greater than half of length
38(41)	Length of spore greater than 20μ
39(40)	Spore large; subcircular and anterior end flattened in front view; sutural edge
07(10)	with markings; length 38 to 45μ , breadth 32 to 38μ , thickness 28 to 35μ , polar
	capsule 15 to $17\mu \log $
	Myxobolus magnus Awerinzew 1913(150)
40(39)	Spore small; extremities equally rounded; length 21μ , breadth 15μ , polar capsule
20(07)	$6\mu \log$
	Myxobolus cyprini Doflein 1898(143)
41(38)	Length of spore less than 20μ . 42
42(43)	Spore with a wide (2 to 3μ) membraneous posterior process; length 12μ , breadth
` .	10μ , length of polar capsule 4.5μ
	Myxobolus cordis Keysselitz 1908(148)
43(42)	Normal spore without appendage 44
44(49)	Breadth of sutural ridge one-third of thickness of spore
45(46)	Length of spore smaller than 10μ ; subcircular in front view; length 7 to 8μ ,
	breadth 6 to 7μ , thickness 5μ
	Myzobolus globosus Gurley 1893(139)
46(45)	Length of spore greater than 10μ
47(48)	Spore large; elliptical in front view; with an intercapsular appendix, sutural edge
	with markings; length 16.9 to 21.6 μ , breadth 13 to 16.2 μ , thickness 9 μ
	Myxobolus gigas Auerbach 1906(145)
48(47)	Spore small; subcircular in front view; markings on entire sutural edge;
	length 10.8 to 11.7 μ , breadth 9.9 to 10.4 μ , thickness 7.2 to 9 μ
40/44\	Myzobolus aeglefini Auerbach 1906(144)
49(44) 50(69)	Sutural ridge narrower
51(68)	Spore with an intercapsular appendix
52(57)	Intercapsular appendix triangular
53(54)	Sutural edge without marking; length 11 to 12μ , breadth 9.25 to 10.5μ
33(3 1)	Myzobolus balleri Reuss 1906(147)
54(53)	Sutural edge with markings. 55
55(56)	Spore small; length 8 to 9.5 μ , breadth 6 to 7.5 μ , thickness 5.5 μ
00(00)	Myxobolus exiguus Thélohan 1895
56 (55)	Spore large; often subcircular; length 11.5μ to 12μ , breadth 7.5 to 8μ , thickness
	5μ
	Myxobolus obesus Gurley 1893(140)
57(52)	Anterior end of spore broadly rounded in front view
58(59)	Posterior portion of spore narrower; polar capsule rather large; length 11.4 to
	13.5 μ , breadth 9.5 to 11 μ , thickness 8.5 to 9.5 μ , polar capsule 5.5 to 6 μ long
	Myxobolus discrepans Kudo 1919(156)
59(58)	Extremities of spore approximately equal
60(61)	Sutural edge without markings; spore 10 to 10.5 µ long, 8 to 8.5 µ broad, polar
	capsule 4.5 µ long
	Myxobolus squamae Keysselitz 1908(147)
61(60)	Sutural edge with markings
62(65)	Markings distinct
	mbers enclosed in parentheses refer to the page of the article on which is found the description of the species
named.	

63(64)	Marking variable in number along posterior margin of spore; spore more elongated; length 12 to 12.5μ, breadth 10 to 10.5μ, polar capsule 5.5 to 6μ long Myxobolus pfeifferi Thélohan 1895
64(63)	Sutural edge with four markings around posterior margin; spore rather short;
01(00)	length 11 to 12μ , breadth 9 to 9.5μ , thickness 4.5 to 5μ , polar capsule 5μ by 2.5μ
	Myxobolus scardinii Reuss 1906(146)
65(62)	Markings indistinct
66(67)	Markings about five at posterior margin; spore larger and shorter; length 11 to
00(01)	12 μ , breadth 9.25 to 10 μ , thickness 4.5 to 5.5 μ , polar capsule 4 to 5 μ by 2.25 μ
	Myxobolus bramae Reuss 1906(147)
67(66)	Markings many along entire sutural edge except anterior tip; spore smaller,
	longer and thicker; length 9.25 to 10μ , breadth 7 to 7.25 μ , thickness 5 to 5.5 μ ,
	polar capsule 4.5μ by 2.5 to 3μ
	Myxobolus cyprinicola Reuss 1906(147)
68 (51)	Intercapsular appendix rounded; sutural edge smooth; length 11μ , breadth 8μ ,
	polar capsule 4 to 6μ long
	Myxobolus musculi Keysselitz 1908(148)
69(50)	Spore without intercapsular appendix
70(75)	Length of spore less than 10μ
71(72)	Spore very much flattened and small; length 6μ , breadth 4.2 to 5μ , polar capsule
	3μ by 2μ
	Myxobolus minutus Nemeczek 1911(150)
72(71)	Thickness of spore about half of length
73(74)	Shell thick; length 9.25 to 10μ , breadth 7.25 to 8.25μ , thickness 4 to 5μ
	Myxobolus sandrae Reuss 1906(146)
74(73)	Shell thin; spore 8.25 to 9.5 μ long, 7.25 to 8.25 μ broad, 4.5 to 5.5 μ thick
	Myxobolus volgensis Reuss 1906(145)
75(70)	Length of spore greater than 10μ
76(85)	Extremities of spore approximately equal
77(80)	Spore elongated (breadth: length=1:1.8 or 1:1.4)
78(79)	Spore larger; length 14 to 17μ , breadth 8.5μ , thickness 5 to 6μ
	Myxobolus oblongus Gurley 1893(139)
79(78)	Spore smaller; length 12 to 15 μ , breadth 9 to 11 μ
	Myxobolus ellipsoides Thélohan 1892(136)
80(77)	Spore shorter (breadth: length=1:1.3, 1:1.2 or 1:1.1)
81(82)	Sutural edge with markings; slightly truncate at anterior end in front view; spore
	9.5 to 11.5 μ long, 8.5 to 9.5 μ broad, 6.5 μ thick, polar capsule 4.75 μ by 1.5
	to 2μ
	Myxobolus mesentericus Kudo 1919(157)
82(81)	Sutural edge without markings
83(84)	Polar capsule larger; spore 13.9 µ long, 11 µ broad, 8 µ thick
	Myzobolus lintoni Gurley 1893(138)
84(83)	Polar capsules smaller; spore 14.5μ long, 11.9μ broad, polar capsule 6μ by 3.7μ
	Myxobolus pleuronectidae Hahn 1917(152)
85(76)	Anterior end of spore more attenuated than posterior
86(89)	Sutural edge with markings 87
87(88)	Markings five or six in number; spore 17 to 18µ long, 10 to 13µ broad, polar cap-
	sule 7 to 8μ
	Myxobolus permagnus Wegener 1910(149)
88(87)	Markings sometimes present; spore 13 to 17μ long, 8 to 10μ broad, 5 to 7μ thick,
	polar capsule 6 to 7μ long
	Myxobolus carassii Klokacewa 1914(150)
37	

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3(6) 4(5)	Length of tail equal to two-thirds of length of main part of spore*	4
	Henneguya gasterostei Parisi 1912	
5(4)		,
	Henneguya mictospera Kudo 1919(173)
6(3)	20 to 24μ, breadth 5 to 6μ, length of polar capsule 4 to 5μ	
7(0)	Henneguya media Thélohan 1892	•
7(2) 8(9)	Shell-valves unstriated	8
O(9)	wider and bifurcated to same direction; total length 19.5 to 22.5 μ , length of main part 8.5 μ , breadth 6 μ , length of tail 8 to 8.5 μ	
	Henneguya légeri Cépède 1905(166)
9(8)	Spore oval; polar capsule shorter; posterior part of main portion narrow; tail	•
	narrower and bifurcated to opposite directions; length of main part 11.5 µ,	
	breadth 7μ , length of tail 9.6μ , polar capsule 3.5μ by 2.5μ	
	Henneguya wisconsinensis Mavor et Strasser 1916(170	•
10(1)	Parasitic in tissue of host	_
11(28)	Tail always appears as a single process	Z
12(13)	Spore small; length 4\mu, breadth 2\mu Henneguya tenuis Vaney et Conte 1901(166	
13(12)	Spore longer and larger, at least 27 µ long.	
14(15)	Sutural edge with eight to ten markings; tail rather long; length of main part 10	7
11(30)	to 11.5 μ, breadth 8 to 8.75 μ, thickness 4 to 5 μ, polar capsule 3 to 4 μ by 2 μ, tail up to 17 μ long	
	Henneguya brachyura Ward 1919(171)
15(14)	Sutural edge without markings	٠.
16(19)	Total length of spore greater than 40μ	7
17(18)	Anterior end rounded; polar capsule large; shell-valves asymmetrical; tail long; main part 10 to 11 \mu long, 6 to 8 \mu broad, 4 \mu thick, tail 30 to 40 \mu long. Cysts	
	elongated and large up to 6 mm. by 2 mm.	
	Henneguye macrura Gurley 1894(164)
18(17)		
	shorter; length 20μ , breadth 8 to 9μ , polar capsule 8μ by 2 to 3μ . Cysts elongated oval and smaller, 1.1 mm. by 0.5 mm.	
	Honneguya creplini Gurley 1894(162	J
19(16)	Total length of spore less than 40μ	0
20(21)	Tail about one-third of main part; total length 38μ , main part 26μ leng, 10 to 11μ broad, 8μ thick, polar capsule 11 to 14μ by 2 to 3μ . Cysts eval, numerous and	
	small (130 µ by 115 µ) Henneguya minuta Cohn 1895(160)	1
21(20)	Tail about three-sevenths of main part; total length 29 to 40 \mu, main part 15 to	•
()	20μ long, 7 to 8μ broad, polar capsule 8μ by 2μ	2
22(25)	Cysts large	3
	ength of main part of spore denotes in all possible cases the distance between the oute	1
	tip and the posterior margin of sporal cavity; and consequently that between the	
	nd the distal end of the tail is the length of the tail.	

Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named. \P



23(24)	Cysts spherical up to 6 mm. in diameter; parasitic in ovary
	Henneguya oviperda (Cohn 1895)(160
24(23)	Cysts elongated up to 2 mm. by 1.5 mm.; parasitic in branchia Henneguya psorospermica Thélohan 1895
25/22)	
25(22)	Cysts rather small or size not observed
26(27)	Elongated cysts 0.75 mm. by 0.375 mm.; parasitic in branchia
	Henneguya texta (Cohn 1895)
27(26)	Parasitic in intestinal wall
	Henneguya peri-intestinalis Cépède 1906(161
28(11)	Tail composed of two processes
29(30)	Total length reaches 87.5 to 110.5 μ ; length of main part 10.5 to 15 μ , breadth
	5μ , length of polar capsule 5μ , length of tail 75 to 100μ
	Henneguya gigantea Nemeczek 1911(168)
30(29)	Total length of spore less than 82μ
31(34)	Sutural edge with markings
32(33)	Tail longer and spore larger; anterior end more rounded; total length 47μ . Cysts
	spherical and large, up to 6 mm.
	Henneguya salminicola Ward 1919(171)
33(32)	Tail shorter and spore smaller; anterior end slightly more attenuated; total
	length 32µ. Cysts lenticular, up to 2 mm. in length
	Henneguya nüsslini Schuberg et Schröder 1905(166)
34(31)	Sutural edge without markings
35(38)	Distal end of tail thread-like
36(37)	Tail 40 to 50μ in length; total length 50 to 60μ , main part 8.5 to 9.5 μ long, 8.5 to
, ,	9.5 μ broad, polar capsule 4.7 to 5.5 μ by 3 μ . Cysts small, up to 50 μ in
	diameter
	Homneguya noapolitana Parisi 1912(170)
37(36)	Tail 10 to 30 m in length; main part 12 m long, 8 m broad
(,	Henneguya miyairii Kudo 1919
38(35)	Distal end of tail tapers to a point and not thread-like
39(40)	Cysts irregular in shape; size up to 2.5 mm. Spore: total length 30 to 40μ ,
(,	main part 11.5 to 15µ long, 5 to 6.5µ broad, polar capsule 6.5 to 8µ by 2 to
	2.5μ , tail 22 to 28μ
	Henneguya lobosa (Cohn 1895)(161)
40(39)	Cysts spherical or oval. 41
41(42)	Anterior end of spore rounded; tail either single or double processes; total length
(/	55μ , length of main part 10μ , breadth 7μ , length of tail 40 to 50μ . Cysts
	spherical or oval up to 32 mm. by 16 mm.
	Henneguya zschokkei (Gurley 1893)(165)
42(41)	Anterior end attenuated; spore large; main part 20 to 22µ long, breadth 8 to
(,	9μ , 6 to 7μ thick, polar capsule 10μ by 2 to 3μ , tail 50 to 60μ long
	Henneguya acerinae Schröder 1906(167)
Income	etely described species
	Henneguya schizura (Gurley 1893)(162)
	Henneguya linearis (Gurley 1893)(163)
	Henneguya gurleyi Kudo 1893(163)
	Henneguya monusa (Gurley 1893)(164)
	Henneguya monura (Gurley 1893)
	Henneguya sirongyiura (Gurley 1894)
	Henneguya brevis Thélohan 1895(162)
	Henneguya sp. (Gurley 1894)
	Henneguya sp. (Gurley 1894)
	Henneguya sp. Nemeczek 1911(169)

Numbers enclosed in parentheses refer to the page of the article on which is found the description of the species named.

SUMMARY

- 1) All species of Myxosporidia which have been observed in various parts of the world, reaching 237 in number, are recorded with figures.
- 2) A new classification of Myxosporidia is proposed after discussion of those of previous authors.
- 3) A complete list of the specific names of the hosts that harbor Myxosporidia, is given together with the names of the organ of infection and the localities from which recorded.
- 4) By study of the geographical distribution of Myxosporidia, it is shown that few species are common both to American and European waters or Asiatic and European waters, while the majority of Myxosporidia are localized in definite and limited regions.
- 5) The study of the organal distribution of Myxosporidia in the host, shows that the gall-bladder is the organ most frequently invaded by the parasite. The kidney, branchia and urinary bladder have less chances of being attacked.
 - 6) One new genus, Wardia, is established.
- 7) Nine new species; Wardia ovinocua, Mitraspora elongata, Chloromyxum trijugum, Chloromyxum catostomi, Myxidium americanum, Myxobolus orbiculatus, Myxobolus discrepans, Myxobolus mesentericus and Henneguya mictospora, are described from fresh-water fish collected in the vicinity of Urbana, Ill.
- 8) Six new species; Sphaerospora carassii, Myxidium kagayamai, Myxobolus misgurni, Myxobolus miyairii, Myxobolus koi and Henneguya miyairii, are recorded from fresh-water fish of Nippon.
- 9) One new species; Chloromyxum wardi, is described from Alaska. This is the second species of Myxosporidia from that part of North America.
- 10) Keys to the genera and species of known Myxosporidia are included.



APPENDIX: NEW MYXOSPORIDIA FROM AUSTRALIA

The following six species described by Johnston and Bancroft did not reach the writer until the page proof was read. For this reason they could not be put in the text and are recorded here separately.

MYXIDIUM THERAPON Johnston et Bancroft

1919 Myxidium therapon Johnston and Bancroft 1919: 520-521

Habitat: In the gall bladder of *Therapon carbo* and *Th. hillii*; Thomson River at Longreach, Australia.

The parasite occurred in one specimen of the former host fish and in nine out of thirteen specimens of the latter. No visible effect of the infection on the part of the host fish was recognized.

Vegetative form: Body pale yellowish to green in color. Form(?). Size varies from 3 to 12mm. in diameter. The protoplasm is differentiated into a clear narrow ectoplasm, about 10μ in width, and a coarsely grained endoplasm. No movements could be seen on slides; but undulations were observed to travel round the margin of the trophozoite. Polysporous.

Spore: Spindle-shaped with slightly pointed extremities. Polar capsules are more or less rounded. Shell with faintly marked longitudinal striation. The sporoplasm is binucleated. Average dimensions: length 9 to 10μ , breadth 4μ , polar capsules 2 to 3μ by 1 to 2μ .

MYXOSOMA OGILBYI Johnston et Bancroft

1919 Myxosoma ogilbyi Johnston and Bancroft 1919: 521-522

Habitat: In the fibrous tissue of the gill arch of *Plectroplites ambiguus;* Thomson River at Longreach, Australia. Three out of nine host specimens examined showed the infection.

Vegetative form: The parasite forms white cysts usually close to the bases of the gill filaments. Cysts are small and rounded, being less than 1mm. in diameter. The authors simply mention that sections revealed the structure usually present in a Myxosporidian cyst.

Spore: Oval with pointed anterior end. The inner margin of the shell is indented posteriad. The sporoplasm contains a single nucleus, but not any iodinophilous vacuole. Average dimensions: length 11 to 13μ , breadth 6 to 8μ , thickness 5μ , polar capsules 5 to 6μ by 2μ .

MYXOBOLUS PLECTROPLITES Johnston et Bancroft

1919 Myxobolus plectroplites Johnston and Bancroft 1919: 522-523

Habitat: In the kidney and gall-bladder of *Plectroplites ambiguus*; Thomson River at Longreach, Australia. The parasite was observed in

four out of nine host fish examined; in two cases in the kidney only, in one case only in the gall-bladder, and in one instance in both gall-bladder and kidney. Cysts were found in the kidney, while only spores were recognized in the gall-bladder.

Vegetative form: The cysts which could only be detected in sections, lie in the connective tissues of kidney. They are of minute size, ranging somewhat widely from 36μ in diameter to 144 by 100μ . According to the authors no definite structure could be found.

Spore: Rounded oval. It bears quite a close resemblance to that of *Myxobolus kylae* (page 153), which is slightly longer, and which has a longer polar filament than the present form. The vacuole, however, is apparently not iodinophilous(?). Average dimensions: length 10 to 12μ , breadth 7 to 8μ , polar capsules 5 by 2μ , length of polar filament 30 to 40μ .

HENNEGUYA AUSTRALIS Johnston et Bancroft

1919 Henneguya australis Johnston and Bancroft 1919: 523-524

Habitat: In the branchiae of *Plectroplites ambiguus*; Thomson River at Longreach, Australia. The parasite was detected in four out of nine host fish examined. The infection was extremely light in all cases.

Vegetative form: The parasites form cysts. They lie embedded in the spongy mass of the gill filament, and in many cases occupy a relatively large area of the section. Cysts showed two layers in section; the outermost clear ectoplasm and inner endoplasm with developing spores, the central portion of which being filled with mature spores. The spores appear to lie in a definite manner, the long axis of the spore commonly being at right angles to the boundary of the cyst, the anterior end of the spore pointing outwards.

Spore: Elongated ovoidal. Anterior end pointed, posterior end drawn out into a tail. The tail appears single when the spore is removed from the cyst but separates soon afterward into two halves which usually diverge widely. Two polar capsules parallel to each other are quite frequently of different length. The sporoplasm contains two nuclei and a small vacuole (iodinophilous?). Average dimensions: length 11 to 15μ , breadth 3 to 5μ , thickness 3 to 4μ , polar capsules 5 to 6μ by 1 to 2μ , length of tail about 20μ .

HENNEGUYA GRACILIS Johnston et Bancroft

1919 Henneguya gracilis Johnston and Bancroft 1919: 524-526

Habitat: In the gill filament of *Therapon hillii*; Thomson River at Longreach, Australia. Out of thirteen specimens examined, eight were infected. Heavy infection was recognized only in one case.

Vegetative form: The cyst is of definite, narrow, pear-shaped form, and lie transversely, i.e., at right angles to the long axis of the gill filament.



Spore: The spore resembles *Henneguya australis*, but is slightly smaller, while the tail is longer in proportion. The spores are arranged with long axis parallel to that of the cyst. Average dimensions: length 10 to 14μ , breadth 2.5 to 3μ , thickness 3μ , polar capsules 5 to 6μ by 1 to 2μ , length of tail about 20 to 26μ .

HENNEGUYA sp. Johnston et Bancroft

1919 Henneguya sp.

Johnston and Bancroft

1919 : 526

Habitat: In the branchiae of Nematalosa elongata; Thomson River at Longreach, Australia.

The authors state that they observed a number of spores of a Henneguya in the scrapings of the gill of one of four host fish.

Vegetative form: Undescribed.

Spore: Undescribed.

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GENERAL EXPLANATION OF FIGURES

For the type species of each genus, both the vegetative form and the spore are illustrated. For the other species, except those which are new, figures of the spore are given, unless the vegetative forms are different from those of the type species or the species were reported in papers which seem to be of less universal distribution.

The original drawings were made with the Abbe drawing apparatus. The combinations used were Zeiss apochromatic objectives 16, 8, 3, and homogeneous oil immersion 2 mm. with compensation oculars 2, 4, 6, 8, 12 and 18. All the other drawings were copied from the original figures of the respective observers, an exact citation of which is given in each case, and were also made with the same drawing apparatus on the same scale except that a few figures were enlarged among those that were taken from other authors.

Magnifications were also calculated and given for those quoted figures, for which the respective authors failed to mention the scale at which the drawings were made.



PLATE I

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- Figs. 1 to 5. Leptotheca agilis.
- Fig. 1. A typical trophozoite in vivo. After Thélohan (1895, Fig. 29); ×750.
- Fig. 2. A young form. After Thélohan (1895, Fig. 31). ×750.
- Fig. 3. A trophozoite in motion. After Doflein (1898, Fig. 5).
- Fig. 4. A trophozoite in contracted condition. After Doflein (1898, Fig. 7).
- Fig. 5. A fresh spore. After Thélohan (1895, Fig. 30). ×1500.
- Fig. 6. A fresh mature spore of Leptotheca elongata. After Thélohan (1895, Fig. 38). ×1500.
- Fig. 7. A fresh mature spore of Leptotheca pares. After Thélohan (1895, Fig. 25). ×1500.
- Fig. 8. A fresh spore of Leptotheca perlata. After Balbiani (1884, Fig. 40).
- Fig. 9. A spore of Leptotheca macrospora. After Auerbach (1909, Fig. 2a). ×1350.
- Fig. 10. A spore of Leptotheca informis, preserved in formol. After Auerbach (1910b, Fig. 1a). × about 2000.
- Fig. 11. A spore of Leptotheca longipes, preserved in formol. After Auerbach (1910b, Fig. 1d). × about 2200.
- Fig. 12. A fresh spore of Leptotheca fusiformis. After Davis (1917, Fig. 1). ×1500.
- Fig. 13. Afresh spore of Leptotheca scisswa. After Davis (1917, Fig. 8). ×1500.
- Fig. 14. A fresh spore of Leptotheca lobosa. After Davis (1917, Fig. 11). ×1500.
- Fig. 15. A fresh spore of Leptotheca glomerosa. After Davis (1917, Fig. 13). ×1500.
- Fig. 16. A trophozoite of Leptotheca sp. After Awerinzew (1908, Pl. 2, Fig. 14). 1/12 and comp. oc. 12.
- Fig. 17. Another trophozoite of the same. After Awerinzew (1908, Pl. 2, Fig. 17). 1/12 and comp. oc. 12.
- Figs. 18 to 20. Ceratomyza arcusta. After Thélohan (1895).
- Figs. 18 and 19. Typical young form. After Thélohan (1895, Figs. 16 and 17).
- Fig. 20. A trophozoite with two spores. After Thelohan (1895, Fig. 18).

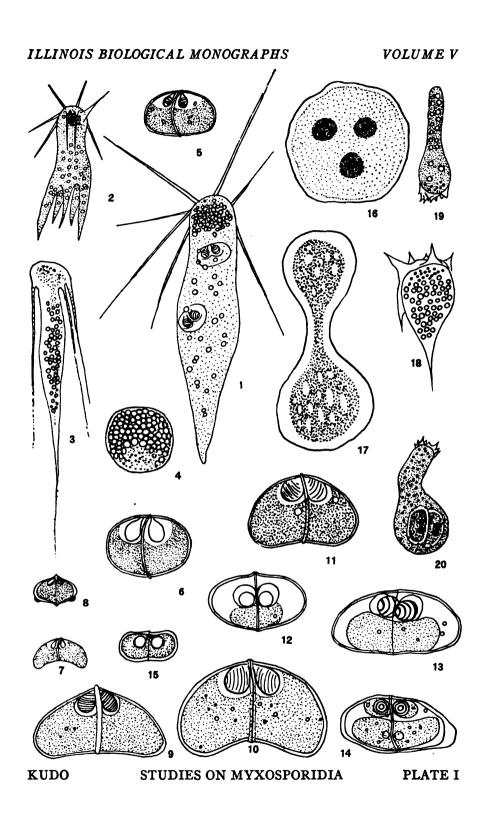


PLATE II

- Fig. 21. A sporulating trophozoite of Caratomyses arcusts. After Parisi (1912, Fig. 6a).
- Fig. 22. A spore of Caratomyza arcusta treated with KOH. After Thélohan (1895, Fig. 19). ×1500.
- Figs. 23 and 24. Ceratomyza sphaerulosa. After Thélohan.
- Fig. 23. A part of the tropzoite (1895, Fig. 2). ×750.
- Fig. 24. A fresh spore (1895, Fig. 3). ×750.
- Fig. 25. A fresh spore of Ceratomyza globulifera. After Thélohan (1895, Fig. 43). ×1500.
- Fig. 26. An adult trophozoite of Ceratomyza appendiculata. After Thélohan (1895, Fig. 4).

 × about 400.
- Fig. 27. A spore of Ceratomyza truncata. After Thélohan (1895, Fig. 51). ×1500.
- Fig. 28. A spore of Ceratomyza reticularis. After Thélohan (1895, Fig. 27). ×1500.
- Fig. 29. A spore of Ceratomyza inaequalis. After Doflein (1898, Fig. 10). ×1250.
- Figs. 30 and 31. Ceratomyza linospora. After Doflein (1898). X about 1900.
- Fig. 30. A spore (1898, Fig. 39).
- Fig. 31. A trophozoite with spores (1898, Fig. 43).
- Figs. 32 and 33. Ceratomyza ramosa. After Awerinzew (1908).
- Fig. 32. A trophozoite (1908, Pl. 2, Fig. 20). Zeiss obj. D and comp. oc. 4.
- Fig. 33. A spore (1908, Pl. 2, Fig. 19). Zeiss obj. E and comp. oc. 4.

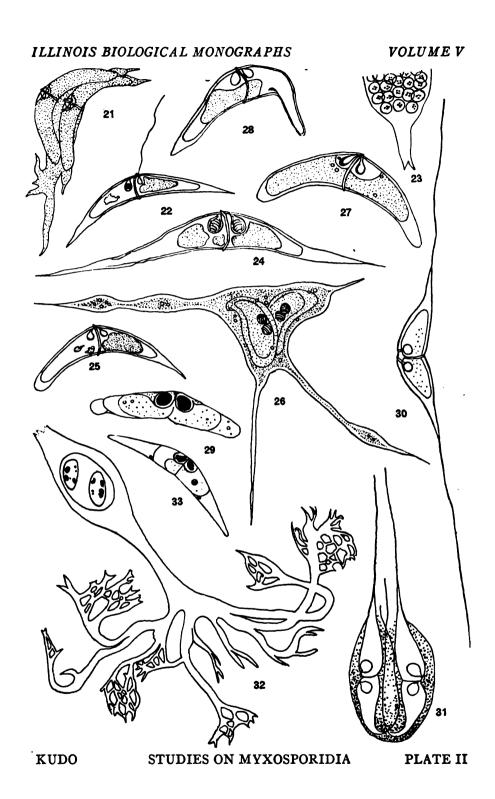


PLATE III

Figs. 34 to 39. Ceratomyza drepanopsettae. Awerinzew (1908).

Fig. 34 and 35. Trophozoites (1908, Pl. 2, Figs. 7 and 9). Obj. D and oc. 4.

Fig. 36. The part of a trophozoite attached to the epithelium of the gall-bladder of the host 1908, Pl. 2, Fig. 10). Obj. E and oc. 4.

Figs. 37 to 39. Spores (1908, Pl. 1, Figs. 2, 3 and 1). Obj. D and oc. 4.

Figs. 40 and 41. Two different views of the spore of Coratomyza tylosuri. After Awerinsew (1913a, Fig. 1). × about 350.

Figs. 42 and 43. Ceratomyza(?) spari. After Awerinzew (1913a, Fig. 2).

Fig. 42. A trophozoite.

Fig. 43. A spore. × about 345.

Figs. 44 to 47. Spores of Ceralomyza acadiensis. After Mavor (1916). Figs. 44 and 45 (1916, Fig. B). ×270. Fig. 46 (1916, Fig. A) ×1800. Fig. 47 (1916, Fig. 40)×2950.

Fig. 48. A spore of Caratomyza coris. After Georgévitch (1916a, Fig. 1).

Fig. 49. A spore of Ceratomyza herowardi. After Georgévitch (1917, Fig. 1).

Fig. 50. A spore of Ceratomyza mesospora. After Davis (1917, Fig. 15). ×1500.

Fig. 51. A spore of Ceratomyza sphairophora. After Davis (1917, Fig. 23). ×950.

Figs. 52 and 53. Spores of Ceratomyza taenia. After Davis (1917, Figs. 26 and 25). ×700.

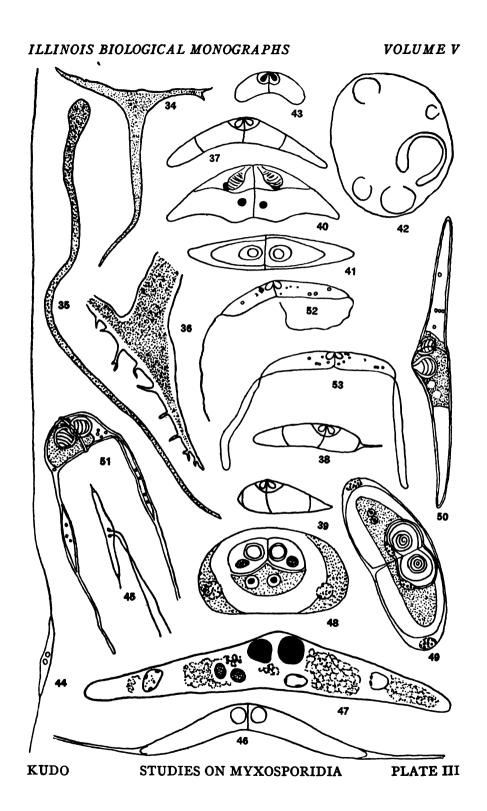


PLATE IV

- Fig. 54. A spore of Ceratomyza attenuata. After Davis (1917, Fig. 28). ×950.
- Figs. 55 and 56. Spores of Ceratomyza recurreta. After Davis (1917, Figs. 32 and 33). ×1500.
- Figs. 57 to 60. Spores of Ceratomyza lunata. After Davis (1917, Figs. 34 to 37). ×1500. Fig. 61. A spore of Ceratomyza abbreviata. After Davis (1917, Fig. 41). ×1500.
- Fig. 62. A spore of Ceratomyza flagellifera. After Davis (1917, Fig. 42). ×1500.
- Fig. 63. A spore of Ceratomyza agglomerata. After Davis (1917, Fig. 45). ×1500.
- Fig. 64. A spore of Ceratomyza amorpha. After Davis (1917, Fig. 47). ×1500.
- Figs. 65 to 67. Spores of Ceratomyza monospora. After Davis (1917, Figs. 57, 56 and 55). ×1500.
- Figs. 68 and 69. Spores of Ceralomyza streptospora. After Davis (1917, Figs. 59 and 60). ×1500.
- Fig. 70. A spore of Ceratomyza aggregata. After Davis (1917, Fig. 63). ×1400.
- Fig. 71. A spore of Ceratomyza undulata. After Davis (1917, Fig. 66). ×1500.
- Figs. 72 and 73. Spores of Ceratomyza navicularia. After Davis (1917, Figs. 69 and 68). ×1500.
- Fig. 74. A spore of Ceratomyza spinosa. After Davis (1917, Fig. 72). ×1500.

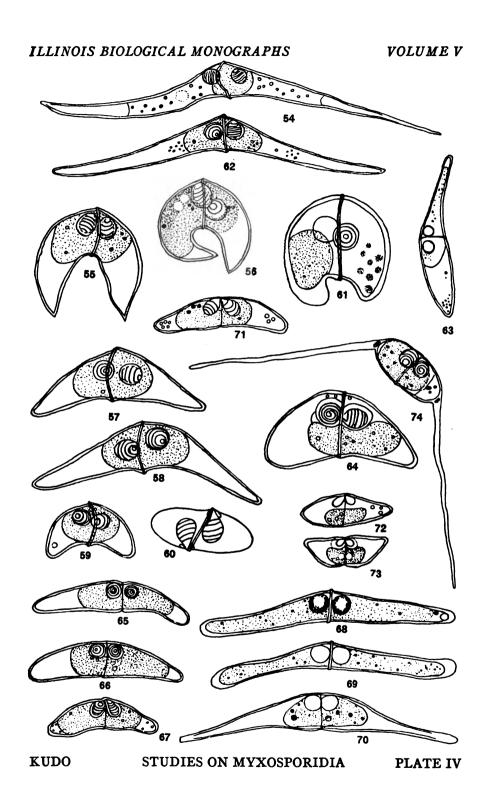


PLATE V

Figs. 75 to 80. Myzoproleus ambiguus. After Doflein (1898).

Figs. 75 and 76. Trophozoites of typical forms (1898, Figs. 12 and 52).

Figs. 77 and 78. Young trophozoites produced by budding (1898, Figs. 55 and 56). Figs. 79 and 80. Spores (1898, Figs. 54 and 64). × about 800 and 1080.

Figs. 81 to 83. Spores of Myzoproteus cordiformis. After Davis (1917, Figs. 79, 80 and 78). ×1500.

Fig. 84. A spore of Myzoproteus cornutus. After Davis (1917, Fig. 85). ×1400.

Figs. 85 to 95. Wardia ovinocua. Original.

Fig. 85. A portion of the cross-section thru an infected overy of Lepomis humilis, showing the parasite in one ovum and the hypertrophied nurse cells and several connective tissue layers. \times 160.

Fig. 86. A portion of the cross-section of a cyst. ×640.

Figs. 87 to 89. Three different views of fresh spore. ×2000.

Figs. 90 and 91. Stained spores. ×1700.

Fig. 92. A spore mechanically pressed and stained with Giemsa's mixature, showing the escaping polar capsules without extruding polar filament, and the sporoplasm. ×1700.

Figs. 93 to 95. Front and lateral views of the shell valves, exhibiting the network-like fine ridges on the surface and the posterior processes. ×1700.

Figs. 96 and 97. Spores of Wardia ohlmacheri. After Ohlmacher (1893, Figs. 4a and 2). 2mm. and oc. 4.

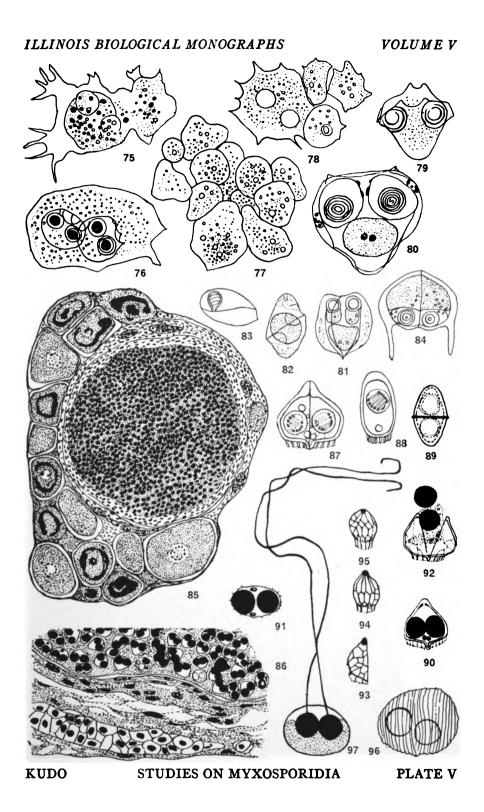


PLATE VI

Figs. 98 to 104. Mitraspora cyprini.

Figs. 98 and 99. Trophozoites from the ureter of Cyprinus carpio in vivo. Original. X about 1500.

Figs. 100 and 101. Different views of fresh spores. Original. X about 2000.

Figs. 102 to 104. Spores. After Fujita (1912, Figs. 1a to 1c).

Figs. 105 to 107. Mitraspora candata. After Parisi (1910 and 1913).

Fig. 105. A trophozoite (1910, Fig. 1).

Figs. 106 and 107. Front and lateral views of the spore (1913, Fig. 20 and 1910, Fig. 2). × about 1600.

Figs. 108 to 113. Chloromyxum leydigi.

Figs. 108 and 109. Trophozoites. After Thélohan (1895, Figs. 7 and 6). ×750.

Figs. 110 and 111. Trophozoites in division. After Doflein (1898, Figs. 57 and 58).

Figs. 112 and 113. Different views of spores. After Thélohan (1895, Figs. 10 and 9). ×1500.

Fig. 114. A spore of Chloromyxum caudatum. After Thélohan (1895, Fig. 36). ×1500. Figs. 115 and 116. Different views of the spore of Chloromyxum quadratum. After Thélohan (1895, Figs. 100a and 100b). ×1500.

Fig. 117. A spore of Chloromyzum quadratum treated with nitric acid. After Thélohan (1895, Fig. 100c). ×1500.

Fig. 118. A spore of Chloromyzum fluviatile. After Thélohan (1892, Fig. 2). ×1500.

Figs. 119 and 120. Different views of the spore of Chloromyzum mucronatum. After Lieber-kühn from Gurley (1894, Pl. 39, Fig. 5). × about 1750.

Figs. 121 and 122. The same after Balbiani (1884, Fig. 41). × about 1200.

Figs. 123 and 125. Spores of Chloromyxum diploxys. After Balbiani from Gurley (1894 Pl. 42, Figs. 13a, 13b and 13c).

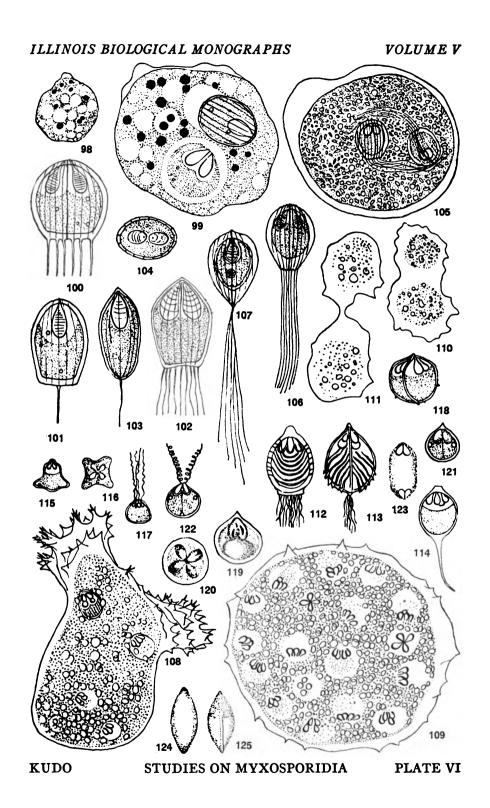


PLATE VII

Fig. 126. Trophozoite of Chloromyxum truttae. After Léger (1906, Fig. 4). ×1000.

Figs. 127 and 128. Trophozoites of Chloromyzum cristatum. After Léger (1906, Fig. 7). ×1000.

Figs. 129 to 133. Chloromyzum dubium. After Auerbach (1908).

Figs. 129 and 130. Trophozoites (1908, Figs. 2 and 1).

Figs. 131 and 132. Spores (1908, Figs. 3 and 4). × about 2250.

Fig. 133. A stained young spore (1908, Fig. 5).

Fig. 134. Trophozoite of Chloromyxum sp. After Awerinzew (1908, Pl. 2: Fig. 12). Objective D and ocular 4.

Fig. 135. Chloromyxum koi. After Fujita (1913). X about 800.

Figs. 136 to 138. Chloromyzum magnum. After Awerinzew (1913a, Fig. 4).

Figs. 136 and 137. Trophozoites.

Fig. 138. A spore. × about 320.

Figs. 139 and 140. Spores of Chloromysum funduli. After Hahn (1915, Figs. 34 and 33). ×2000.

Figs. 141 to 146. Chloromyzum misgurni. After Kudo (1916). ×1750.

Figs. 141 and 142. Trophozoites (1916, Figs. 3f and 3g)

Figs. 143 to 145. Different views of fresh spore (1916, Figs. 3a, 3c and 3b).

Fig. 146. A spore treated with potassium hydrate (1916, Fig. 3e).

Figs. 147 to 152. Chloromyxum fujitai. After Kudo (1916). ×1750.

Fig. 147. A trophozoite (1916, Fig. 4a).

Fig. 148. A fresh spore (1916, Fig. 4e).

Fig. 149. A spore stained with Giemsa's mixture (1916, Fig. 4g).

Figs. 150 to 152. Two surface views and an optical cross-section of a stained spore, showing the ridges on the shell valves (1916, Figs. 4b, 4d and 4c).

Figs. 153 to 156. Spores of Chloromyzum clupeidae.

Figs. 153 to 155. Fresh spores. After Linton (1901, Fig. 3). "Variously magnified."

Fig. 156. A spore. After Hahn (1917b, Fig. 10). ×1650.

Figs. 157 and 158. Two views of Chloromyzum granulosum. After Davis (1917, Figs. 137 and 138). ×1500.

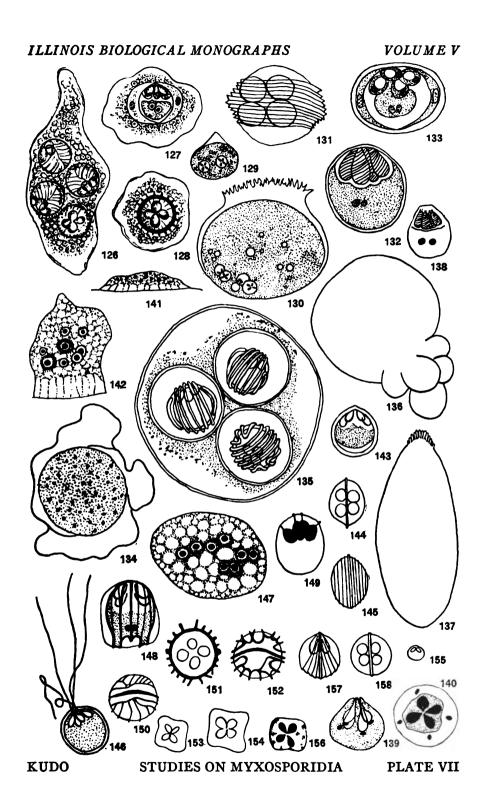


PLATE VIII

Figs. 159 to 182. Chloromyzum trijugum. Original. Figs. 159 to 174. Trophozoites of various form and age.

Figs. 159 and 160. Trophozoites of medium size. ×640.

Fig. 161. A trophozoite, ten minutes after it was removed from the host. ×1700.

Figs. 162 and 163. The movements of pseudopodia of the same specimen in ten minutes. ×1700.

Fig. 164. The same specimen after thirty minutes. ×1700.

Figs. 165 to 167. A trophozoite, showing the change of pseudopodus in five and ten minutes. ×1700.

Figs. 168 to 172. Small trophozoite with different numbers of the nuclei. Fig. 172 is probably a disporous form. ×2350.

Fig. 173. A monosporous trophozoite with a young spore. ×2350.

Fig. 174. A young spore. ×2350.

Figs. 175 to 177. Different views of fresh spores. ×1700.

Fig. 178. A Giemsa stained spore. ×1700.

Figs 179 and 180. Side views of the valves showing the ridges by Giemsa staining. ×1700.

Fig. 181. A spore treated with potassium hydrate solution, and stained with Giemsa solution. $\times 1700$.

Fig. 182. A spore from which the sporoplasm is leaving the shell. From the Giemsa smear of the infected bile. $\times 1700$.

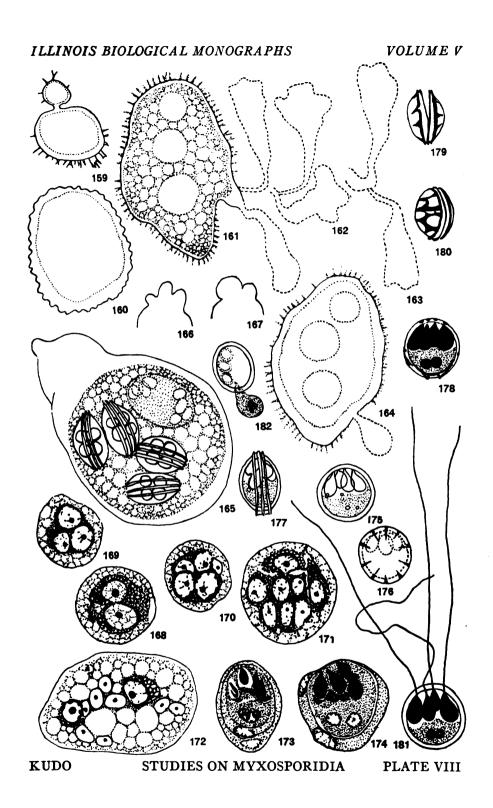


PLATE IX

Fig. 183 to 186. Sphaerospora divergens.

Fig. 183. Trophozoite. After Thélohan (1895, Fig. 12). ×750.

Figs. 184 and 185. Two views of spore. After Auerbach (1912, Pl. 5, Fig. 4). × about 1500.

Fig. 186. A spore treated with nitric acid. After Thélohan (1895, Fig. 13). ×1500.

Figs. 187 and 88. Spores of Sphaerospora elegans. After Thélohan (1890b, Fig. 1). X about 1000.

Fig. 189. A spore of Sphaerospora rostrata. After Thélohan (1895, Fig. 93). ×about 1635.

Figs. 190 to 192. Spores of Sphaerospora masovica. After Cohn (1902, Fig. 3). ×1000.

Fig. 192. Spore with extruded polar filaments and "starren Fäden" by warming.

Figs. 193 and 194. Spores of Sphaerospora platessae. After Woodcock (1904, Fig. 7d). ×900.

Figs. 195 to 197. Spores of Sphaerospora angulata. After Fujita (1912, Fig. 3). × about 2800.

Figs. 198 and 199. Spores of Sphaerospora polymorpha. After Davis (1917, Figs. 91 and 92) ×1500.

Figs. 200 to 204. Sphaerospora carassis. Original.

Fig. 200. A trophozoite. ×2250.

Figs. 201 to 203. Different views of spores. ×1800.

Fig. 204. A young spore. \times 2250.

Figs. 205 to 209. Sinuolinea dimorpha. After Davis (1916).

Fig. 205. A fresh trophozoite (1916, Fig. 1). ×1400.

Figs. 206 and 207. Trophozoites with erythrocytes in different stages of disintegration (1916, Figs. 2 and 57). ×640.

Fig. 208. A stained disporous trophozoite (1916, Fig. 41).

Fig. 209. A stained gemmule (1916, Fig. 72).

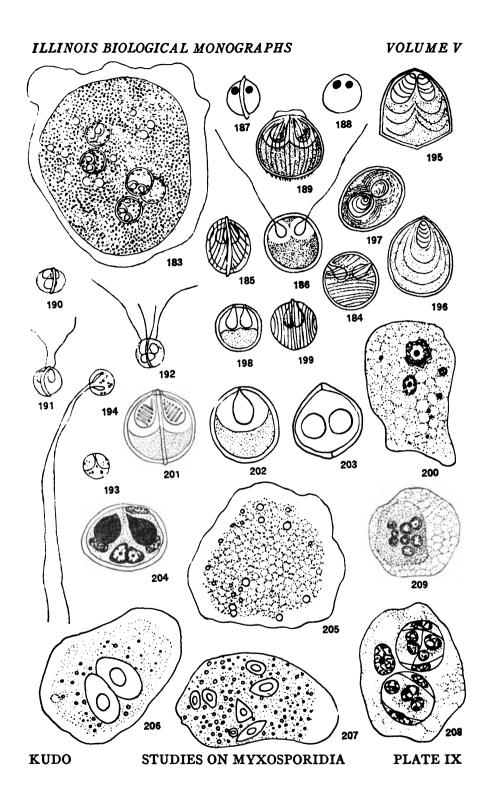


PLATE X

Figs. 210 to 213. Sinuclines dimorphs. After Davis (1916 and 1917).

Fig. 210. A living trophosoite (1916, Fig. 56). ×640.

Fig. 211. A living trophosoite from which a gemmule is just escaping (1916, Fig. 60). ×640.

Figs. 212 and 213. Spores (1917, Figs. 99 and 100). ×1400.

Figs. 214 to 216. Spores of Sinuolinea capsularis. After Davis (1917, Figs. 105 to 107). ×1500.

Figs. 217 and 218. Spores of Sinuclines arborescens. After Davis (1917, Figs. 109 to 110). ×1500.

Fig. 219. Spore of Sinuclinea opacita. After Davis (1917, Fig. 112). ×1500.

Fig. 220. Spore of Simulinea brachiophera. After Davis (1917, Fig. 113). X1500.

Figs. 221 to 224. Myzidium lieberkülmi. After Bütschli (1881 and 1882).

Fig. 221. A trophozoite (1882, Pl. 38, Fig. 12). × about 60.

Fig. 222. A trophozoite (1882, Pl. 38, Fig. 13). ×160.

Figs. 223 and 224. Trophosoites (1881, Figs. 27 and 26).



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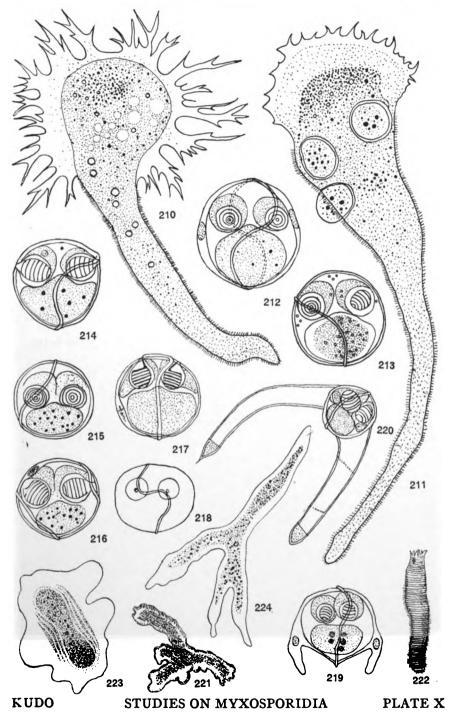


PLATE XI

Figs, 225 to 240. Myxidium lieberk#kni.

Fig. 225. A trophozoite. After Lieberkühn from Gurley (1894, Pl. 43, Fig. 1a). ×330.

Figs. 226 and 227. Trophozoites. After Bütschli (1881, Figs. 25 and 31).

Figs. 228 to 230. Stained trophozoites. After Thélohan (1895, Figs. 44, 46 and 45). ×750.

Fig. 231. A trophosoite forming daughter individuals by budding. After Cohn (1895 Fig. 5).

Fig. 232. Four figures showing the separation of a bud. After Cohn (1895, Figs. 6a, 6b, 6d and 6e).

Fig. 233. A cross-section of a trophozoite, showing the ectoplasm, mesoplasm and endoplasm. After Cohn (1895, Fig. 2).

Fig. 234. A trophozoite. After Laveran and Mesnil (1902, Fig. 3). ×500.

Fig. 235. A part of a trophozoite. After Laveran and Mesnil (1902, Fig. 3). ×800.

Figs. 236 and 237. Young trophosoites undergoing division. After Laveran and Mesnil (1902, Figs. 4 and 5). \times 1000.

Fig. 238. An isolated spore. After Bütschli (1881, Fig. 32). × about 2500.

Figs. 239 and 240. Fresh and stained spores. After Thélohan (1895, Figs. 47 and 48). $\times 1500.$

Figs. 241 to 251. Myxidium incurvatum.

Fig. 241. A monosporous trophozoite. After Parisi (1912, Fig. 1).

Fig. 242. A spore. After Thélohan (1895, Fig. 54). ×1500.

Figs. 243 and 244. Different views of spore. After Parisi (1912, Fig. 1).

Fig. 245. A young spore. After Georgévitch (1916, Fig. 11).
Figs. 246 to 248. Spores. After Georgévitch (1916, Figs. 10, 9 and 8).
Figs. 249 to 251. Spores. After Davis (1917, Figs. 119 to 121). ×1500.

Fig. 252. Trophozoite of Myzidium sphaericum. After Thélohan (1895, Fig. 28). ×1500.

Fig. 253. A spore of Myzidium histophilum. After Thélohan (1895, Fig. 49). ×1500.

Fig. 254. A spore of Myzidium sp. After Leydig from Gurley (1894, Pl. 47, Fig. 6).

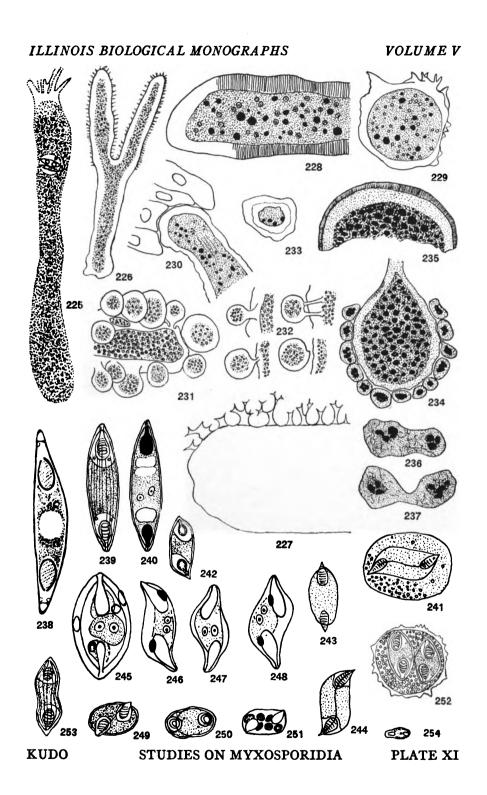


PLATE XII

Figs. 255 to 257. Myzidium danilewskyi. After Laveran (1898).

Figs. 255 and 256. Longitudinal and transverse sections thru renal tubules, showing the trophozoites (1898, Figs. 1 and 2). Fig. 255. ×350.

Fig. 257. Spores (1898, Figs. 4, 5 and 6). ×800.

Fig. 258. Spores of Myxidium giganteum. After Doflein (1898, Fig. 48). × about 500.

Figs. 259 to 261. Spores of *Myzidium giardi*. After Cépède (1906a, Figs. 1, 3 and 2). ×2000.

Figs. 262 to 265. Spores of Myxidium pfeifferi. After Auerbach (1908, Figs. 6 and 7). × about 2000, except Fig. 265.

Fig. 266. A spore of Myxidium inflatum. After Auerbach (1909, Fig. 3a). × about 1500.

Fig. 267. A spore of Myzidium bergense. After Auerbach (1910a, Fig. 57). × about 1820.

Fig. 268. A spore of Myxidium procerum. After Auerbach (1910a, Fig. 58). × about 2000. Figs. 269 to 271. Spores of Myxidium mackiei. After Bosanquet (1910, Fig. 12). ×

Figs. 269 to 271. Spores of Myxidium mackiei. After Bosanquet (1910, Fig. 12). × about 1250.

Figs. 272 and 273. Spores of Myxidium macrocapsulare. After Auerbach (1910d, Figs. 1a and 1b). ×3000.

Figs. 274 to 276. Myxidium sp. After Awerinzew (1908 and 1911).

Fig. 274. A monosporous trophozoite (1911, Fig. C).

Fig. 275. A disprous trophozoite (1908, Pl. 2, Fig. 6). Obj. E and oc. 4.

Fig. 276. A spore (1908, Pl. 1a, Fig. 17). × about 1000.

Figs. 277 and 278. Spores of Myxidium depressum. After Parisi (1912, Figs. 2a and 2b). × about 1600.

Figs. 279 and 280. Spores of Myxidium oviforme. After Parisi (1912, Fig. 3). × about 1600.

Figs. 281 to 284. Spores of Myxidium anguillae. After Ishii (1915, Fig. 3a). ×1450.

Figs. 285 and 286. Myxidium sp. After Mavor (1915).

Fig. 285. A spore treated with ammonia water (1915, Fig. 3a). ×660.

Fig. 286. A spore (1915, Fig. 3b). ×1320.

Figs. 287 to 290. Spores of Myzidium gadi. After Georgévitch (1916, Figs. 1, 4, 3 and 2).

Fig. 228. A spore from Solea vulgaris.

Figs. 289 and 290. Young spores.

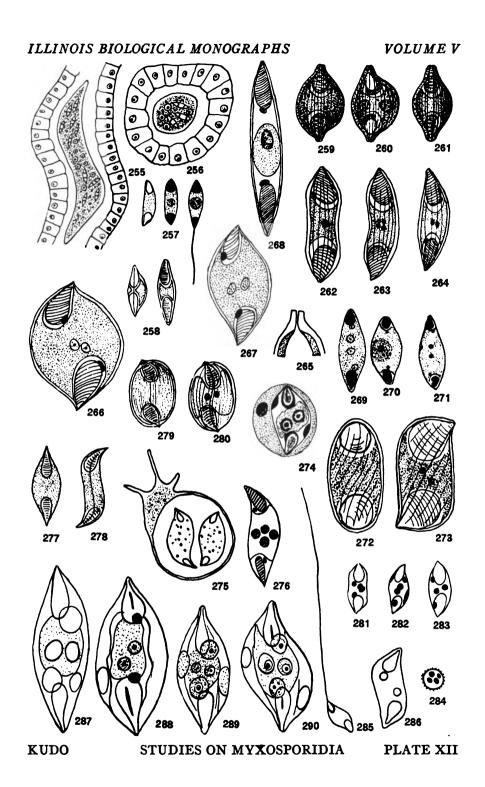


PLATE XIII

Fig. 291. A spore of Myzidium glutinosum. After Davis (1917, Fig. 124). ×1400.

Figs. 292 and 293. Spores of *Myzidium phyllium*. After Davis (1917, Figs. 126 and 127). ×2000.

Figs. 294 and 295. Spores of Myxidium kagayamai. After Kudo (1916, Fig. 2). ×1750 and ×1000 respectively.

Figs. 296 to 307. Sphaeromyza balbianii.

Fig. 296. A trophozoite. After Thélohan (1895, Fig. 57). X3.

Fig. 297. A trophozoite. After Davis (1917, Fig. 128). ×640.

Fig. 298. A trophozoite in plasmotomy. After Georgévitch (1916, Fig. 15).

Figs. 299 and 300. Spores. After Thélohan (1895, Figs. 58 and 59). ×1500.

Fig. 301. An end of a spore. After Thélohan (1895, Fig. 60).

Fig. 302. A spore treated with nitric acid. After Thelohan (1895, Fig. 61).

Fig. 303. A spore. After Parisi (1912, Fig. 4). × about 1750.

Fig. 304. A spore. After Davis (1917, Fig. 130). ×2100.

Figs. 305 to 307. Mature and young spores (Figs. 306 and 307). After Georgévitch (1916, Figs. 17, 20 and 19).

Figs. 308 to 311. Sphaeromyza immersa. After Lutz (1889: 301).

Fig. 308. An infected gall-bladder of Bufo aqua (1889, Fig. 1). X1.

Fig. 309. Spores (1889, Figs. 4, 5 and 6).

Fig. 310. A spore (1889, Fig. 10). ×600.

Fig. 311. A spore with extruded polar filaments (1889, Fig. 7).

Figs. 312 to 314. Spores of Sphaeromyza incurvata. After Doflein (1898, Fig. 49). × about 1000.

Figs. 315 and 316. Sphaeromyza sabrasesi. After Schröder (1907).

Fig. 315. A trophozoite (1907, Fig. 1). ×15.

Fig. 316. A cross section thru a trophozoite (1907, Fig. 3). ×1500.

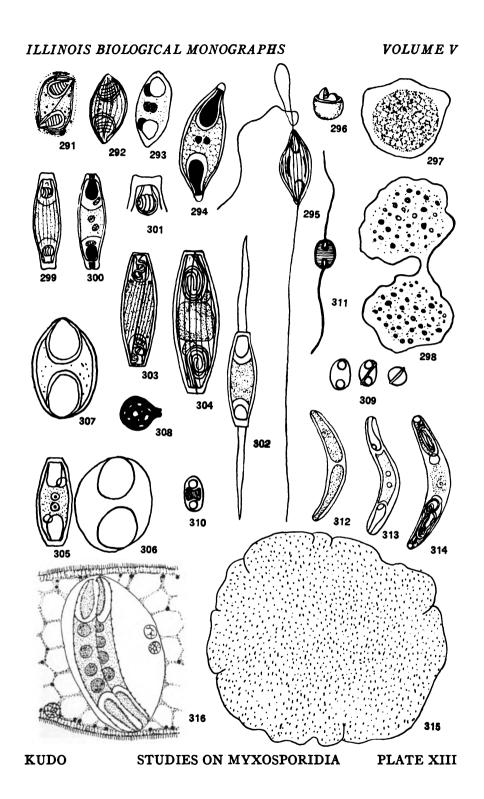


PLATE XIV

Figs. 317 to 322. Spores of Sphaeromyza sabrasesi.

Figs. 317 to 319. After Laveran and Mesnil (1900, Figs. 1, 3 and 4). ×1500.

Fig. 318. A spore treated with nitric acid (1900, Fig. 3).

Figs. 320 and 321. Spores. After Schröder (1907, Figs. 39 and 41). ×1500.

Fig. 322. A polar capsule. After Schröder (1907, Fig. 45).

Figs. 323 and 324. Spores of Sphaeromyza hellandi. After Auerbach (1909, Fig. 5). X about 1500.

Figs. 325 and 326. Spores of Sphaeromyza exneri. After Awerinzew (1913a, Fig. 3). X about 365.

Fig. 327. A spore of Sphaeromyza gasterostei. After Georgévitch (1916, Fig. 22).

Figs. 328 to 331. Zschokkella hildae. After Auerbach (1910a and 1912).

Fig. 328. A monosporous trophozoite (1912, Pl. 5, Fig. 2).

Figs. 329 to 331. Spores (1910a, Fig. 62).

Figs. 332 and 333. Spores of Zschokkella nova. After Klokacewa (1914, Fig. 2). X about 2500.

Figs. 334 to 338. Spores of Zschokkella acheilognathi. After Kudo (1916).

Figs. 334 to 336. Different views of fresh spores (1916, Figs. 3d, 3e and 3f). ×2250.

Fig. 337. A young spore. Original. ×2785.

Fig. 338. A stained spore (1916, Fig. 3h). ×2800.

Figs. 339 and 340. Spores of Zschokkella globulosa. After Davis (1917, Figs. 135 and 134). ×1500.

Figs. 341 to 343. Spores of *Myxasoma dujardini*. After Thélohan (1895, Figs. 90, 91, and 89). ×1500.

Figs. 344 to 347. Spores of Myzosoma funduli. After Kudo (1918a, Fig. A). ×1500.

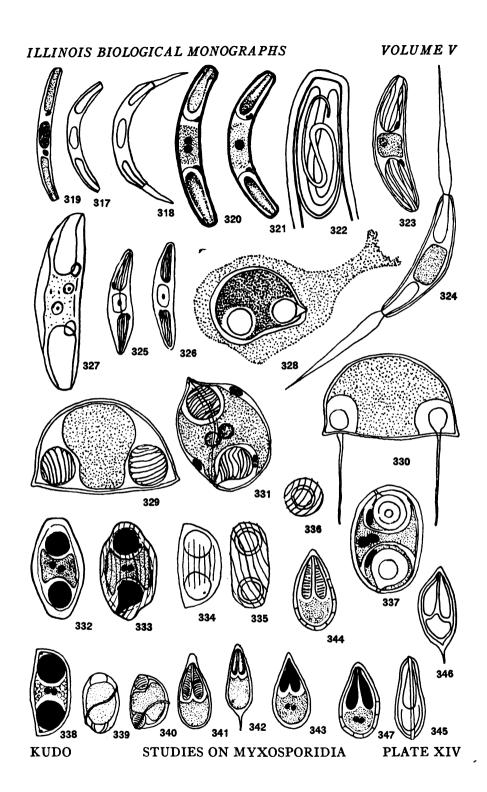


PLATE XV

- Fig. 348. A spore of Myzosoma(?) lobatum. After Nemeczek (1911, Fig. 18). ×1000.
- Fig. 349 to 354. Lentospora cerebralis. After Plehn (1905). ×1200.
- Fig. 349. Various young ameboid forms; stained (1905, Textfig. 5).
- Fig. 350. A stained larger form (1905, Textfig. 5).
- Fig. 351. A trophozoite with two spores (1905, Textfig. 4).
- Fig. 352. Various spores (1905, Textfig. 2).
- Fig. 353. A stained spore (1905, Textfig. 3).
- Fig. 354. A spore with extruded polar filaments (1905, Textfig. 2).
- Fig. 355. A spore of Lentospora multiplicata. After Reuss (1906, Fig. 8). ×1500.
- Figs. 356 to 359. Spores of Lentospora asymmetrica. After Parisi (1912, Fig. 7). × about 1500.
- Figs. 360 to 362. Spores of Leniospora acuta. After Fujita (1912, Fig. 2). ×2000.
- Figs. 363 and 364. Spores of Myzobelus piriformis. After Thelohan (1895, Figs. 117 and 116). ×1500.
- Figs. 365 and 366. Spores of Myzobolus unicapsulatus. After Müller (1841, Fig. 5).
- Fig. 367. A spore of Myzobolus fuhrmanni. After Auerbach (1909, Fig. 1b). ×1840.
- Fig. 368. A spore of Myzobolus oculi-leucisci. After Trojan (1909, Fig. 3). ×2000.
- Figs. 369 and 370. Spores of Myzobolus toyamai. After Kudo (Original and 1917, Fig. 40). ×2500.
- Figs. 371 and 372. Spores of Myzobolus notatus. After Mavor (1916, Figs. 6C and 6B). ×2600.
- Figs. 373 and 374. Spores of Myzobolus robitae. After Southwell and Prashad (1918, Figs. 26 and 27). × about 1720 and 700 respectively.
- Figs. 375 and 376. Spores of Myxobolus seni. After Southwell and Prashad (1918, Figs. 29 and 30). × about 1700.
- Figs. 377 and 378. Spores of Myzobolus misgumi. Original. ×1500.

ILLINOIS BIOLOGICAL MONOGRAPHS

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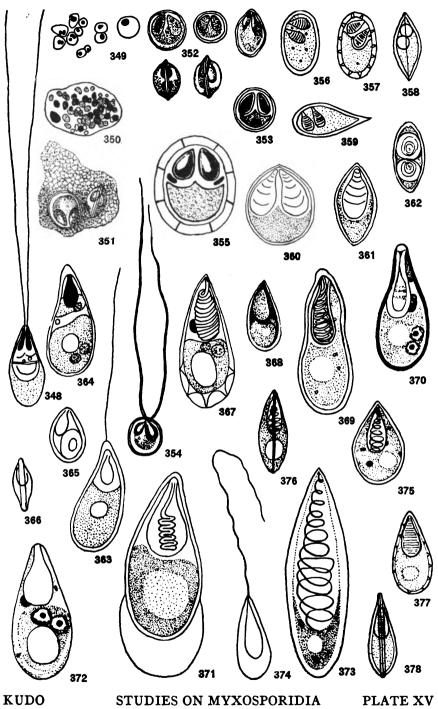


PLATE XVI

Figs. 379 to 385. Myxobolus pfeifferi.

Figs. 379 and 380. Parts of section thru cyst. After Keysselitz (1908a, Pl. 15, Figs. 1 and 2).

Fig. 381. A spore. After Thélohan (1895, Fig. 77). ×1500.

Fig. 382. An optical section of spore. After Keysselitz (1908a, Fig. A).

Fig. 383. A spore treated with Lugol's solution. After Keysselitz (1908a, Pl. 14: Fig. 92).

Fig. 384. A spore with extruded filaments. After Keysselitz (1908a, Fig. C).

Fig. 385. A stained young spore. After Keysselitz (1908a, Pl. 14, Fig. 81).

Fig. 386. A spore of Myzobolus dispar. After Thélohan (1895, Fig. 86). X about 1500.

Figs. 387 to 389. Spores of Myxobolus ellipsoides. Fig. 389. An abnormal spore with six polar capsules. After Thélohan (1895, Figs. 112, 113 and 115).

Fig. 390. A part of the infected intestine of Mugil auralus, showing cysts of Myzobolus exiguus. After Parisi (1912, Fig. 9e). × about 3.

Fig. 391. A spore of Myzobolus exiguus. After Thélohan (1895, Fig. 98). X 1500.

Figs. 392 to 395. Spores of Myxobolus exiguus. After Parisi (1912, Fig. 9). ×2500.

Fig. 396. A spore of Myzobolus oviformis. After Thelphan (1895, Fig. 81). ×1500.

Figs. 397 to 398. Spores of Myxobolus mulleri. After Thélohan (1895, Figs. 96 and 97). ×1500.

Figs. 399 and 400. Spores of Myxobolus mulleri. After Bütschli (1881, Figs. 1 and 2).

Fig. 401. A spore of Myzobolus mulleri in conc. sulphuric acid. After Bütschli (1881, Fig. 6).

Figs. 402 and 403. Abnormal spores of Myzobolus mülleri. After Bütschli (1881, Figs. 9 and 8).

Figs. 404 and 405. Spores of Myzobolus lintoni. After Linton (1891, Figs. 3 and 5).

Fig. 406. Diagram of the cross section of a spore of Myxobolus lintoni. After Linton (1891, Fig. 8).

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Fig. 408. A stained spore of Myzobolus lintoni. After Gurley (1894, Pl. 26, Fig. 7). × about 2000.

Figs. 409 and 410. Spores of Myzobolus globosus. After Gurley (1894, Pl. 26, Fig. 7). * about 2900.

Fig. 411. Spores of Myzobolus inaequalis. After Müller (1841, Fig. 6).

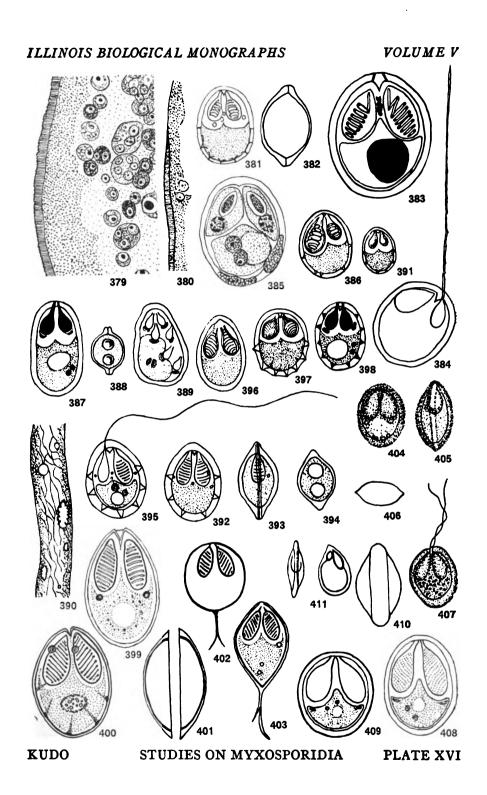


PLATE XVII

Figs. 412 to 416. Spores of Myzobolus oblongus. Figs. 412 to 414. After Gurley (1894, Pl. 26, Fig. 6). ×2300. Figs. 415 and 416. After Müller (1841, Fig. 9).

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Figs. 438 and 439. Spores (1906a, Figs. 5a and 3d). ×about 1320.

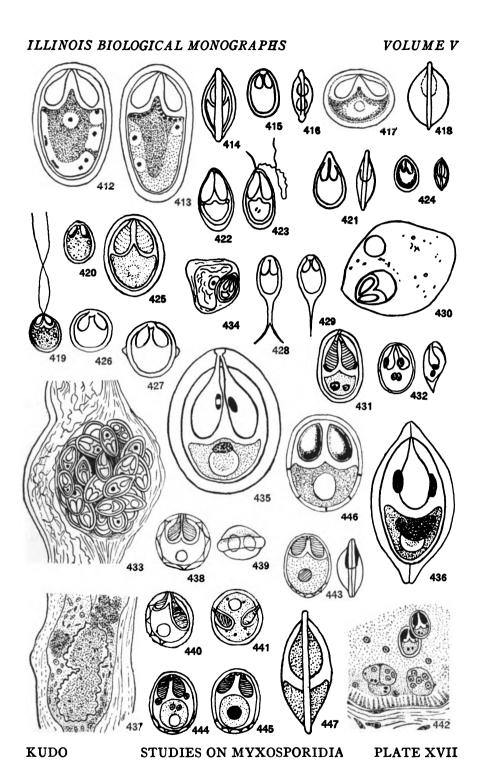
Figs. 440 and 441. Abnormal spores (1906a, Figs. 5b and 5c). Xabout 1320.

Figs. 442 to 445. Myzobolus gigas. After Auerbach (1906b).

Fig. 442. A part of the section of a cyst (1906b, Fig. 1).

Figs. 443 to 445. Spores (1906b, Figs. 3a, 3c, 5 and 3b). × about 850.

Figs. 446 and 447. Spores of Myzobolus volgensis. After Reuss (1906, Fig. 1). ×2000.



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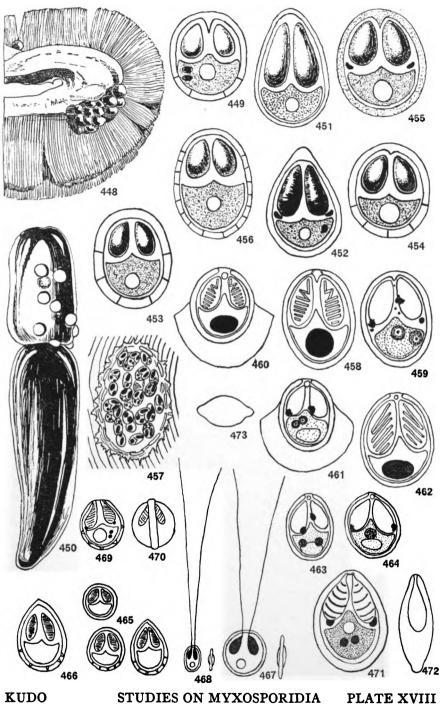
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PLATE XVIII

- Fig. 448. The branchia of Lucioperca volgonsis with cysts of Myzobolus volgonsis. After Reuns (1906, Fig. 2). ×2.25.
- Fig. 449. A spore of Myzobolus scardinii. After Reuse (1906, Fig. 3). ×1500.
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- Fig. 465. Spores of Myzobolus sp. After Wegener (1910, Fig. 44). ×1050.
- Fig. 466. A spore of Myzobolus parmagnus. After Wegener (1910, Fig. 45). ×1050.
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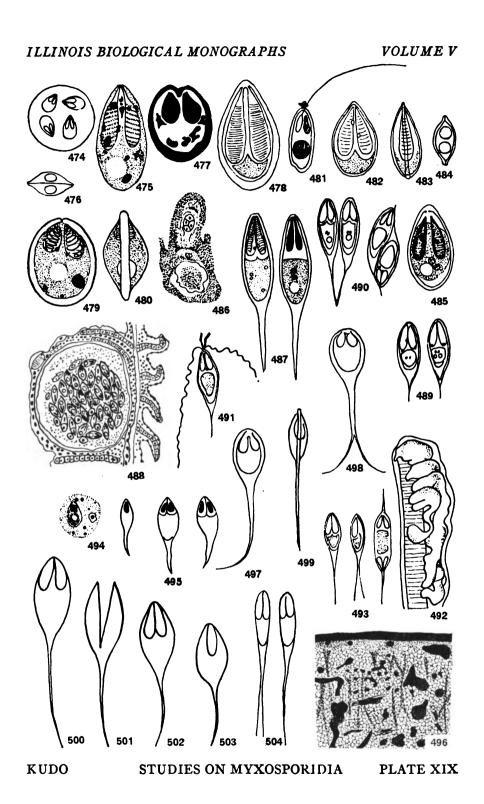


PLATE XX



Fig. 505. Spores of Henneguya Gurleyi. After Gurley (1894, Pl. 33, Figs. 8c, 6 and 7). × about 3100.

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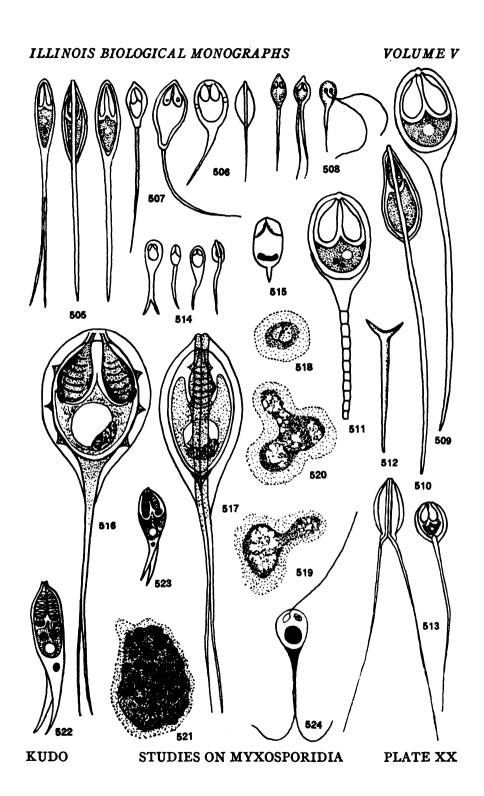


PLATE XXI

Figs. 525 and 526. Spores of *Henneguya acerinae*. After Schröder (1906, Figs. 5 and 6). ×1650.

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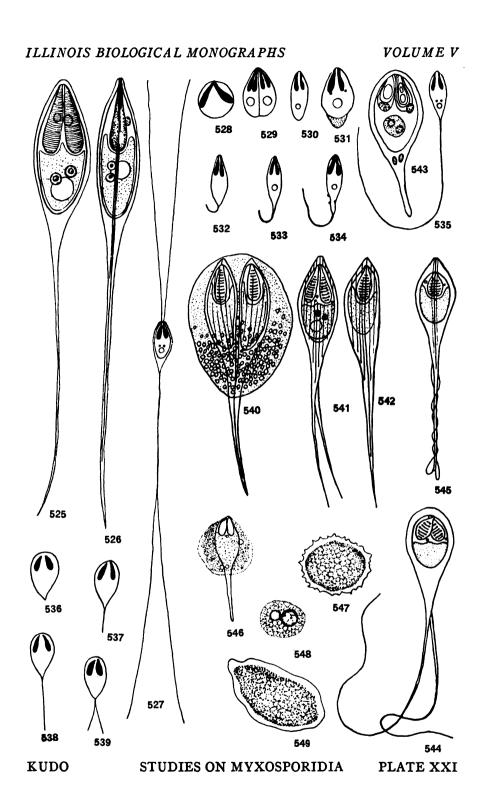


PLATE XXII



Figs. 550 to 557. Henneguya mictospora. Original.

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Figs. 567 and 568. Abnormal spores. ×1500.

Fig. 571. A spore stained with Lugol's solution. ×1500.

Fig. 572. A spore stained with Giemsa's mixture. ×2360.

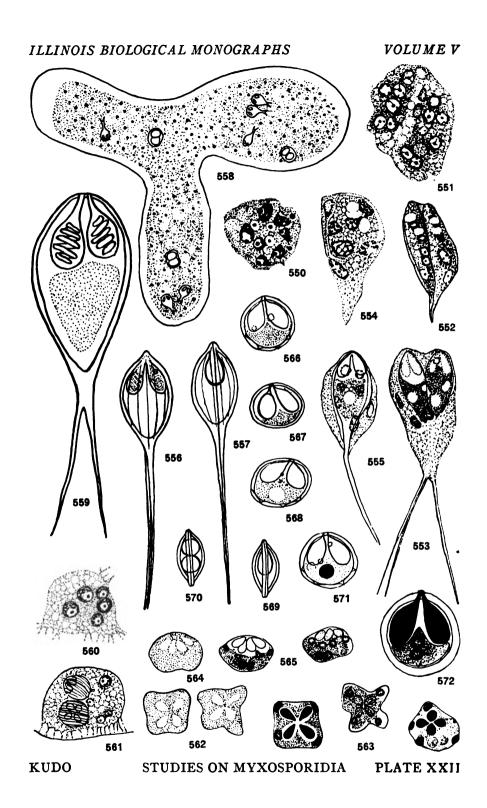


PLATE XXIII

Figs. 573 to 576. Myzobolus orbiculatus. Original.

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Fig. 586. A trophozoite with attached Ceratomyza acadiensis (1915, Fig. 8). ×830.

Fig. 587. A trophozoite (1915, Fig. 6).

Figs. 588 and 589. Trophozoites of genus et species incertae. After Mavor (1916a, Figs. 3d and 3b). ×660.

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Figs. 598 to 601. Unstained preserved spores, showing different views. × about 1500.

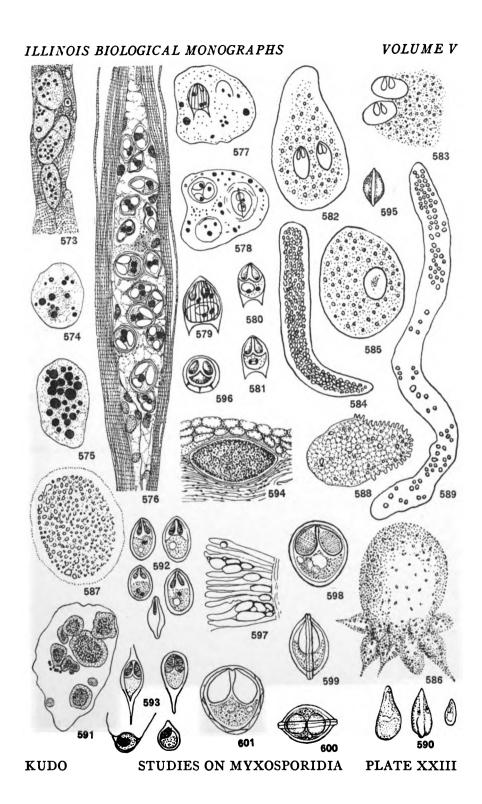


PLATE XXIV.

- Figs. 602 to 621. Mitraspora elongata. Original. ×1500, except Figs. 620 and 621, ×2500. Giemsa staining, unless otherwise stated.
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- Fig. 630. A Giemsa stained spore.
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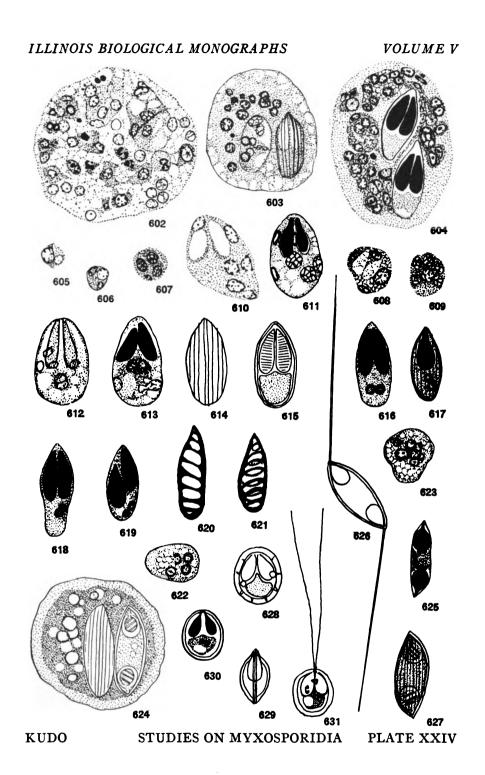


PLATE XXV

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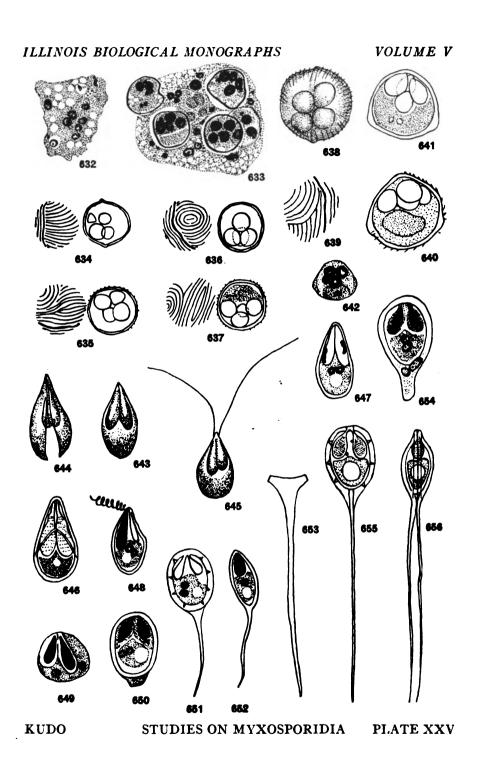
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